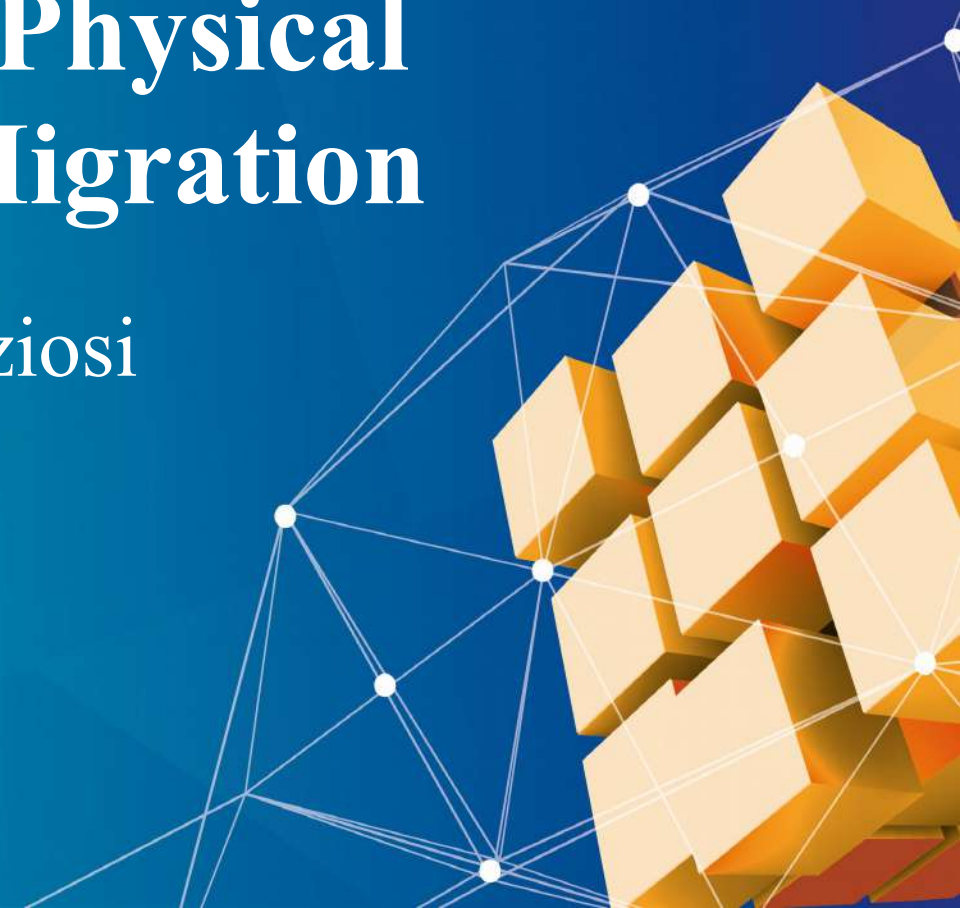


Modelling Physical Limits of Migration

Luigi Preziosi

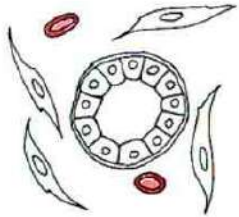


POLITECNICO
DI TORINO

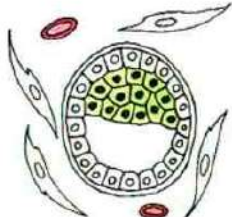


Compartmentalization or invasion

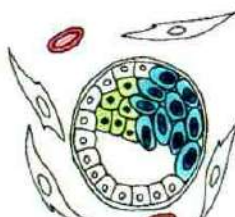
Normal duct



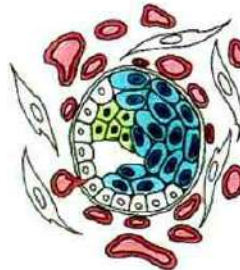
Hyperplasia



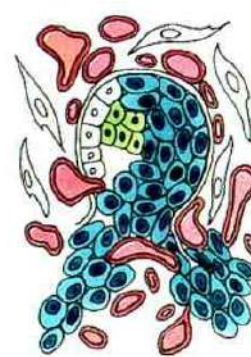
Dysplasia/ CIS



Angiogenic CIS



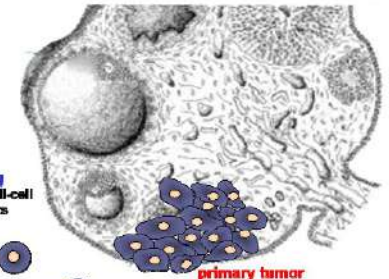
Invasive carcinoma



Breast

Ovary

Ovarian cancer dissemination



1) **Surface shedding**
Initial disruption of cell-cell and cell-matrix contacts

3) **Retraction, sub-mesothelial adhesion**
disruption of cell-cell contacts in multi-cellular aggregates

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migration/invasion through mesothelial layer and into sub-mesothelial EC matrix

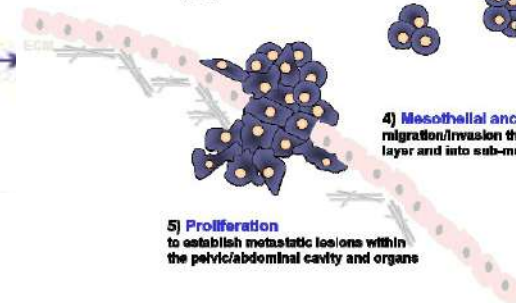
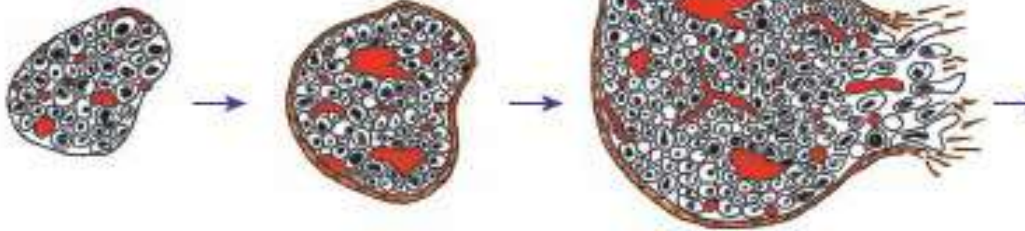
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Pancreas

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Small Tumor

Large Tumor/
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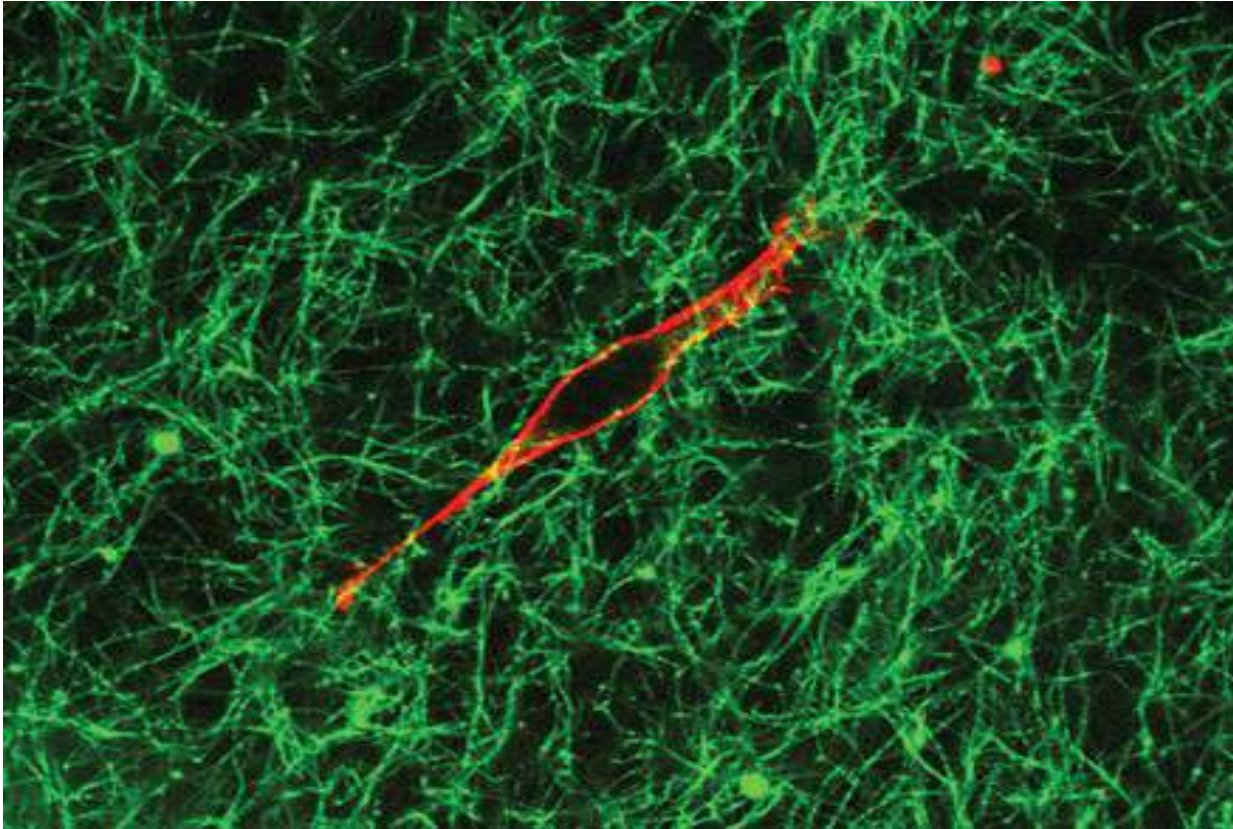




The carpenter syndrome

**To a carpenter with a hammer
the entire world is a nail**

Physical limit of migration



(P. Friedl, K. Wolf)

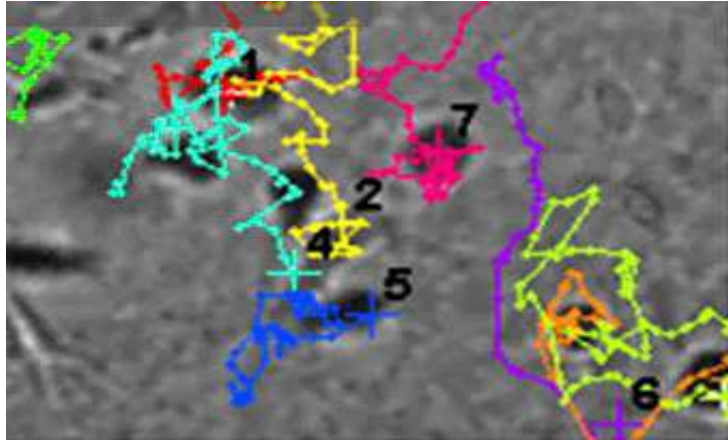


HT1080 migration
in rat tail collagen (1.7 mg/ml)
in presence of MMP inhibitor



Neutrophil migration
in rat tail collagen (1.7 mg/ml)
in presence of IL-8

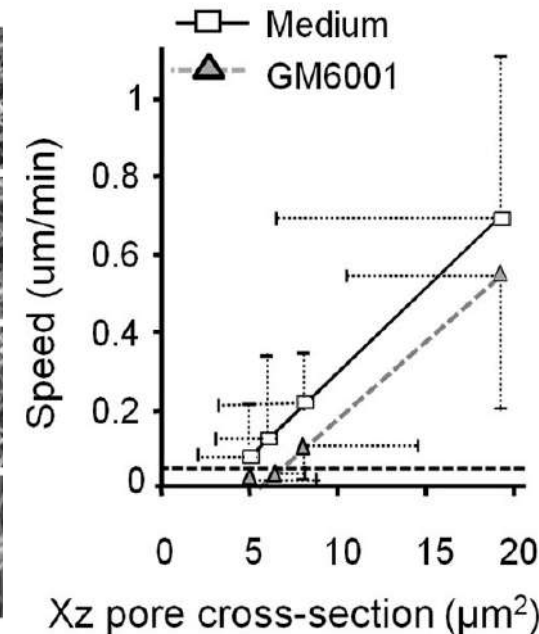
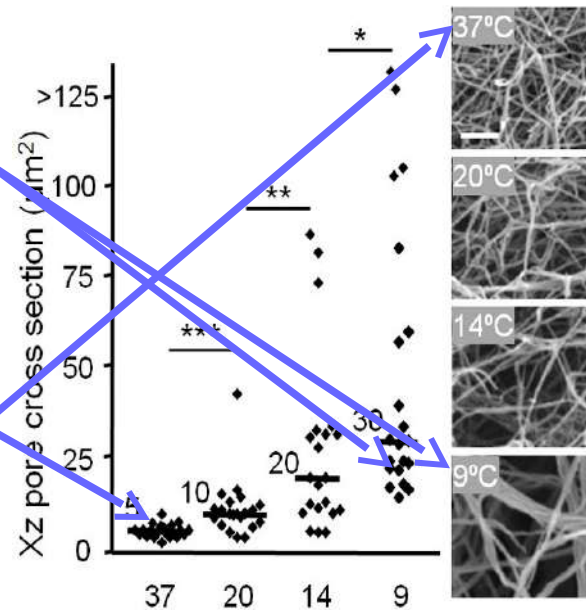
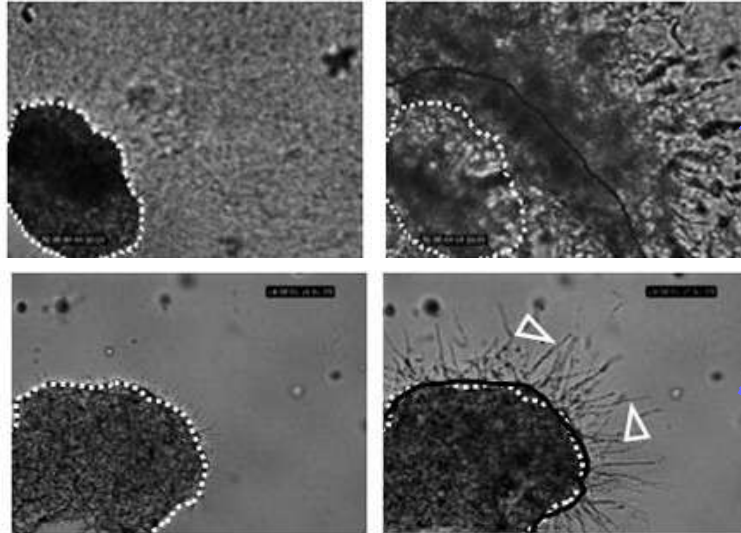
Physical limit of migration



Wolf, te Lindert, Krause, Alexander, te Riet, Wills, Hoffman, Figdor, Weiss, Friedl
J. Cell Biol. **201**, 1069-1084 (2013)

0h

18h



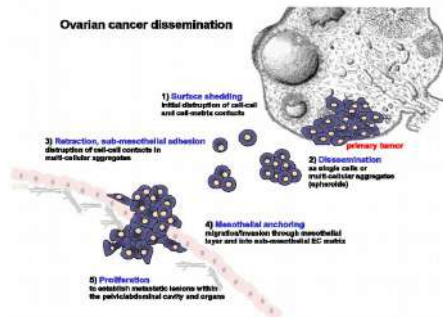
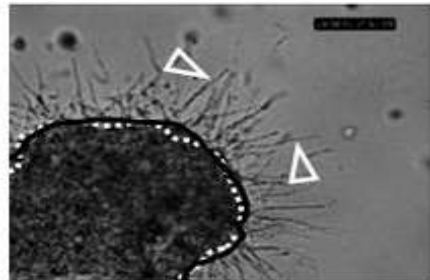
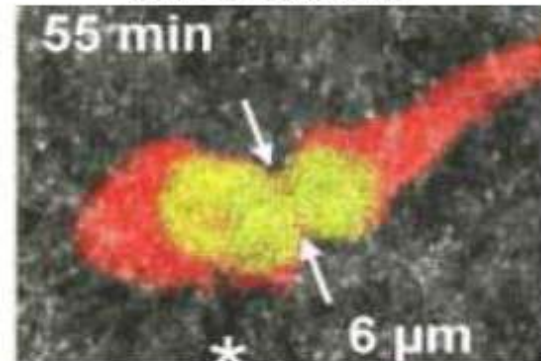
Plan of the talk

- Cell mechanics

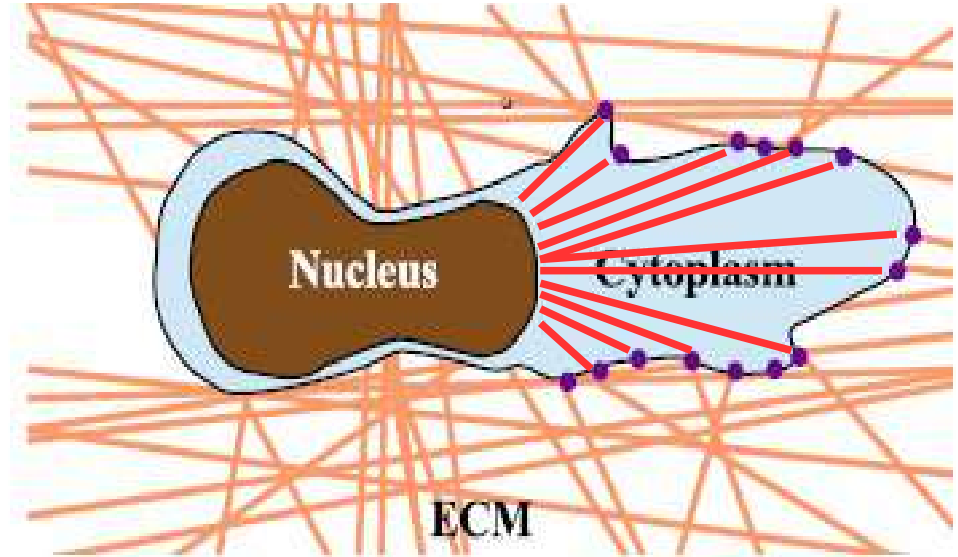
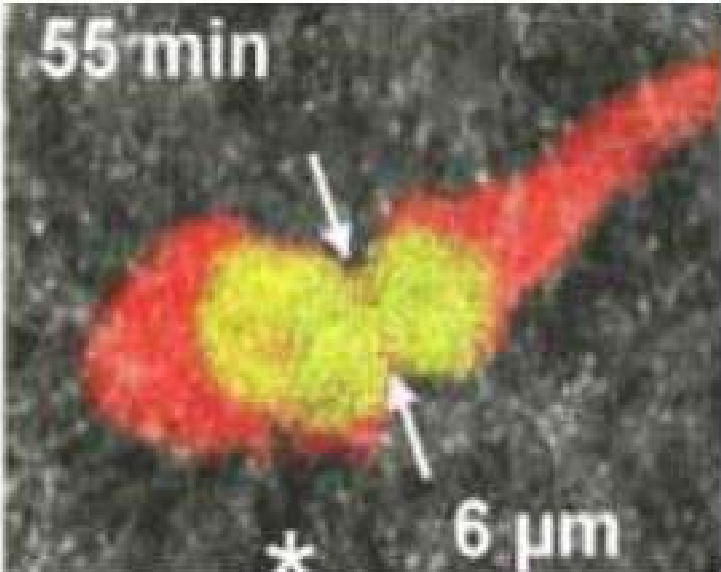
- Individual cell-based model

- Multiphase model

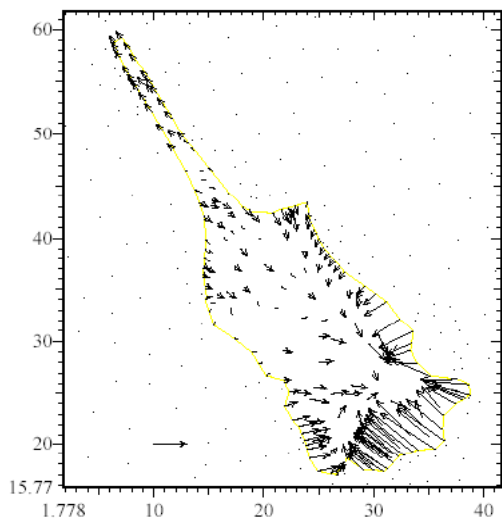
- Asymptotic limit for interface condition



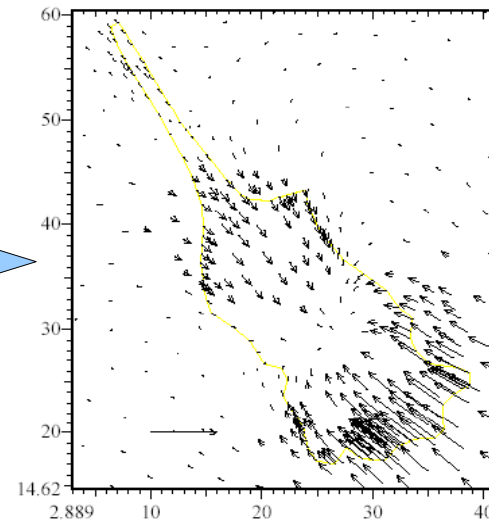
Cell pulling and nucleus squeezing



Traction force microscopy

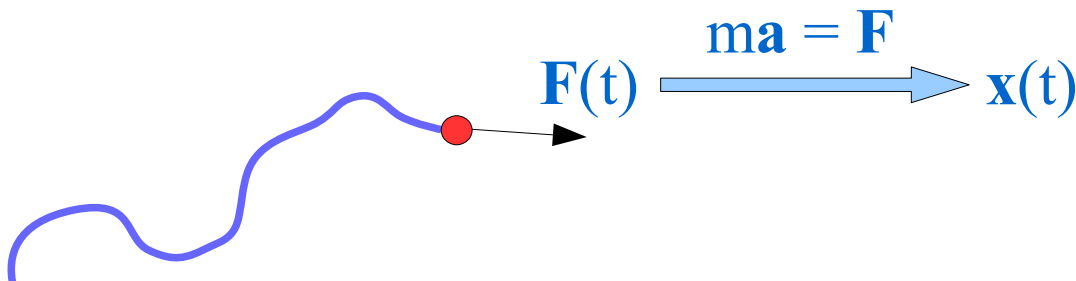


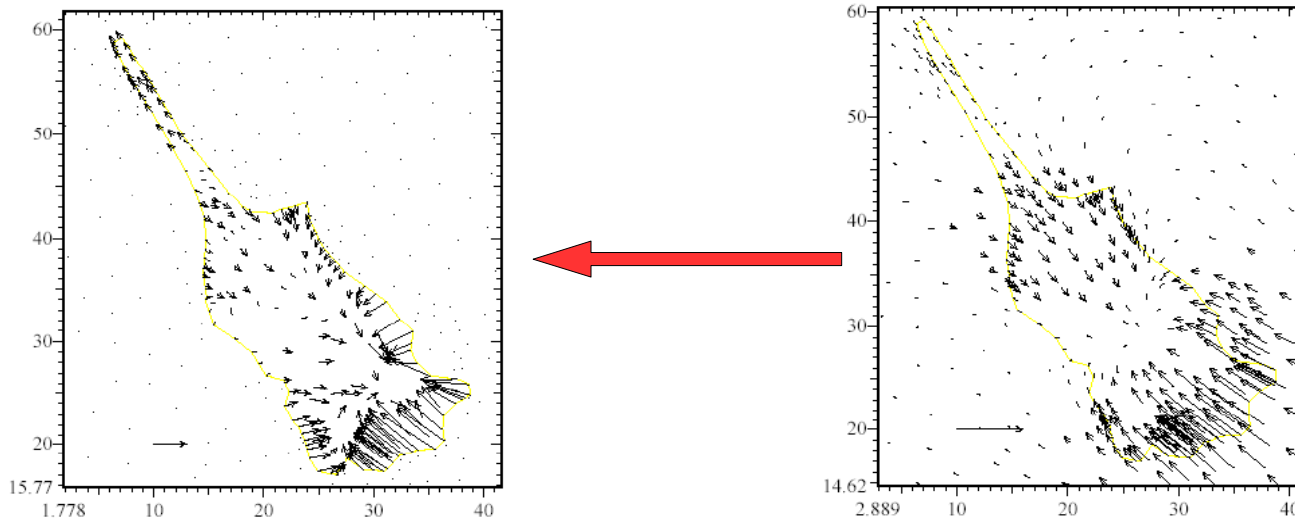
Direct
problem



Cell traction determines ECM deformation

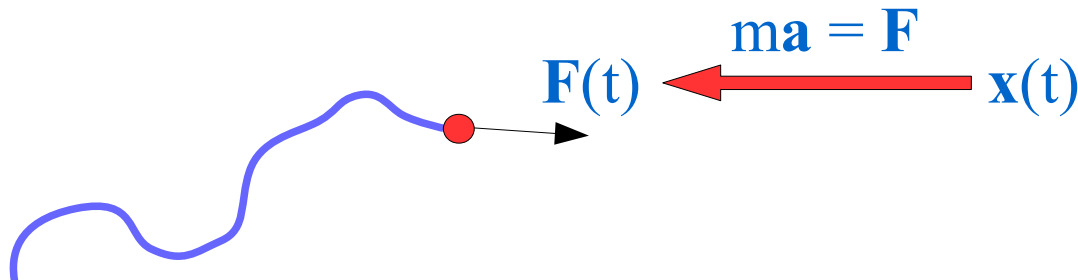
Given the force, determine the motion



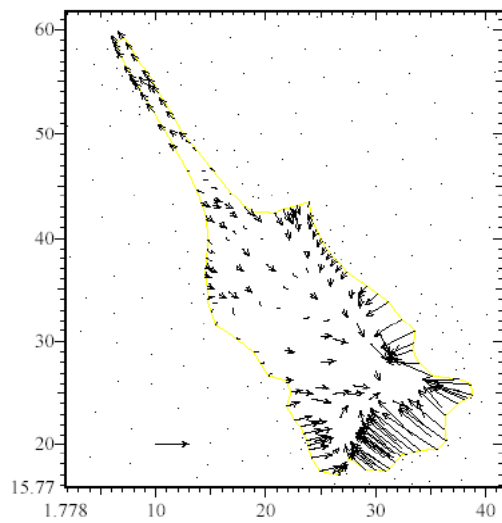


Deformation is determined by cell traction

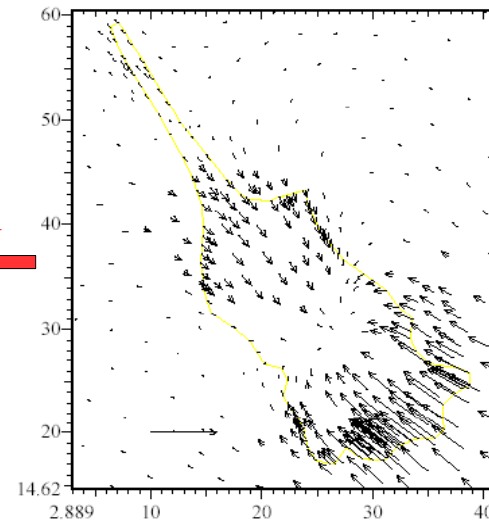
Given the motion determine the force



Traction force microscopy

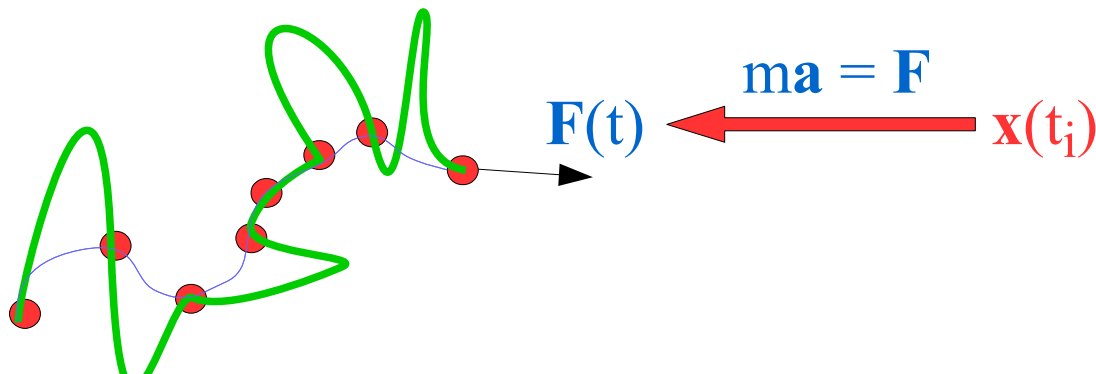


Inverse problem

Deformation is determined by cell traction

Given some information on the motion, determine the most plausible force



Given the deformations \mathbf{u}_{0i} in \mathbf{x}_i

For any admissible force \mathbf{F}
exerted in Ω_c

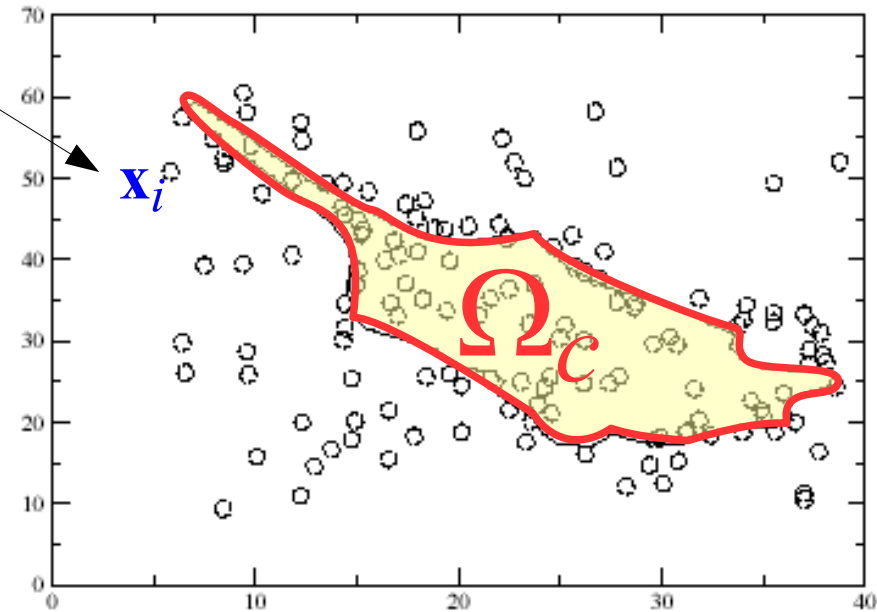
Compute $\mathbf{u}(\mathbf{x})$ such that

$$\mathcal{F}(\mathbf{u}) = \mathbf{F}$$

Compute the difference between
 $\mathbf{u}(\mathbf{x}_i)$ and \mathbf{u}_{0i}

Among all admissible \mathbf{F} choose the most “economical”

$$\mathcal{J}(\mathbf{F}) = \sum_i \left| [\mathcal{F}^{-1}(\mathbf{F})](\mathbf{x}_i) - \mathbf{u}_{0i} \right|^2 + \varepsilon \|\mathbf{F}\|^2$$



Linear elasticity
operators

$$-\hat{\mu}\Delta\mathbf{u} - (\hat{\mu} + \hat{\lambda})\nabla(\nabla \cdot \mathbf{u})$$

$$-\hat{\mu}\Delta\mathbf{p} - (\hat{\mu} + \hat{\lambda})\nabla(\nabla \cdot \mathbf{p})$$

Indicator function of Ω_c



$$-\frac{\chi_c}{\varepsilon}\mathbf{p},$$

$$\mathbf{u}|_{\partial\Omega} = 0,$$

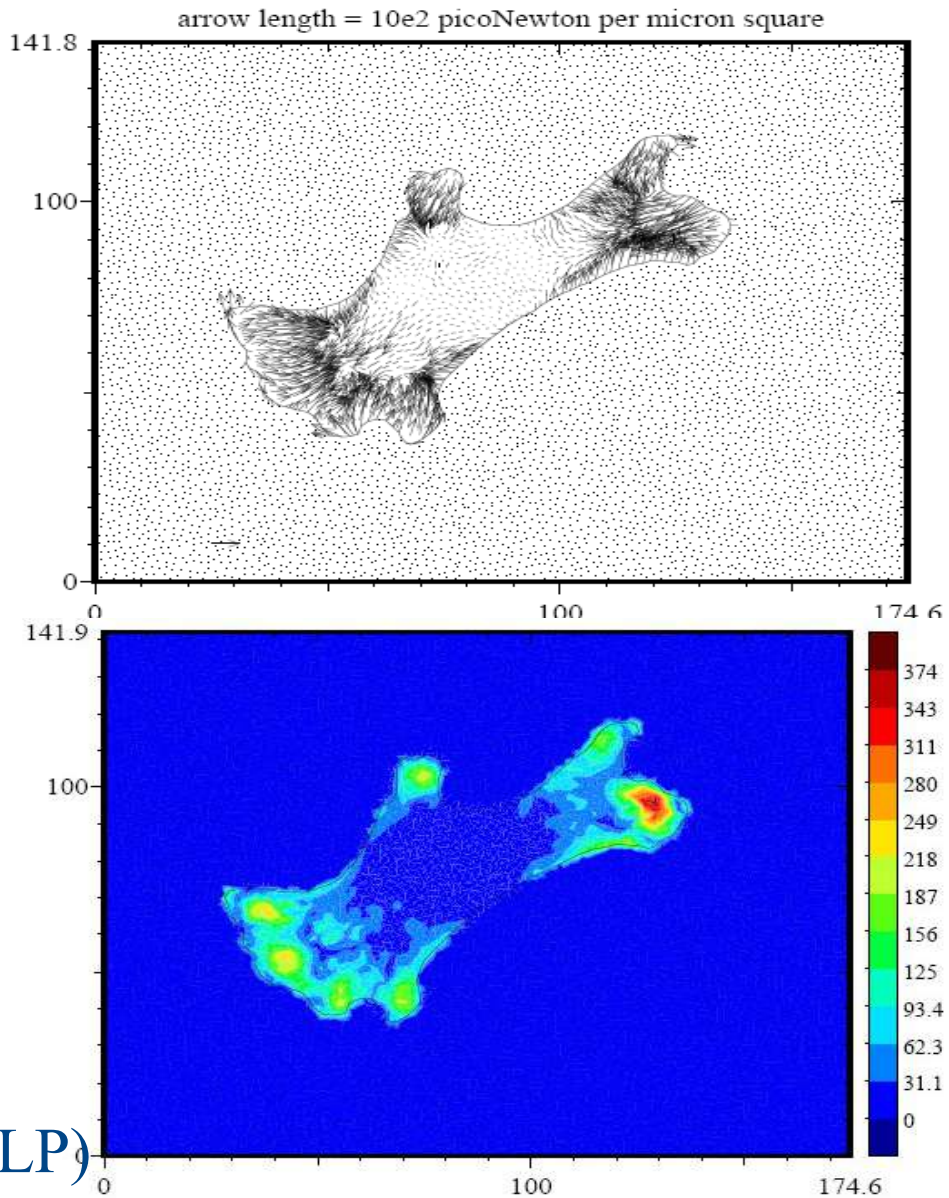
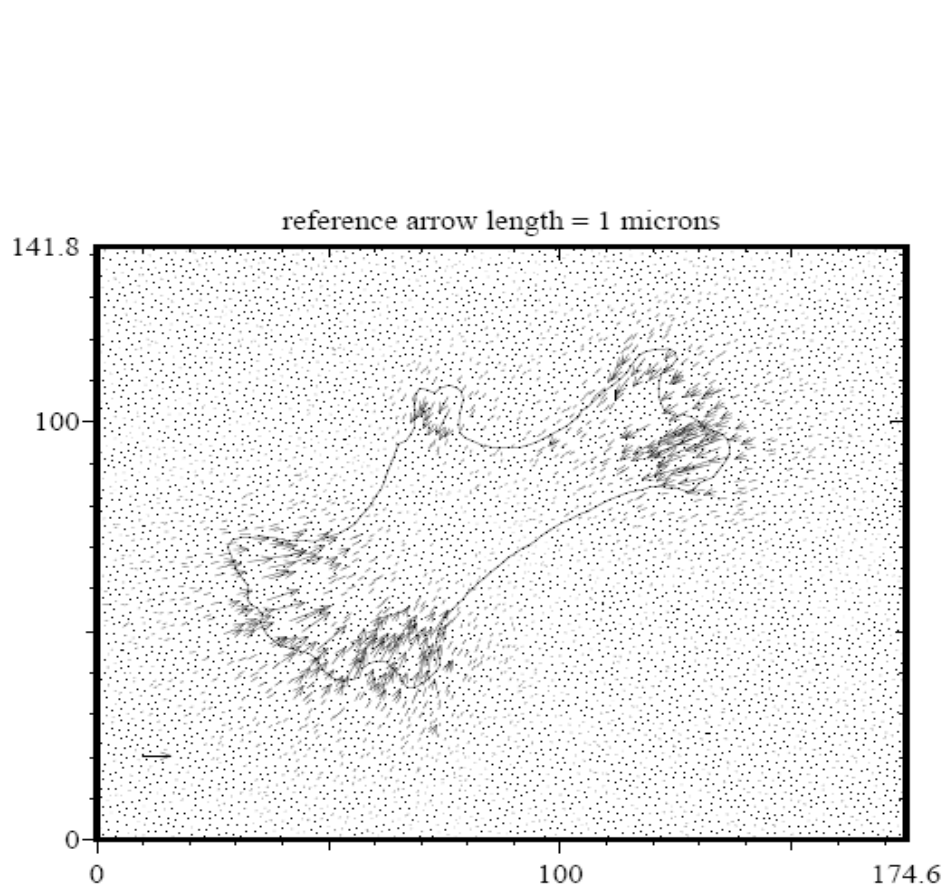


Indicator function of Ω_0

$$= \chi_o\mathbf{u} - \mathbf{u}_0, \quad \mathbf{p}|_{\partial\Omega} = 0.$$

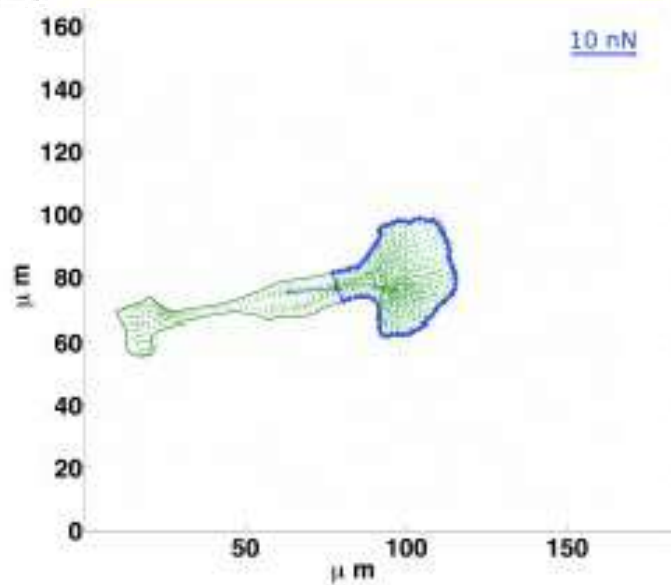
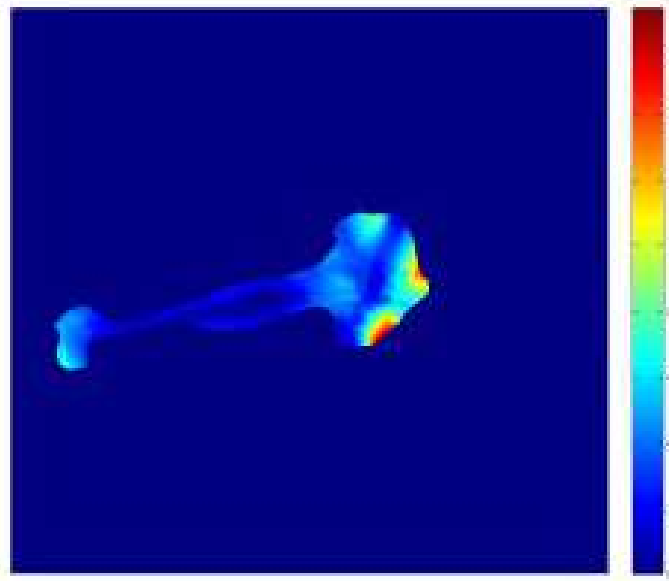
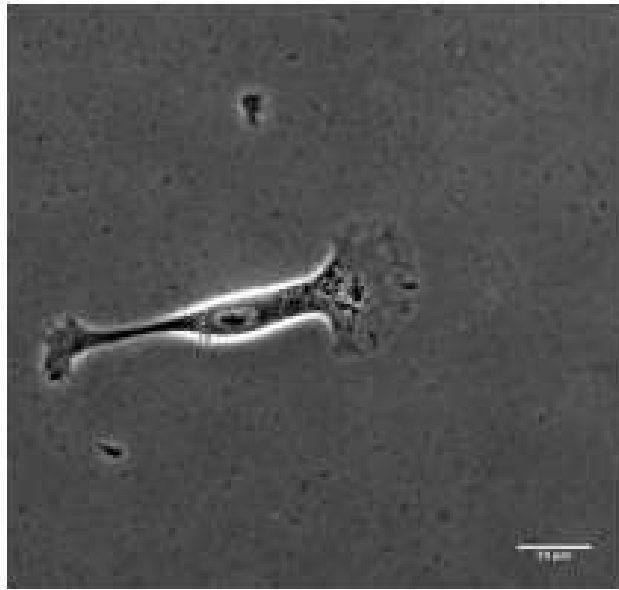
Self-adjoint problem

Traction force microscopy



(Ambrosi, Vitale, Verdier, Etienne, LP)

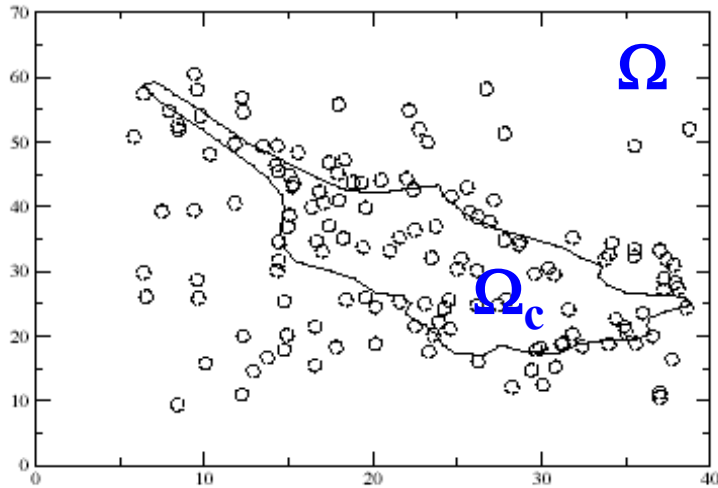
Traction force microscopy



Traction force microscopy in 3D

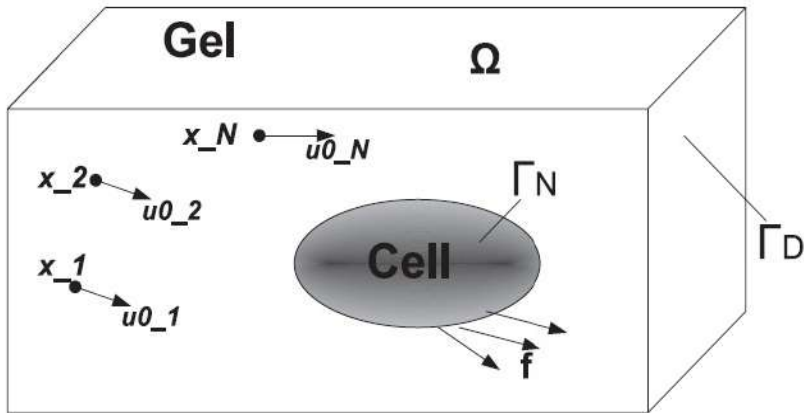
2D

- Measures in Ω (also below the cell)
- Forces in Ω_c (only under the cell)

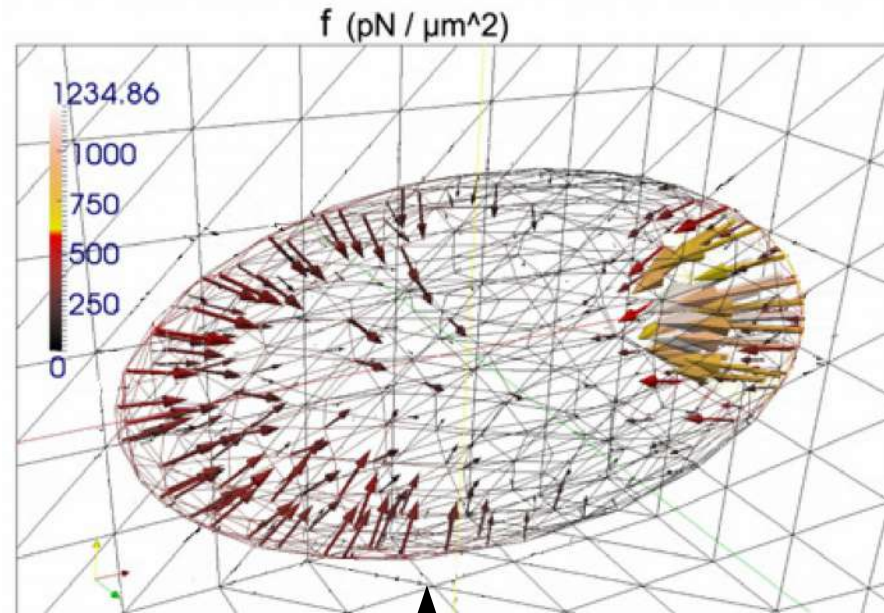
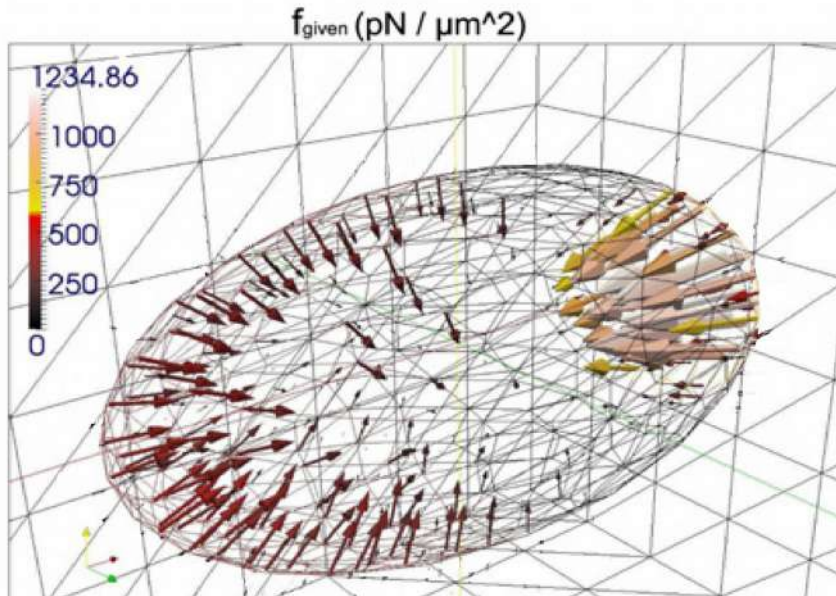


3D

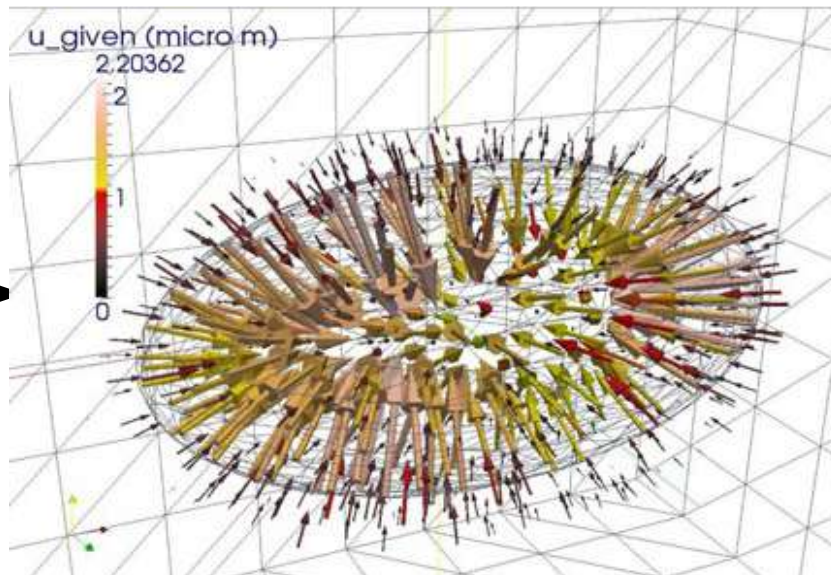
- Measures only outside Ω_c
- Forces on Γ_N (the cell membrane)



Traction force microscopy in 3D



Direct
problem

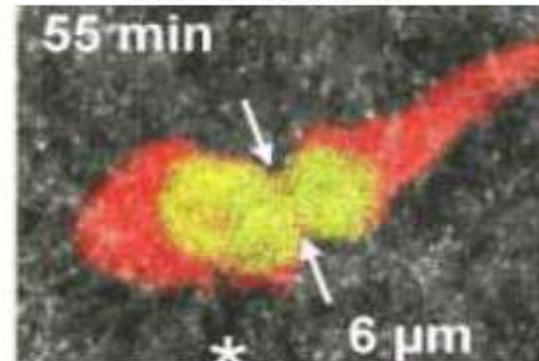


Inverse
problem

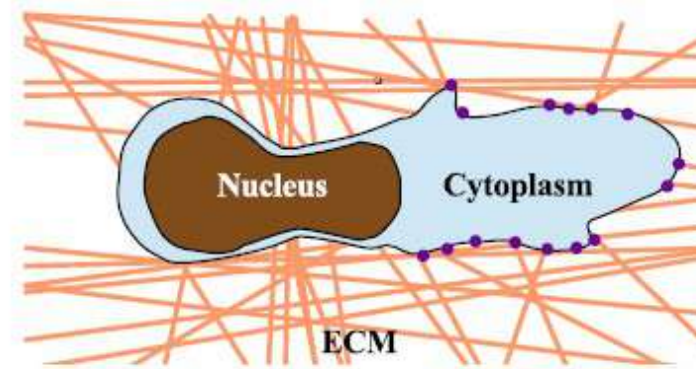


Nucleus squeezing

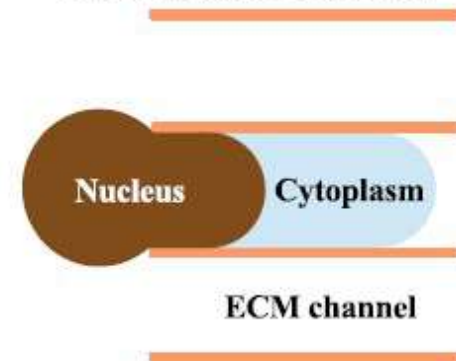
BIOLOGICAL EXPERIMENT



BIOLOGICAL REPRESENTATION



MATHEMATICAL REPRESENTATION



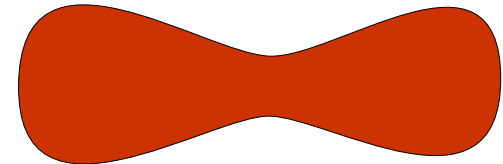
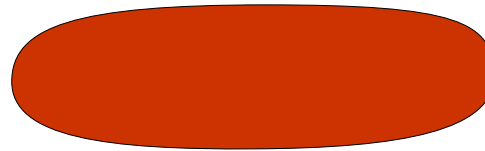
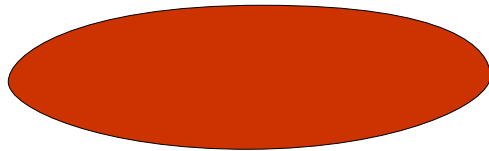
Work done by traction $>$ Energy required to squeeze the nucleus

C. Giverso & L.P., *Biomech. Model. Mechanobiol.* **13**, 481-502 (2014)

C. Giverso, A. Arduino & L.P., *Bull. Math. Biol.* **80**, 1017-1045 (2018)

Work done by traction $>$ Energy required to squeeze the nucleus

- Given the deformation gradient \mathbf{F}

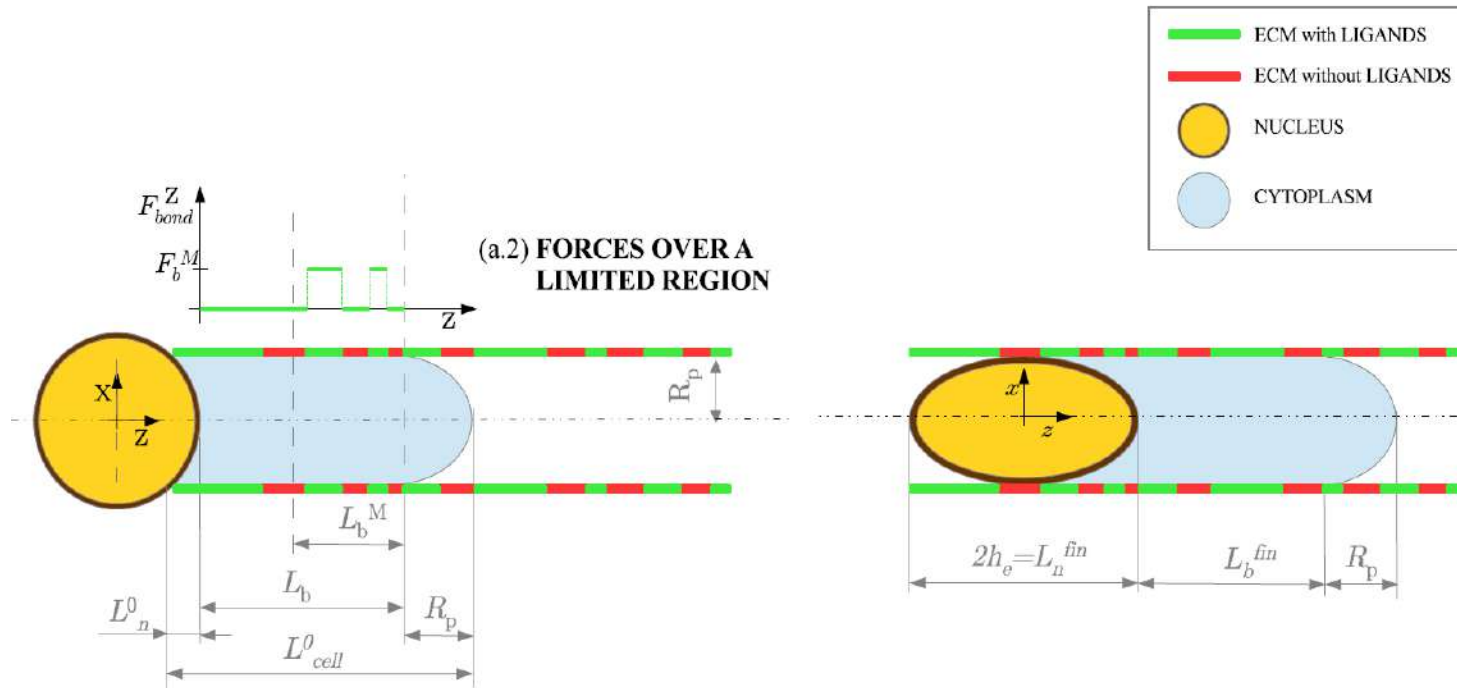


- Given the constitutive equation of the material

- Compute $\mathbf{B} = \mathbf{F} \mathbf{F}^T$

- Compute the elastic energy, e.g. $W(\mathbf{I}_{\mathbf{B}}) = \frac{\mu}{2}(\mathbf{I}_B - 3)$

Work done by traction > Energy required to squeeze the nucleus



$$\mathbf{F}_{adhesion} = \int_S \rho_b(\mathbf{X}) \alpha_{ECM}(\mathbf{X}) \mathbf{F}_{bond}(\mathbf{X}) dS$$

$$\mathcal{W}_{adhesion} = F_{adhesion}^Z \Delta L$$

Invasion criterium

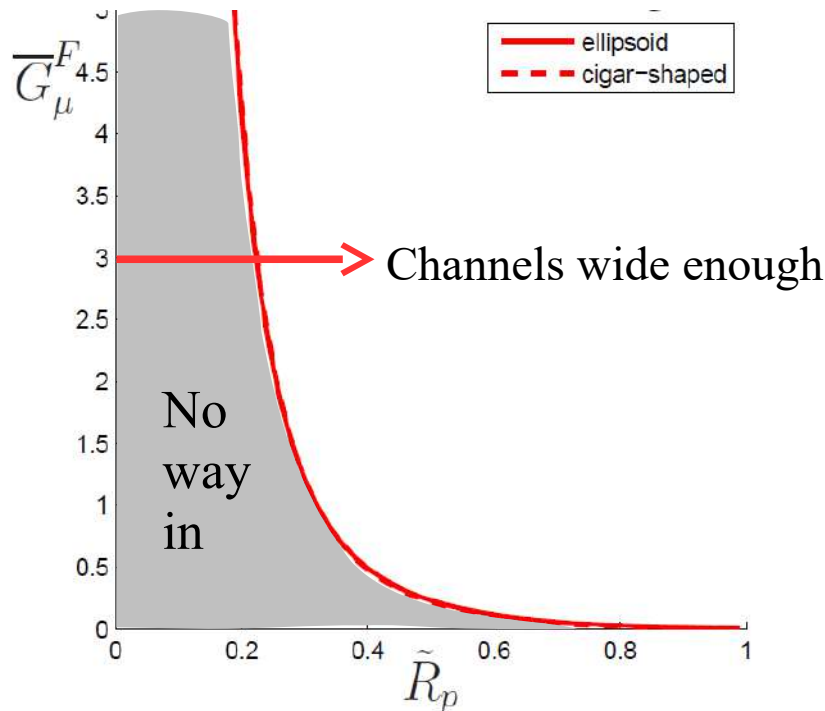
Work done by traction > Energy required to squeeze the nucleus

Elastic nucleus	Ellipsoid	$G_{\mu}^F \geq \frac{2}{3} \frac{2\tilde{R}_p^2 + \frac{1}{\tilde{R}_p^4} - 3}{\tilde{R}_p \tilde{L}_b^{(*)} \Delta \tilde{L}_{ellips}}$
	Cigar	$G_{\mu}^F \geq \frac{2}{3} \frac{\mathcal{I}(\tilde{R}_p)}{\tilde{R}_p \tilde{L}_b^{(*)} \Delta \tilde{L}_{cigar}}$

$$G_{\mu}^F = \frac{\rho_b \alpha_{ECM} F_b^M}{\mu}$$

Traction
Nucleus stiffness

$$\tilde{R}_p = R_p / R_n$$



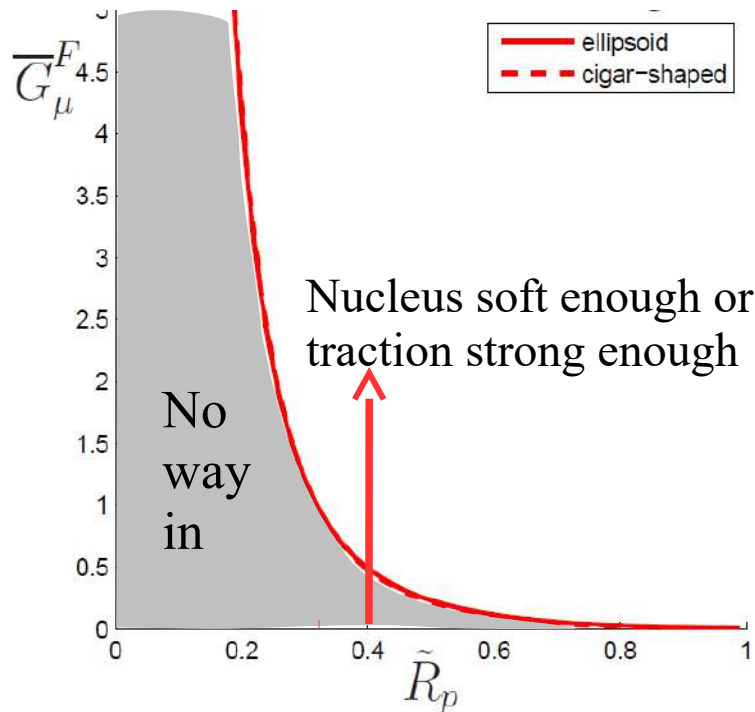
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	Cigar	$G_{\mu}^F \geq \frac{2}{3} \frac{\mathcal{I}(\tilde{R}_p)}{\tilde{R}_p \tilde{L}_b^{(*)} \Delta \tilde{L}_{cigar}}$

$$G_{\mu}^F = \frac{\text{Traction}}{\text{Nucleus stiffness}}$$

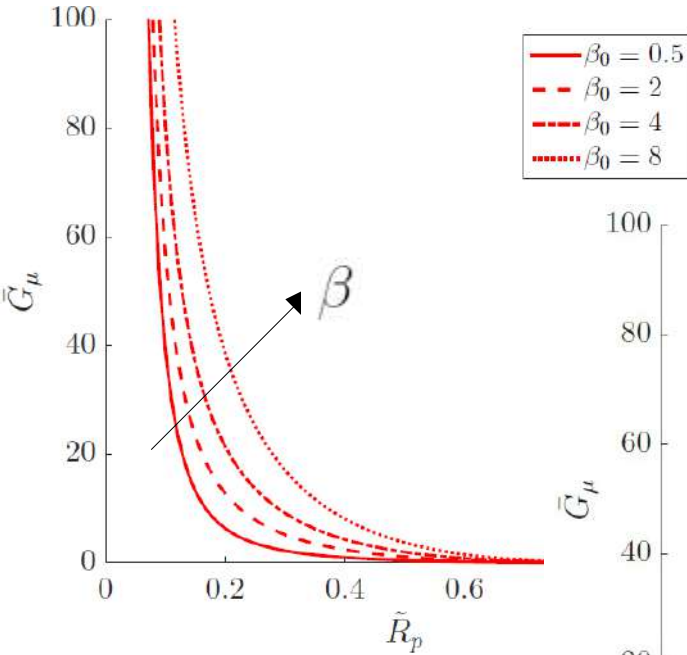
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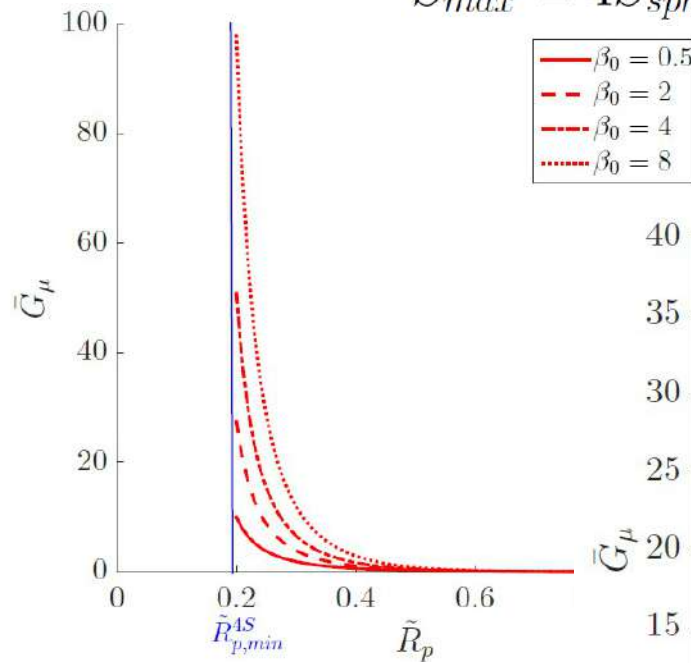
$$\tilde{R}_p = R_p / R_n$$

Effect of nuclear membrane

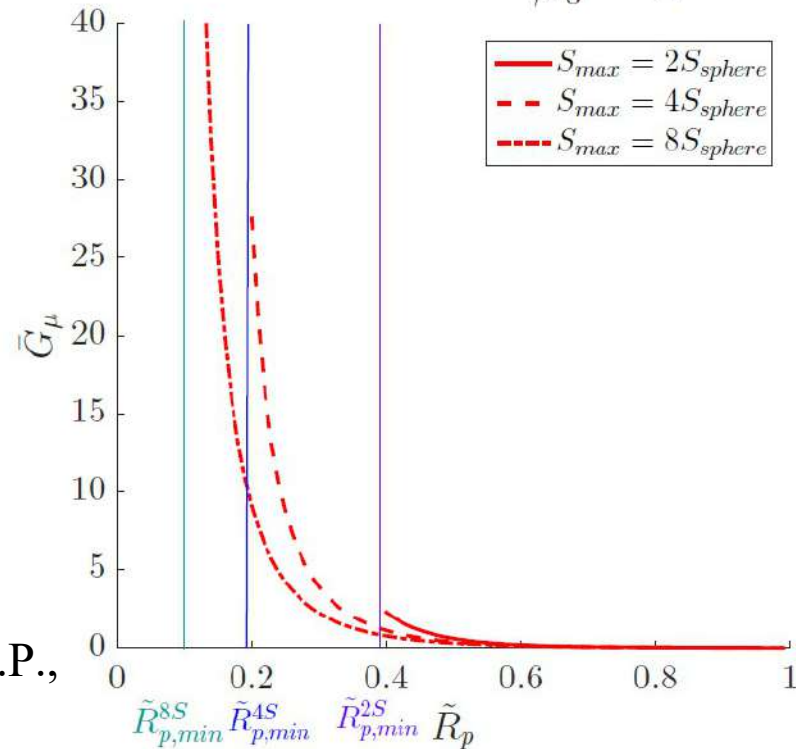
$$\lambda(\Delta S) = \lambda_0 \left(\frac{\Delta S}{S_{max} - S} \right)$$



$$S_{max} = 4S_{sphere}$$



$$\beta_0 = 2$$



$$\tilde{R}_p = R_p / R_n$$

Membrane

$$\beta := \frac{\lambda R_n}{\mu}$$

Nucleus

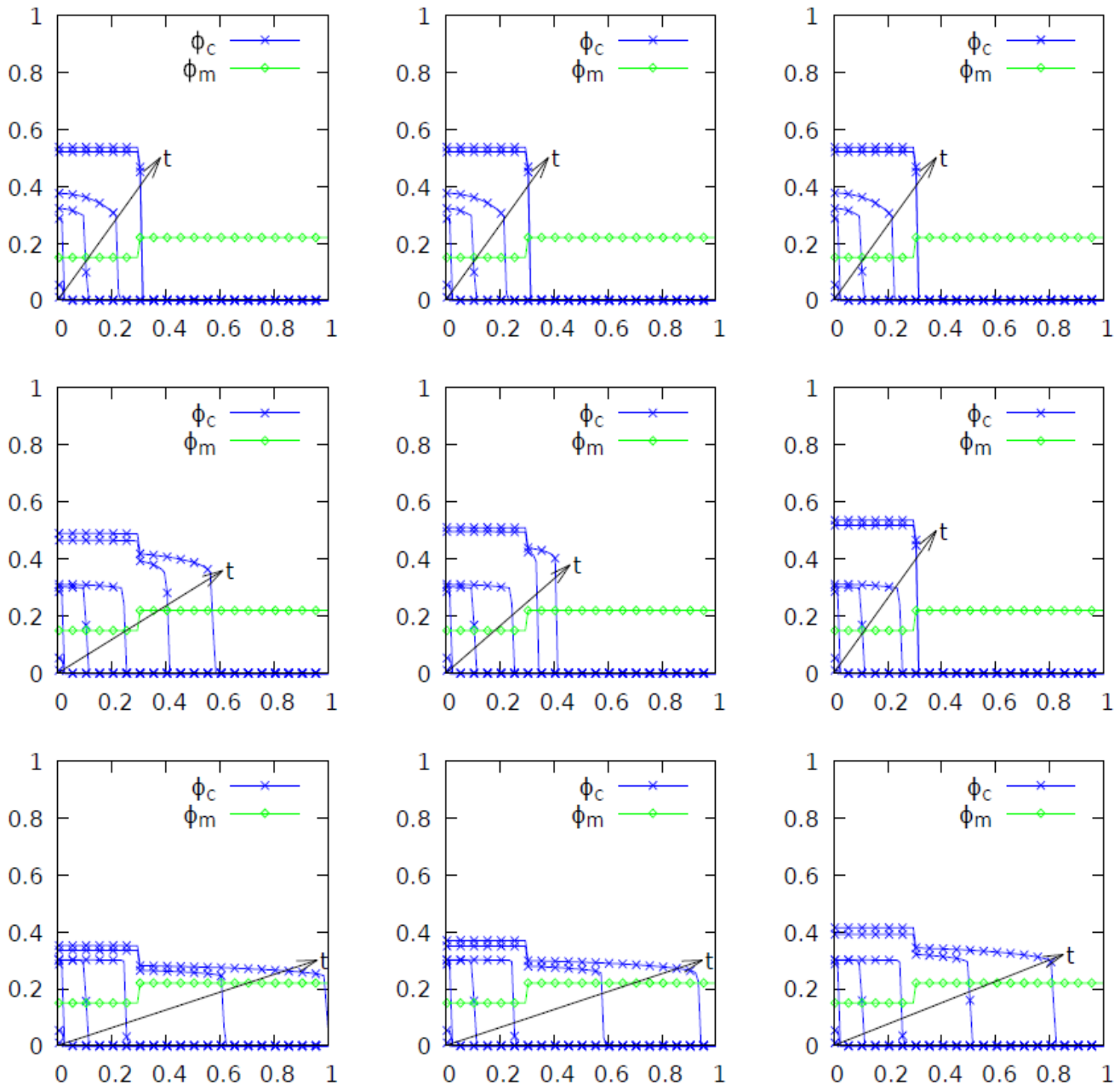
C. Givero, A. Arduino & L.P.,
Bull. Math. Biol. (2017)

Nuclear membrane stiffness

 $\beta_0 = 0.5$
 $\beta_0 = 2$
 $\beta_0 = 8$

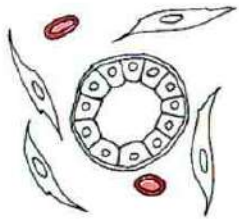
$$\beta := \frac{\lambda R_n}{\mu}$$

Bulk stiffness

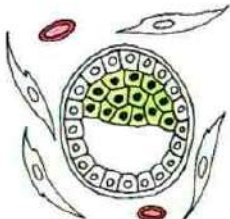


Compartmentalization or invasion

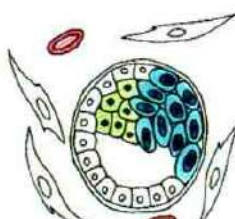
Normal duct



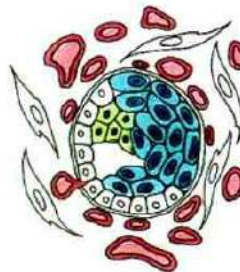
Hyperplasia



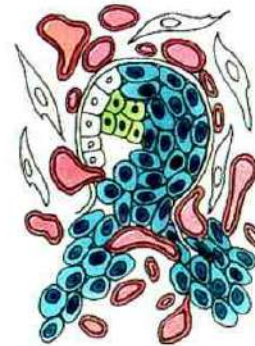
Dysplasia/ CIS



Angiogenic CIS



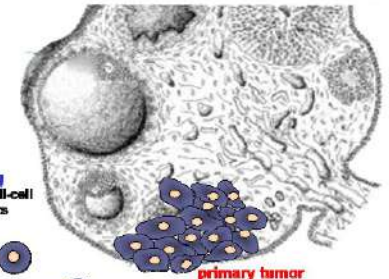
Invasive carcinoma



Breast

Ovary

Ovarian cancer dissemination



1) **Surface shedding**
Initial disruption of cell-cell and cell-matrix contacts

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disruption of cell-cell contacts in multi-cellular aggregates

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migration/invasion through mesothelial layer and into sub-mesothelial EC matrix

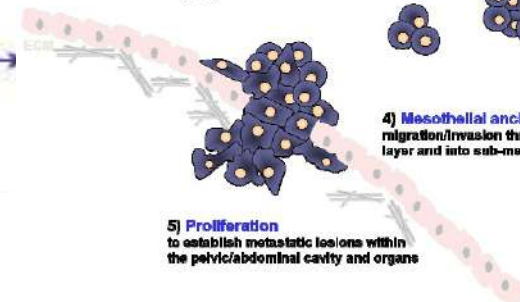
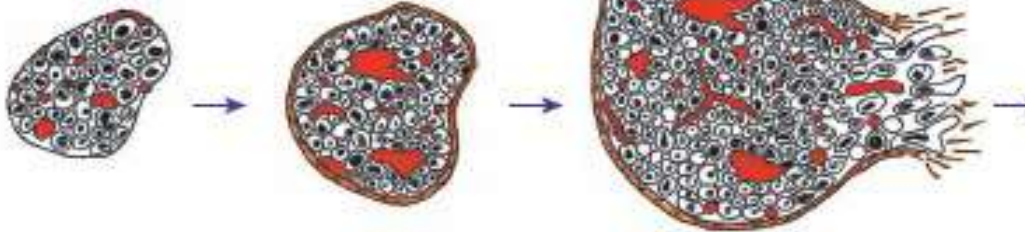
5) **Proliferation**
to establish metastatic lesions within the pelvic/abdominal cavity and organs

Pancreas

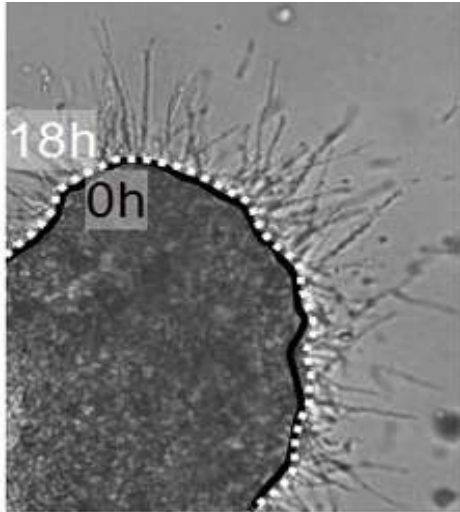
Angiogenic Dysplasia/CIS

Small Tumor

Large Tumor/
Invasive Carcinoma



Transfer to continuous models



$$\begin{cases} \frac{\partial \phi_c}{\partial t} + \nabla \cdot (\phi_c \mathbf{v}_c) = \Gamma_c \\ \nabla \cdot \mathbb{T}_c + \mathbf{m}_{cm} = \mathbf{0} \end{cases}$$

Growth

Stress

$$-\Sigma(\phi_c)\mathbb{I}$$

Cellular (solid) pressure

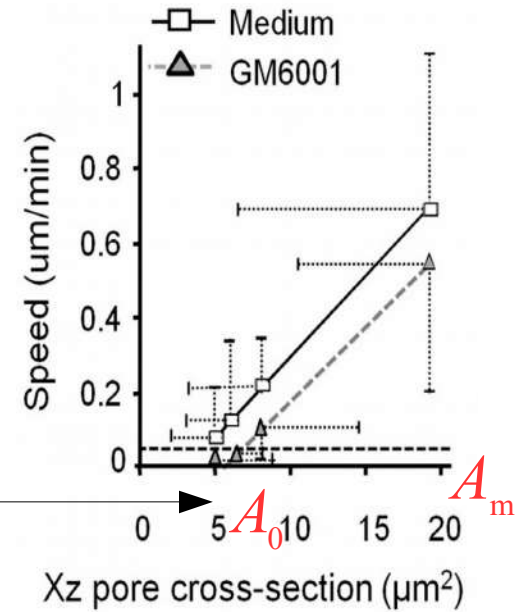
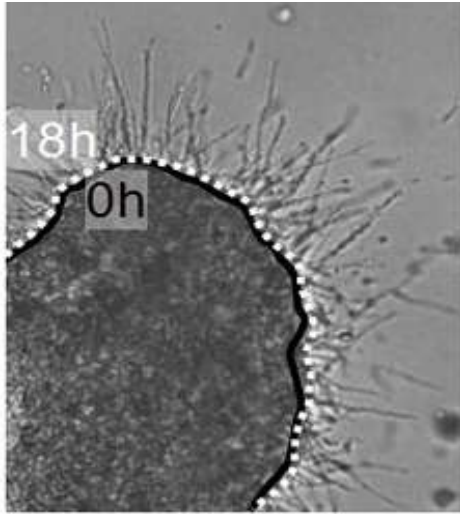
Interaction force

$$-\mathbb{M}_c^{-1}(\mathbf{v}_c - \cancel{\mathbf{v}_m})$$

Motility tensor

$$\mathbf{v}_c = -\mathbb{M}_c \nabla \Sigma$$

Transfer to continuous models



Implications for interaction force m_{cm} and motility M ?

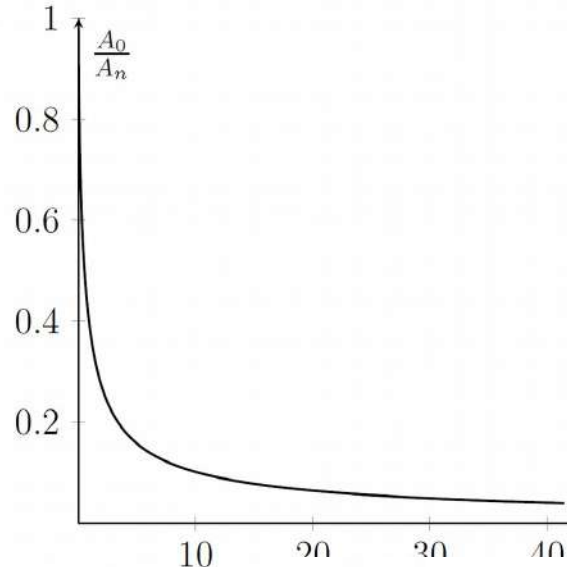
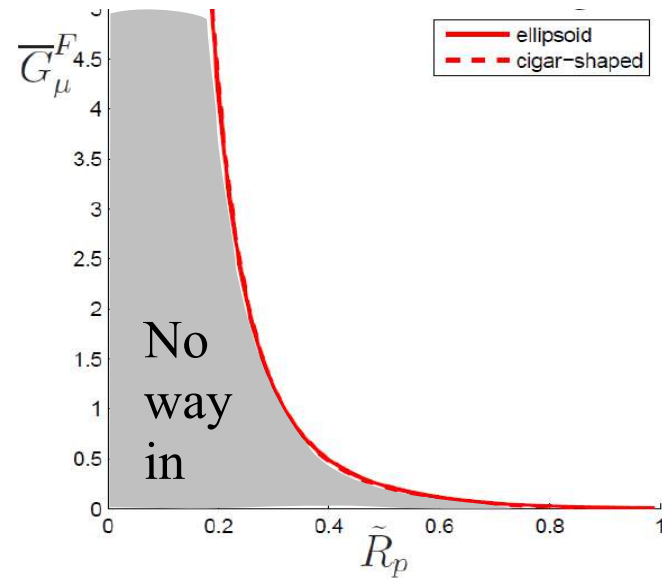
A_0 depends on:

- ✓ Pore vs. nucleus size
- ✓ Nucleus elasticity
- ✓ Cell adhesion
- ✓ Active traction

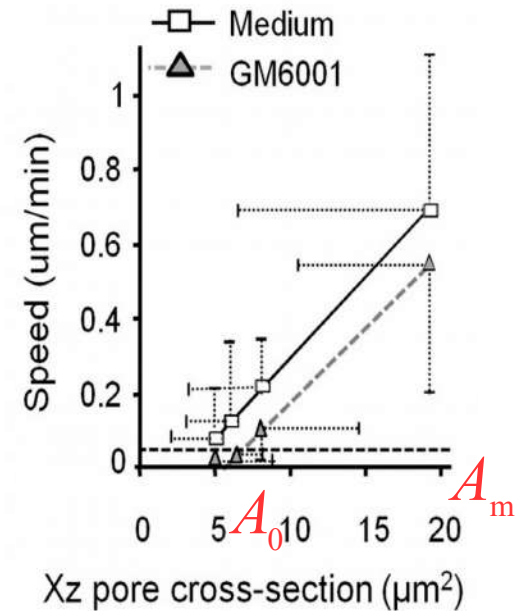
$$M_c \propto \frac{[A_m(\phi_m) - A_0]_+}{\left(1 + \frac{A_m(\phi_m) - A_0}{A_1}\right)^n} \quad \text{II}$$

Use in a continuous models

$$G_{\mu}^F = \frac{\rho_b \alpha_{ECM} F_b^M}{\mu}$$



$$G = \frac{1}{3\mu} |\text{tr} \mathbf{T}_c|$$



Implications for interaction force m_{cm} and motility K ?

A_0 depends on:

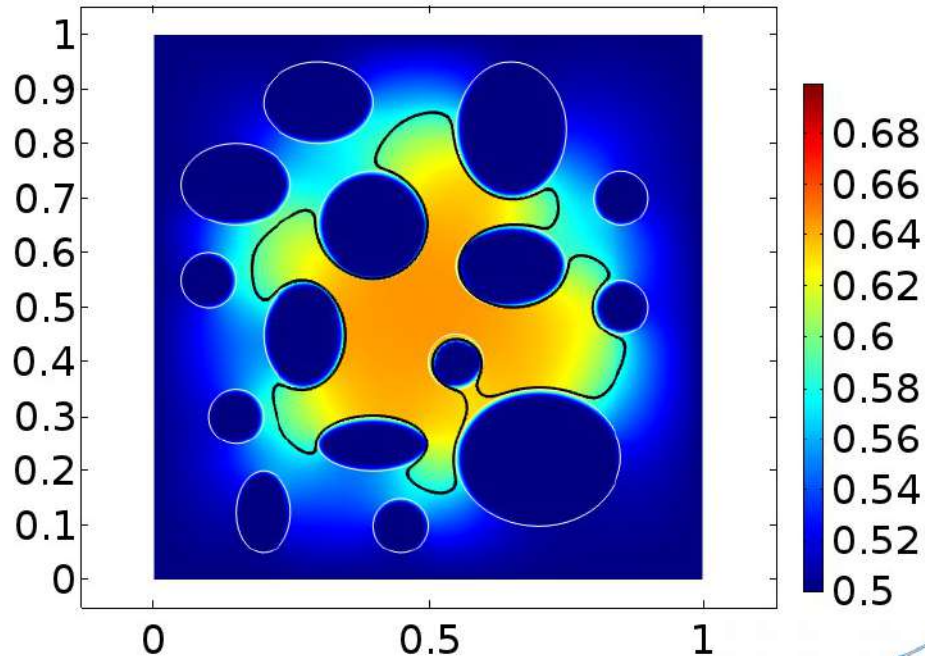
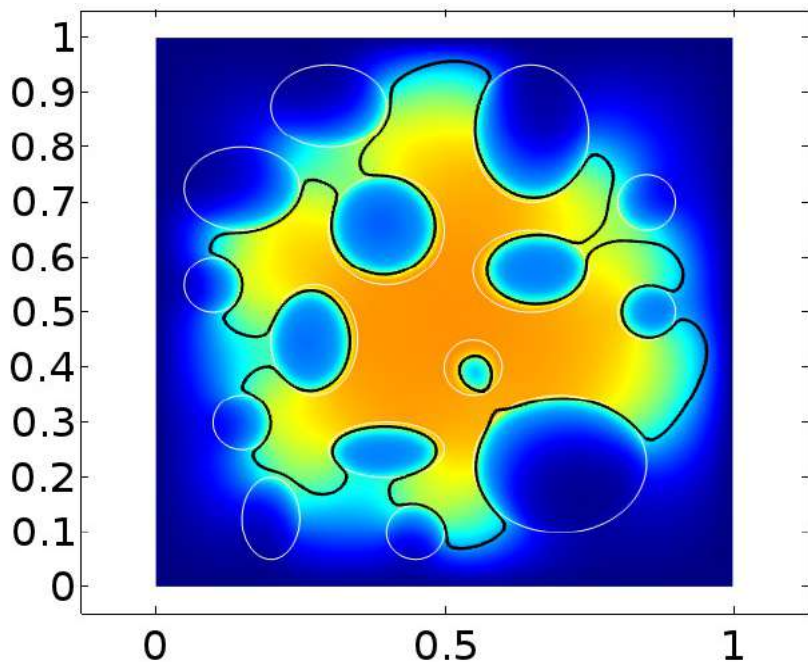
- ✓ Pore vs. nucleus size
- ✓ Nucleus elasticity
- ✓ Cell adhesion
- ✓ Active traction

$$\frac{\partial \phi_c}{\partial t} = \nabla \cdot (\phi_c \mathbb{M}_c \nabla \Sigma) + G_c$$

$$\propto \frac{[A_m(\phi_m) - A_0]_+}{\left(1 + \frac{A_m(\phi_m) - A_0}{A_1}\right)^n}$$

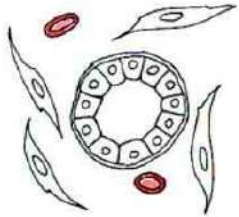
Growth in heterogeneous environments

$$\left\{ \begin{array}{l} \frac{\partial \phi_{c_i}}{\partial t} + \nabla \cdot (\phi_{c_i} \mathbf{v}_{c_i}) = \Gamma_{c_i} \left[\gamma_c^i \mathcal{H}_\varepsilon(\psi_0^i - \psi) - \delta_c^i \right] \phi_c^i \\ \mathbf{v}_{c_i} = \alpha \frac{[A_m(\phi_m) - A_0^i]_+}{\left(1 + \frac{A_m(\phi_m) - A_0^i}{A_1}\right)^n} \nabla \cdot \mathbf{T}_c \end{array} \right.$$

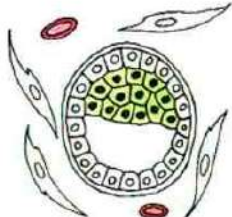


Compartmentalization or invasion

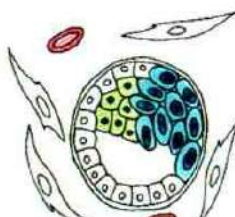
Normal duct



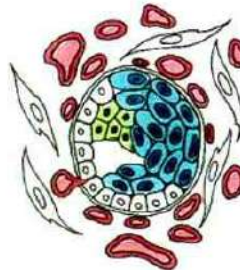
Hyperplasia



Dysplasia/ CIS



Angiogenic CIS



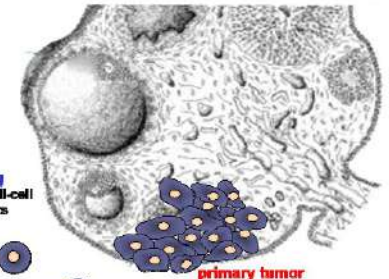
Invasive carcinoma



Breast

Ovary

Ovarian cancer dissemination



1) **Surface shedding**
Initial disruption of cell-cell and cell-matrix contacts

3) **Retraction, sub-mesothelial adhesion**
disruption of cell-cell contacts in multi-cellular aggregates

2) **Dissemination**
as single cells or multi-cellular aggregates (spheroids)

4) **Mesothelial anchoring**
migration/invasion through mesothelial layer and into sub-mesothelial EC matrix

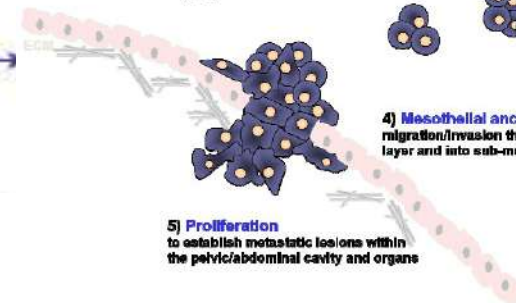
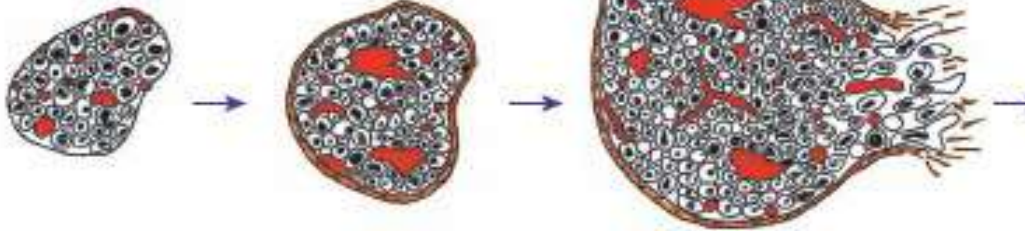
5) **Proliferation**
to establish metastatic lesions within the pelvic/abdominal cavity and organs

Pancreas

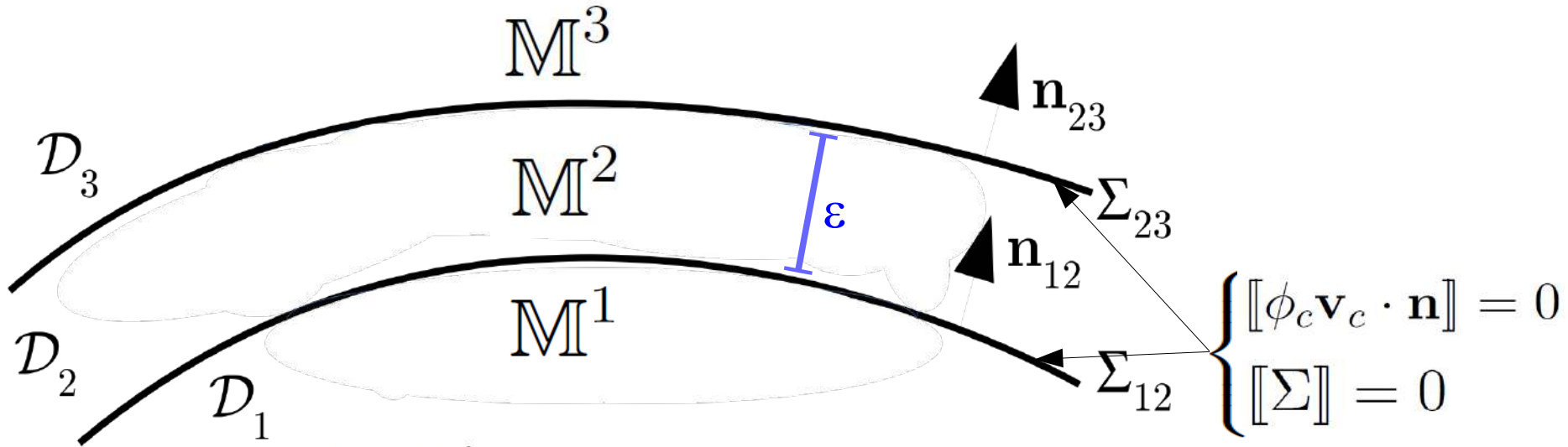
Angiogenic Dysplasia/CIS

Small Tumor

Large Tumor/
Invasive Carcinoma



Basal membranes

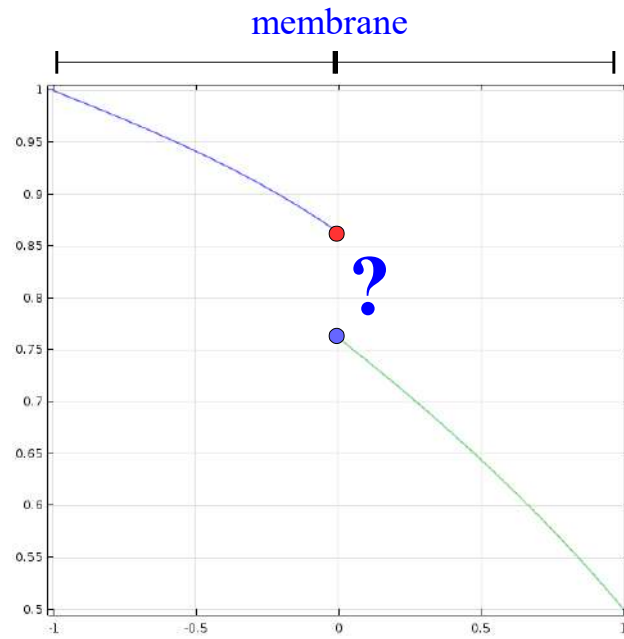
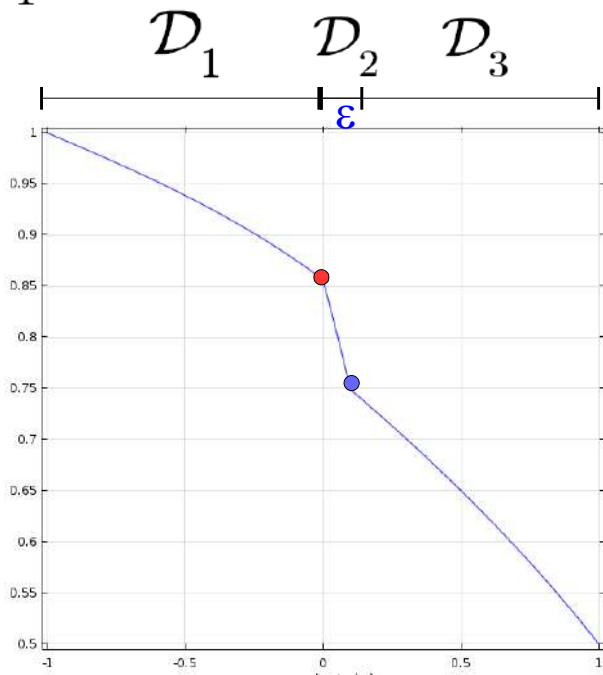
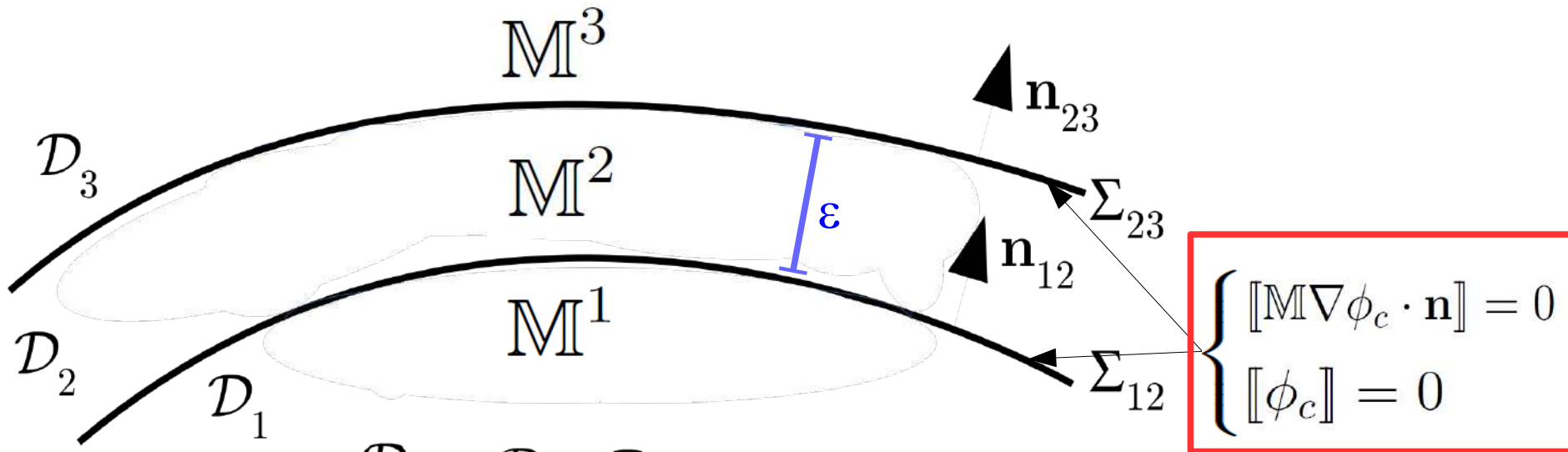


$$\begin{cases} \frac{\partial \phi_c^i}{\partial t} + \nabla \cdot (\phi_c^i \mathbf{v}_c^i) = \Gamma_c^i \\ \mathbf{v}_c^i = -M^i \nabla \Sigma(\phi_c^i) \end{cases}$$

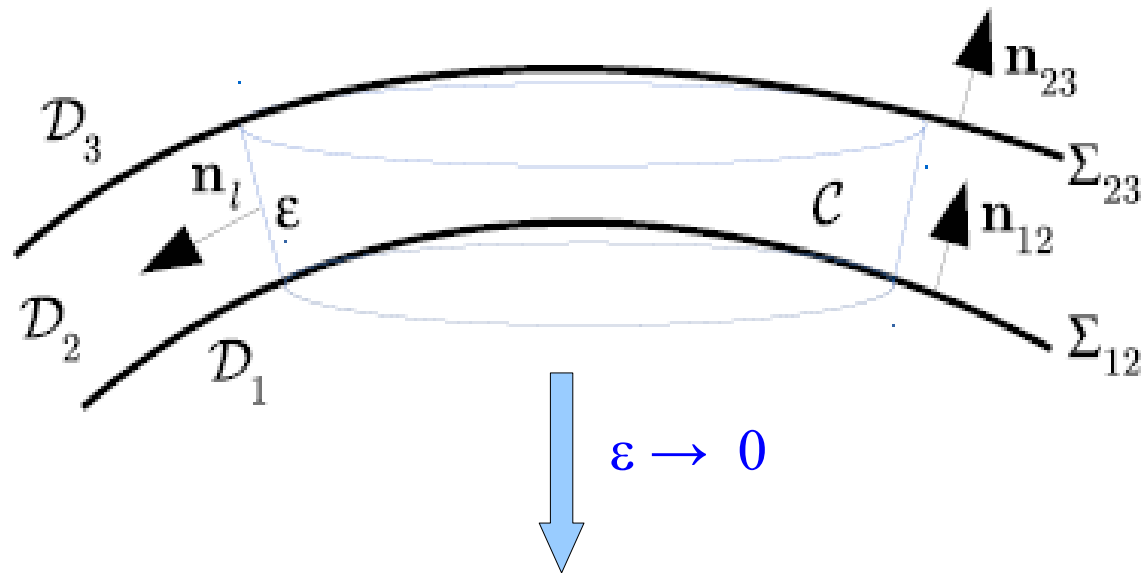


$$\frac{\partial \phi_c^i}{\partial t} = \nabla \cdot [\phi_c^i M^i \nabla \Sigma(\phi_c^i)] + \Gamma_c^i$$

Basal membranes



Basal membranes

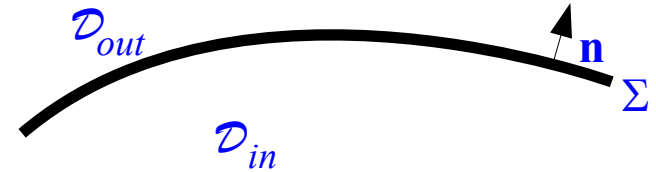


$$\phi_c^1 M^1 \nabla \Sigma(\phi_c^1) \cdot \mathbf{n} = \phi_c^3 M^3 \nabla \Sigma(\phi_c^3) \cdot \mathbf{n} = M_\sigma [\Pi]$$

where $\Pi'(\phi_c) = \phi_c \Sigma'(\phi_c)$

$$M_\sigma = \lim_{\varepsilon \rightarrow 0} \frac{M^2}{\varepsilon}$$

$$\left\{ \begin{array}{l} \frac{\partial \phi_c}{\partial t} = \nabla \cdot [\phi_c M_{cm} \nabla \Sigma(\phi_c)] + \Gamma_c \\ \llbracket \phi_c M_{cm} \nabla \Sigma(\phi_c) \cdot \mathbf{n} \rrbracket = 0 \quad \text{on } \Sigma \\ M_\sigma \llbracket \Pi \rrbracket = \phi_c M_{cm} \nabla \Sigma(\phi_c) \cdot \mathbf{n} \quad \text{on } \Sigma \end{array} \right.$$



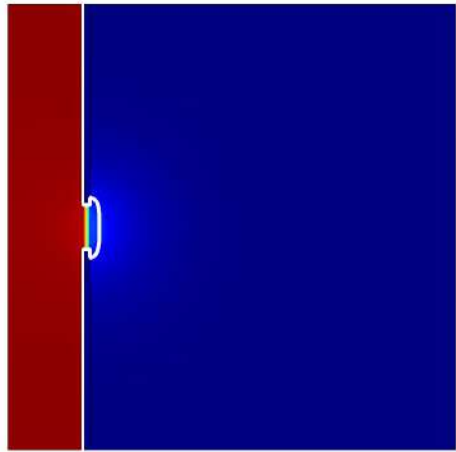
where $\Pi'(\phi_c) = \phi_c \Sigma'(\phi_c)$

$$\text{if } \Sigma(\phi_c) = A \ln \frac{\phi_c}{\phi_0} \quad \longrightarrow \quad \phi_c \Sigma'(\phi_c) = A \quad \longrightarrow \quad \Pi = A \phi_c$$

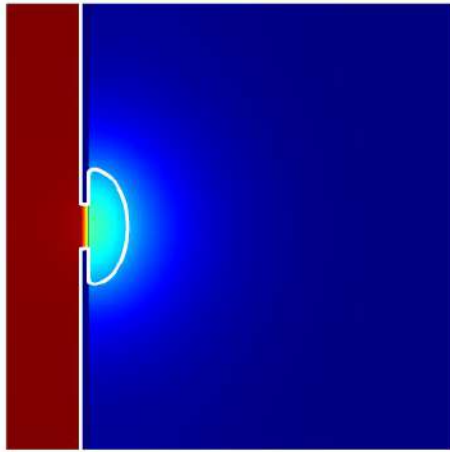
$$\longrightarrow M_\sigma \llbracket \phi_c \rrbracket = M_{cm} \nabla \phi_c \cdot \mathbf{n}$$

Kedem-Katchalsky interface condition

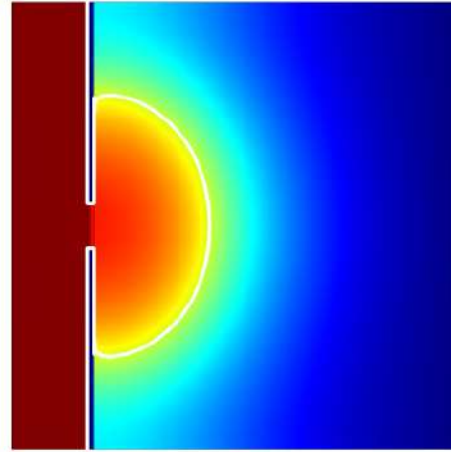
Invasion through a duct wall



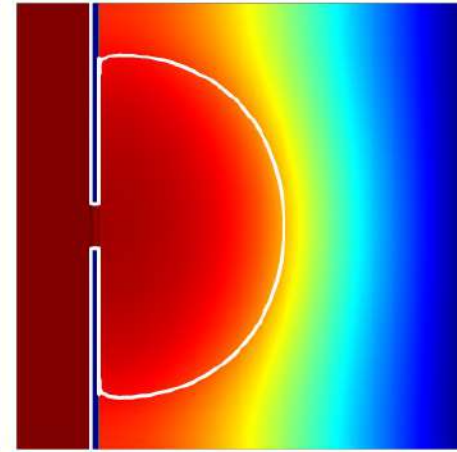
(a) $t = 5$



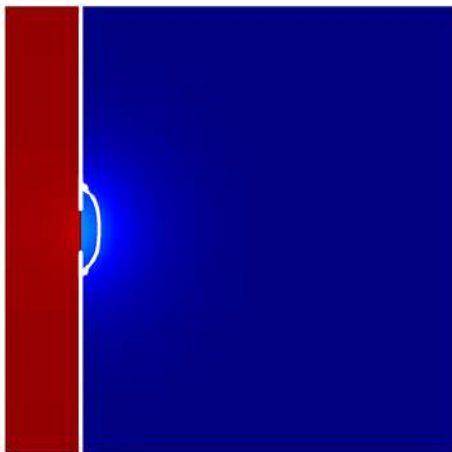
(b) $t = 10$



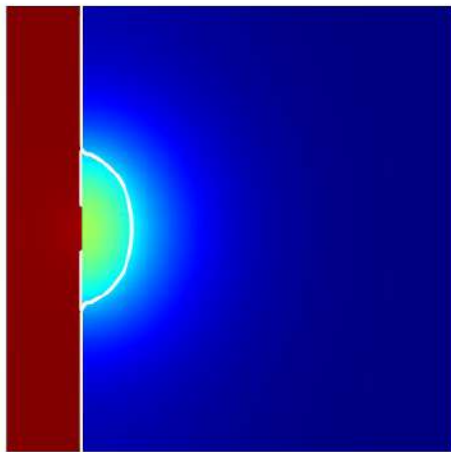
(c) $t = 20$



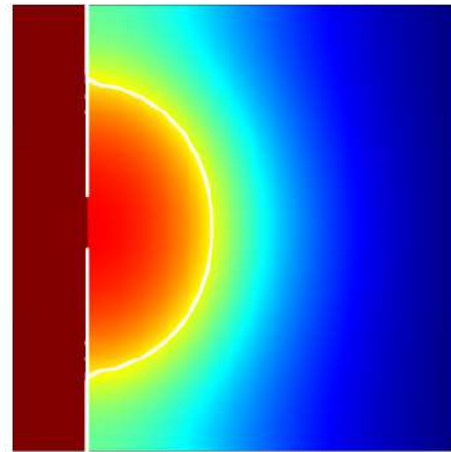
(d) $t = 30$



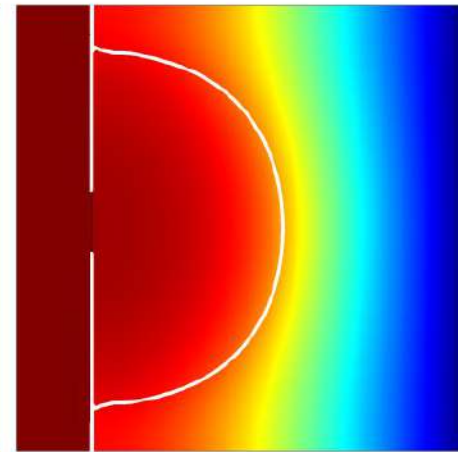
(e) $t = 5$



(f) $t = 10$

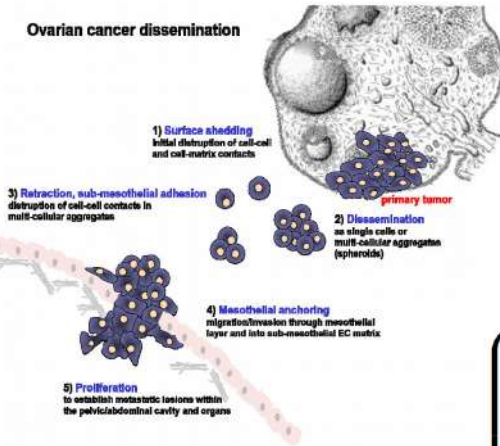


(g) $t = 20$



(h) $t = 30$

Ovarian cancer dissemination



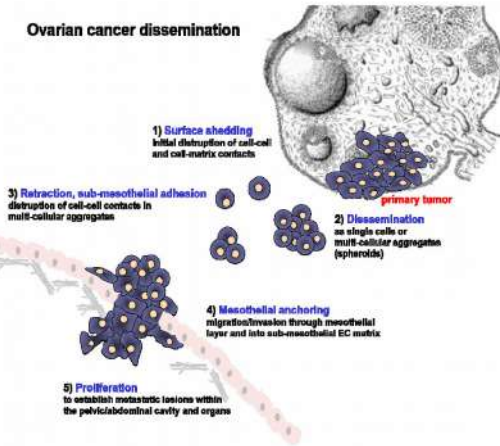
Adding tumour produced metallo-proteinases

$$\left\{ \begin{array}{l} \frac{\partial c_i}{\partial t} = \gamma_c \rho_i H(\varphi) + D_c \Delta c_i \quad \text{in } \mathcal{D}_i, \quad i = 1, 2, 3, \\ D_c \nabla c_i \cdot \mathbf{n}_{ij} = D_c \nabla c_j \cdot \mathbf{n}_{ij} \quad \text{on } \Sigma_{ij}, \quad i = 1, 2, \quad j = i + 1, \\ c_i = c_j \quad \text{on } \Sigma_{ij}, \quad i = 1, 2, \quad j = i + 1. \end{array} \right.$$

$$\mu_{23}[\hat{c}](t, \mathbf{x}) := \bar{\mu}_{23} \frac{(\hat{c}(t, \mathbf{x}) - 1)_+}{K_c + (\hat{c}(t, \mathbf{x}) - 1)},$$

$$\hat{c}(t, \mathbf{x}) = \frac{\beta}{\alpha(A_0 - A_1)} c(t, \mathbf{x})$$

Ovarian cancer dissemination



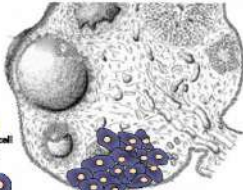
Adding tumour produced metallo-proteinases

$$\frac{dA}{dt} = \alpha (A_1 - A) + \beta c_{MMP}$$

$$M_\sigma = \bar{M}_\sigma \frac{[A - A_0]_+}{1 + (A - A_0)}$$

Invasion of ovary cancer cells

Ovarian cancer dissemination



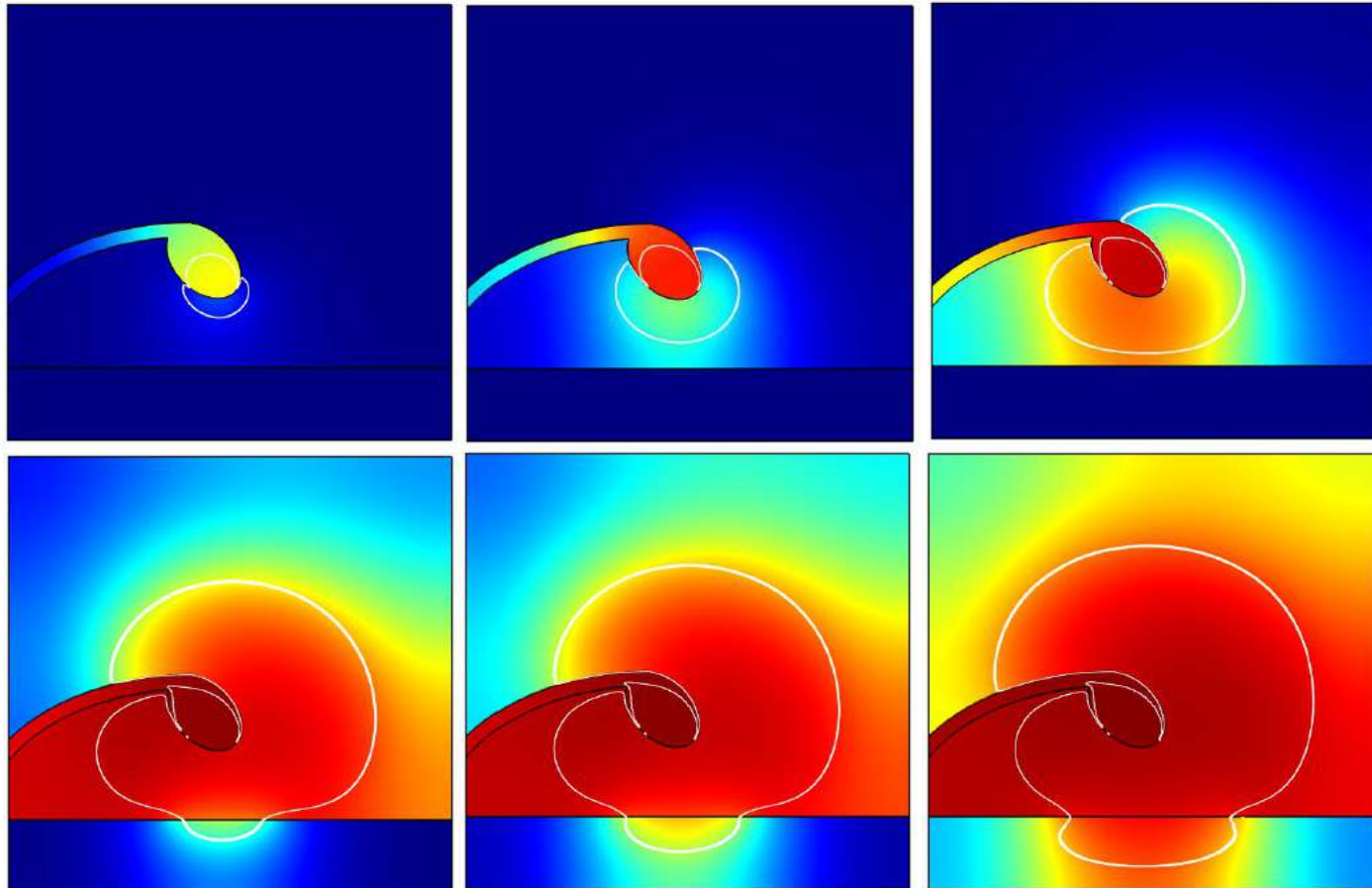
1) **Surface shedding**
initial disruption of cell-cell and cell-matrix contacts

3) **Retraction, sub-mesothelial adhesion**
disruption of cell-cell contacts in multi-cellular aggregates

2) **Dissemination**
as single cells or multi-cellular aggregates (spheroids)

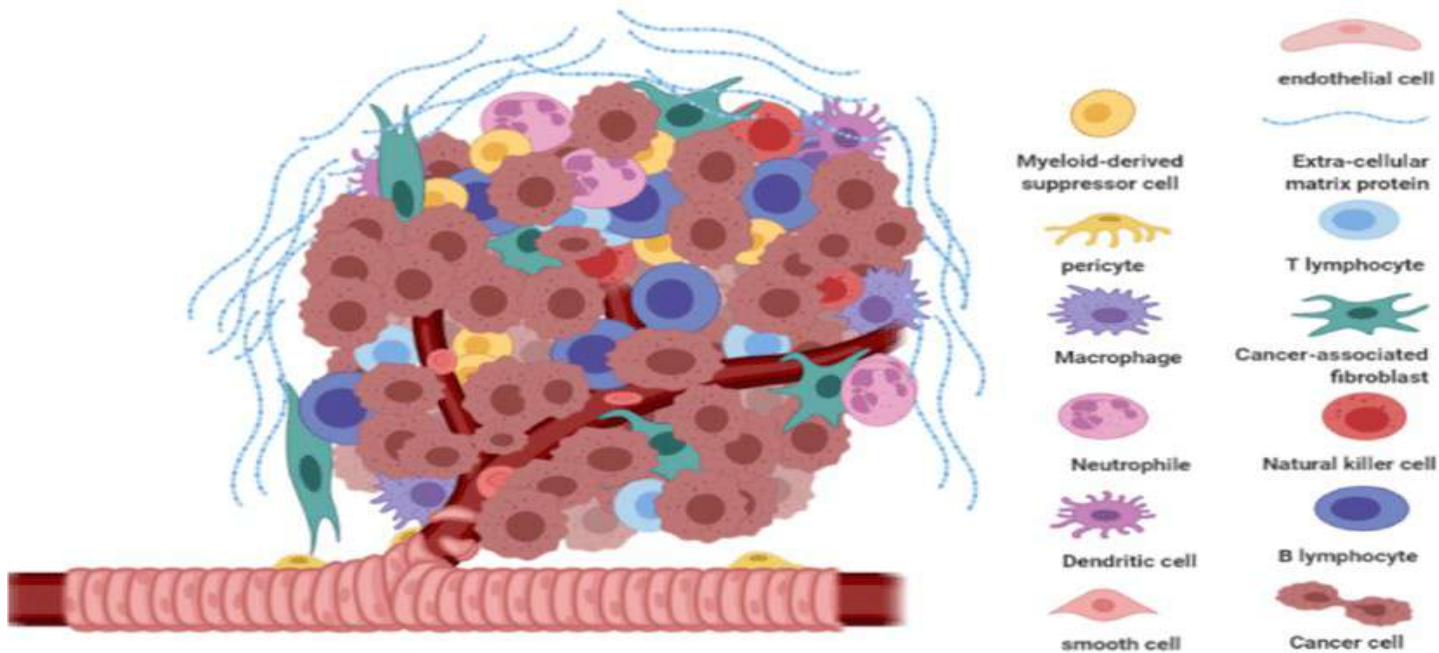
4) **Mesothelial anchoring**
migration/invasion through mesothelial layer and into sub-mesothelial EC matrix

5) **Proliferation**
to establish metastatic lesions within the peritoneal cavity and organs



More populations

$$\begin{cases} \frac{\partial \phi_{c_i}}{\partial t} + \nabla \cdot (\phi_{c_i} \mathbf{v}_{c_i}) = \Gamma_{c_i} \\ \mathbf{v}_{c_i} = -\mathbb{M}_{c_i m} \nabla \Sigma \quad \text{with } \Sigma = \Sigma(\phi_{c_i}, \Phi_c) \quad \Phi_c = \sum_i \phi_{c_i} \end{cases}$$



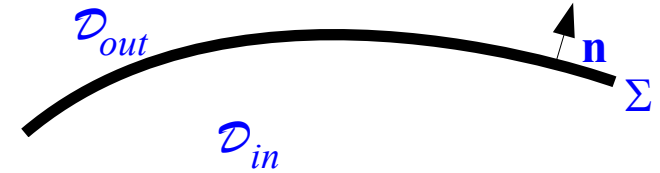
More populations

$$\begin{cases} \frac{\partial \phi_{c_i}}{\partial t} + \nabla \cdot (\phi_{c_i} \mathbf{v}_{c_i}) = \Gamma_{c_i} \\ \mathbf{v}_{c_i} = -\mathbb{M}_{c_i m} \nabla \Sigma \quad \text{with } \Sigma = \Sigma(\cancel{\phi_{c_i}}, \Phi_c) \quad \Phi_c = \sum_i \phi_{c_i} \end{cases}$$

⇒ All velocities are “proportional”

If $\mathbb{M}_{c_i m} = \mathbb{M}_{cm}$ and $\Sigma = \Sigma(\Phi_c)$ ⇒ $\mathbf{v}_{c_i} = \mathbf{v}_c$

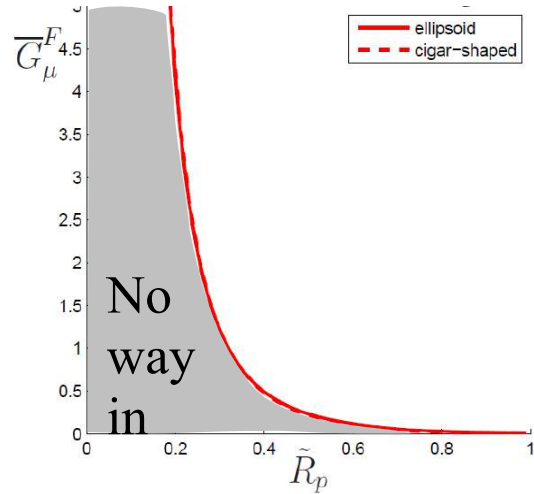
If $M_{\sigma}^1, M_{\sigma}^2 \neq 0$



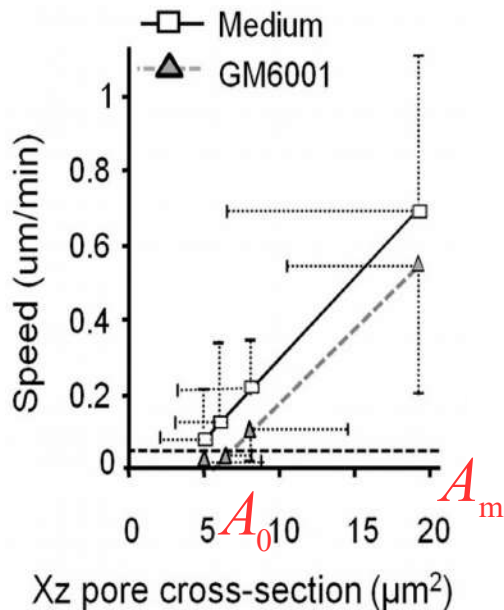
$$\left\{ \begin{array}{l} \frac{\partial \phi_c^n}{\partial t} = \nabla \cdot [\phi_c^n M_{cm}^n \nabla \Sigma(\Phi_c)] + \Gamma_c^n \\ [\phi_c^n M_{cm}^n \nabla \Sigma(\Phi_c) \cdot \mathbf{n}] = 0 \quad \text{on } \Sigma \\ [\Pi] = \left(\phi_c^1 \frac{M_{cm}^1}{M_{\sigma}^1} + \phi_c^2 \frac{M_{cm}^2}{M_{\sigma}^2} \right) \nabla \Sigma(\Phi_c) \cdot \mathbf{n} \quad \text{on } \Sigma \end{array} \right.$$

where $\Pi'(\Phi_c) = \Phi_c \Sigma'(\Phi_c)$

Work done by traction > Energy required to squeeze the nucleus

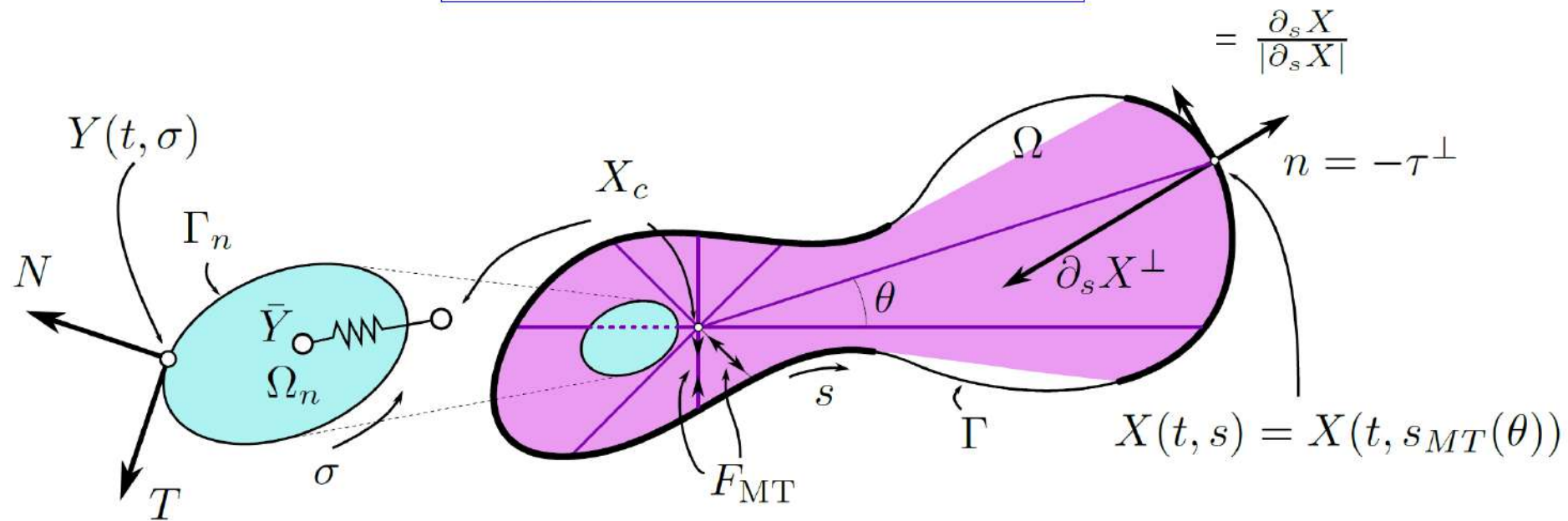
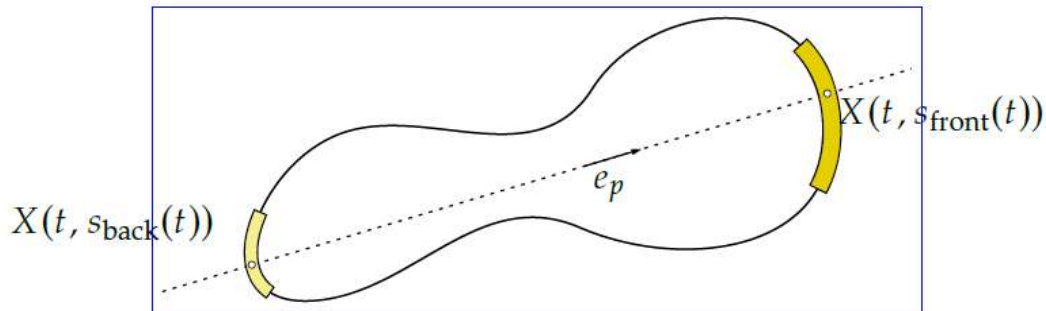


$$\left\{ \begin{array}{l} \frac{\partial \phi_c}{\partial t} = \nabla \cdot [\phi_c M_{cm} \nabla \Sigma(\phi_c)] + \Gamma_c \\ [\phi_c M_{cm} \nabla \Sigma(\phi_c) \cdot \mathbf{n}] = 0 \quad \text{on } \Sigma \\ M_\sigma [\Pi] = \phi_c M_{cm} \nabla \Sigma(\phi_c) \cdot \mathbf{n} \quad \text{on } \Sigma \end{array} \right.$$

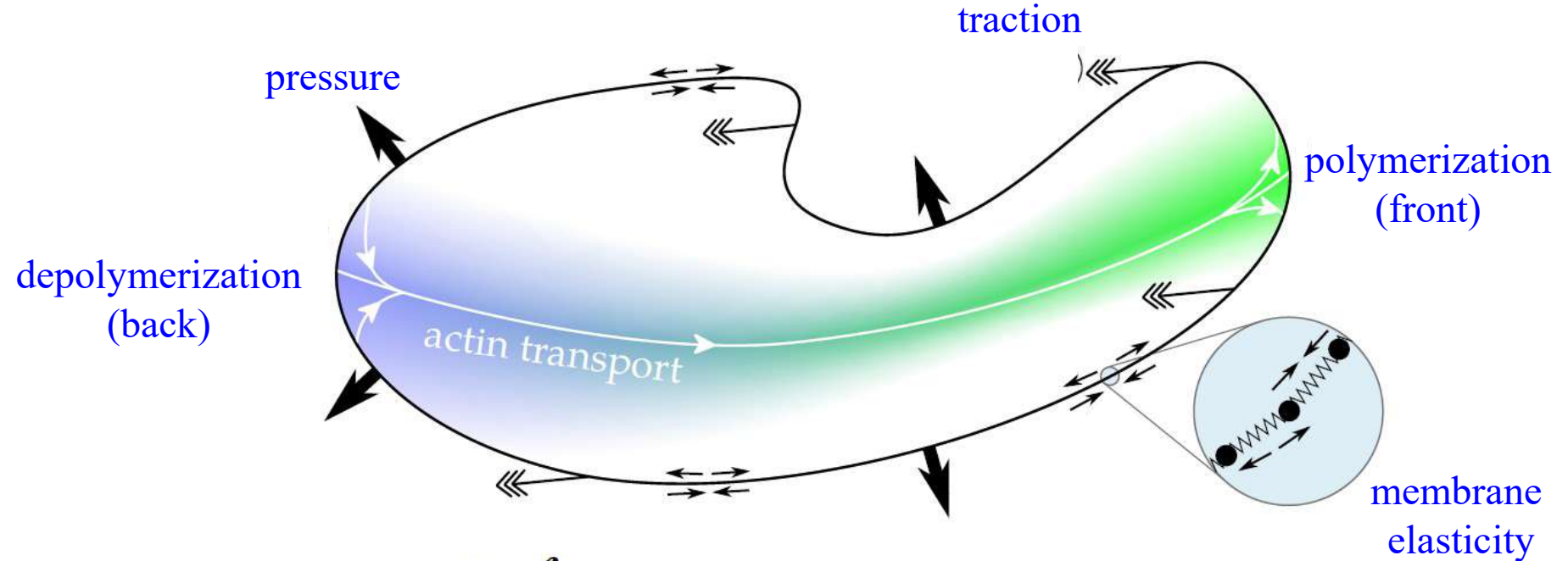


$$\propto \frac{[A_m(\phi_m) - A_0]_+}{\left(1 + \frac{A_m(\phi_m) - A_0}{A_1}\right)^n}$$

Single cell model



Single cell model: cytosol

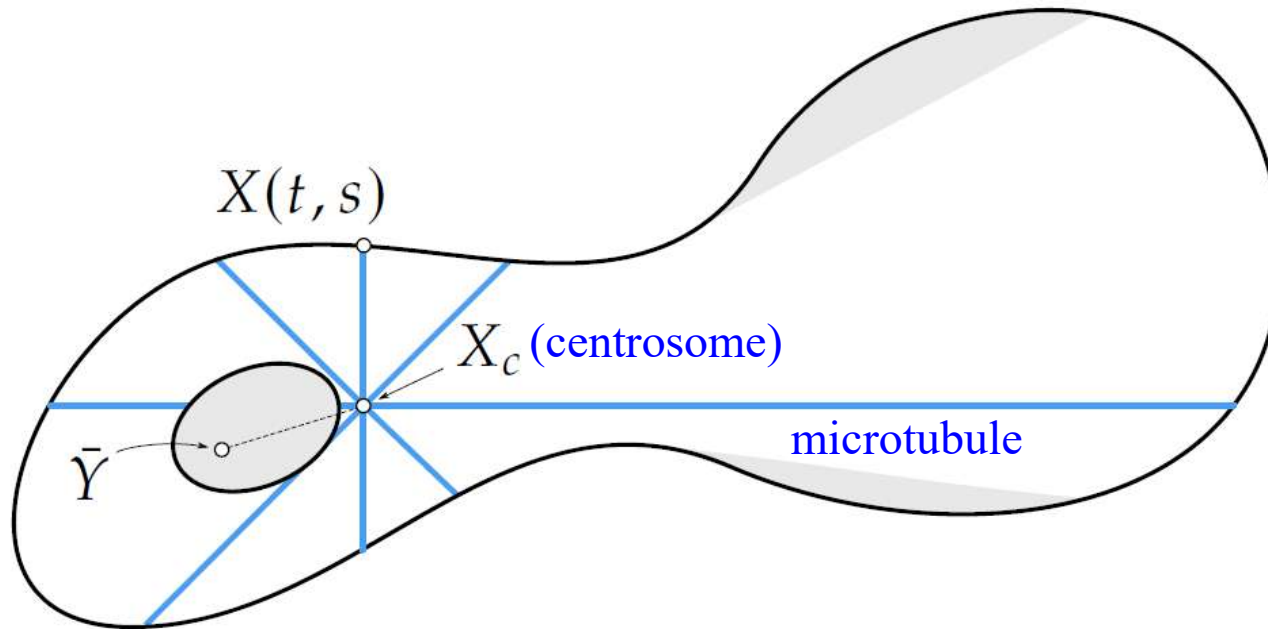


$$E_p(X) = -p |\Omega(t)| = \frac{p}{2} \int_{\mathbb{T}^1} X \cdot \partial_s X^\perp ds \quad \text{Cell pressure}$$

$$E_{el}(X) = \frac{1}{2} \int_{\mathbb{T}^1} (|\partial_s X| - 1)_+^2 ds \quad \text{Membrane elasticity}$$

$$E_{\text{obst},\delta}(X) = \int_{\mathbb{T}^1} W_\delta(X) ds \quad \text{Cell-wall interactions}$$

Single cell model: the nucleus



Bending stiffness

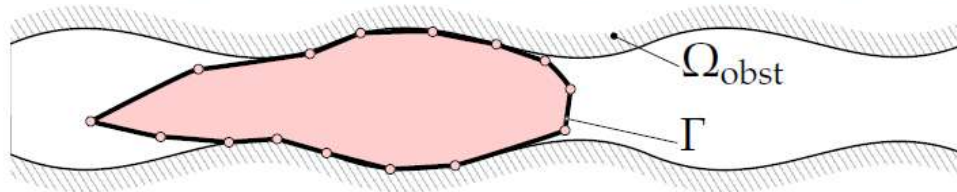
Pressure

$$E_n = \frac{k_b}{2} \int_{\Gamma_n} \kappa^2 d\ell + \int_{\Gamma_n} W_n(Y(\ell)) d\ell + \Delta p_n \int_{\Omega_n} dA$$

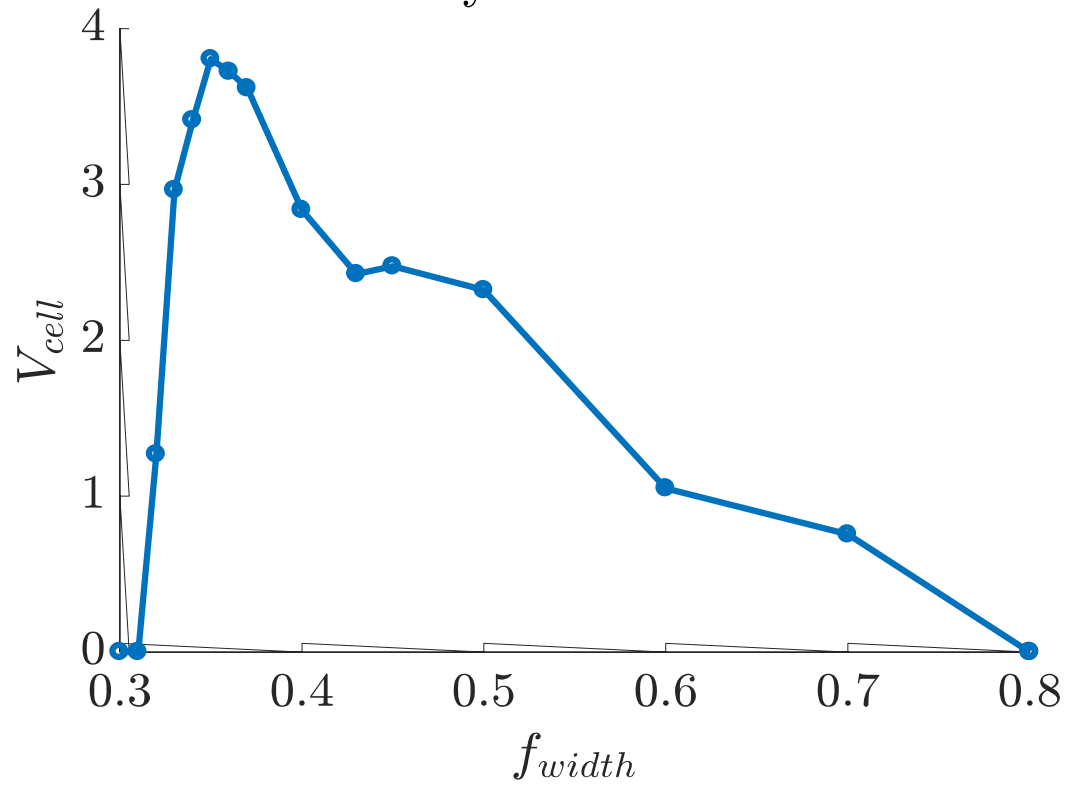
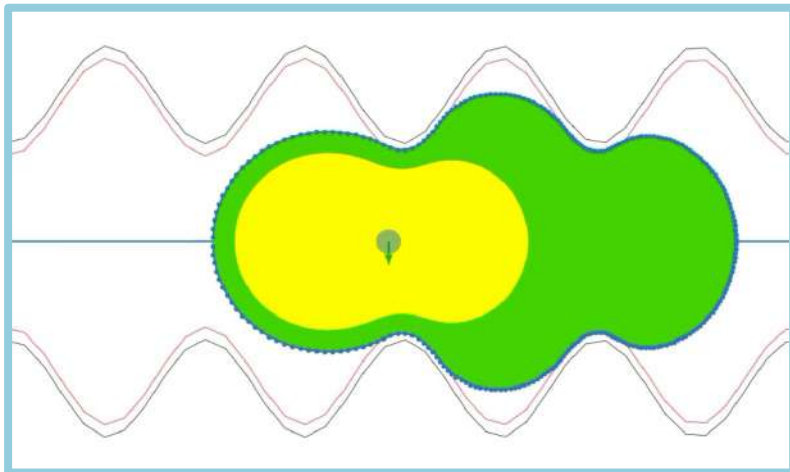
The potential W_n encodes elasticity and the interactions with cortex and centrosome.



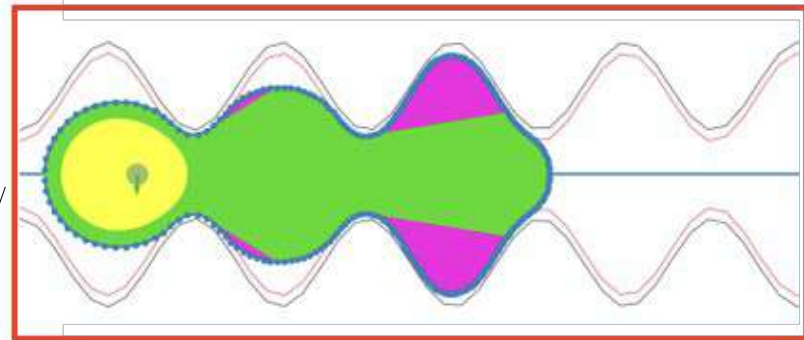
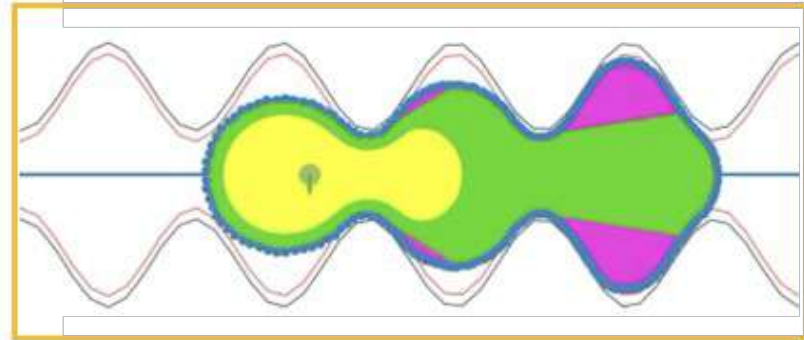
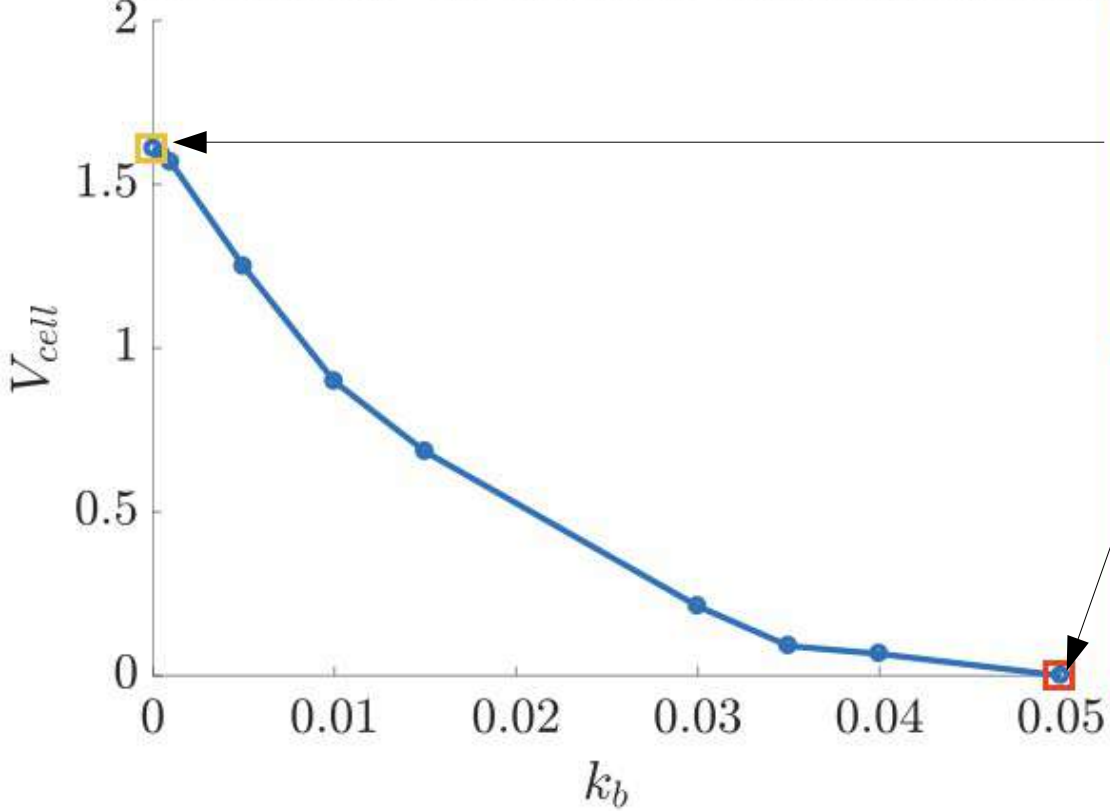
Simulations

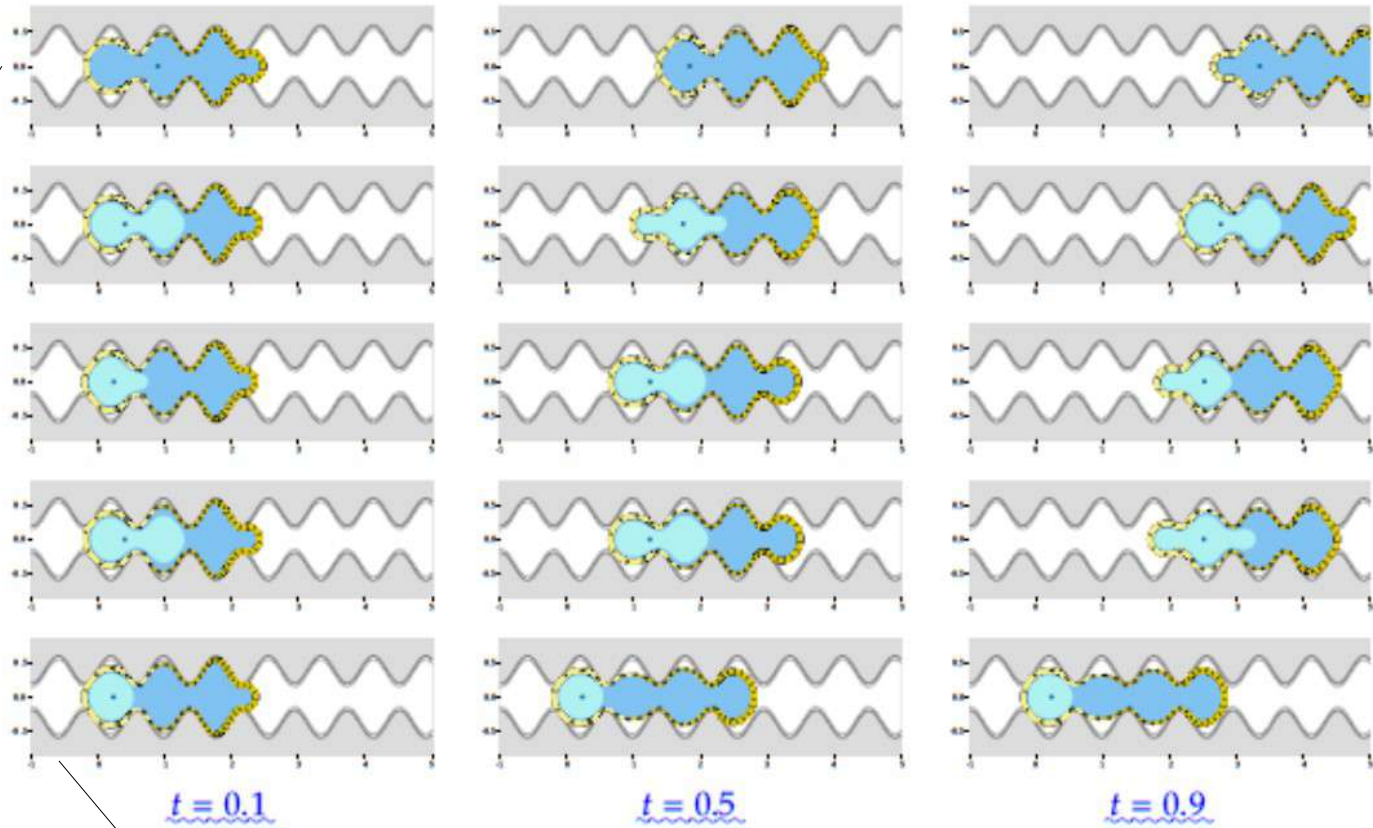


Cell velocity vs. Channel width

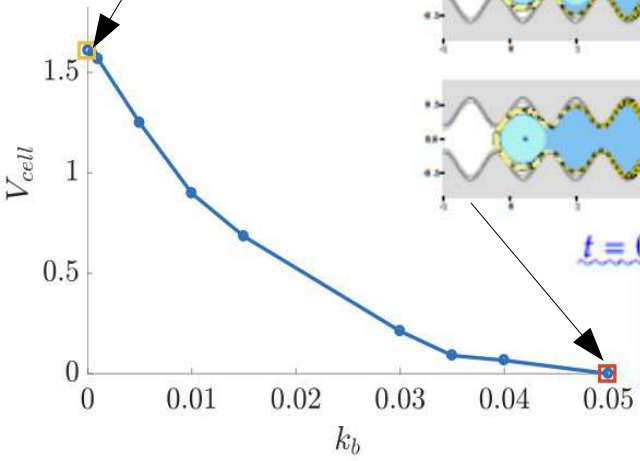


Cell velocity vs. Nucleus bending stiffness

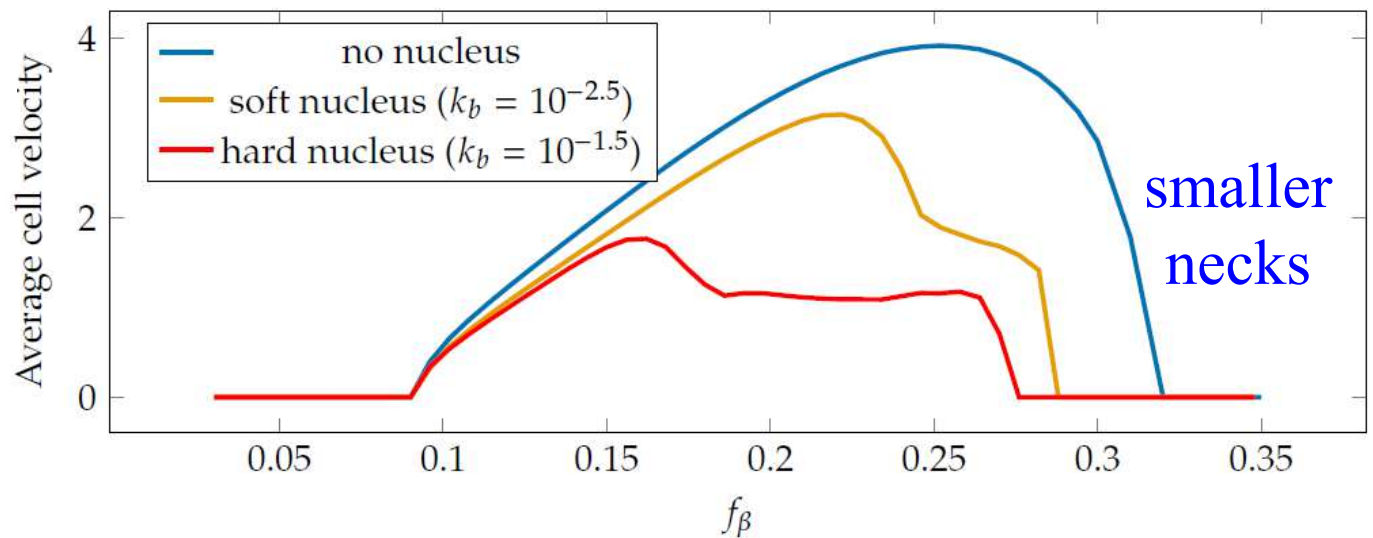
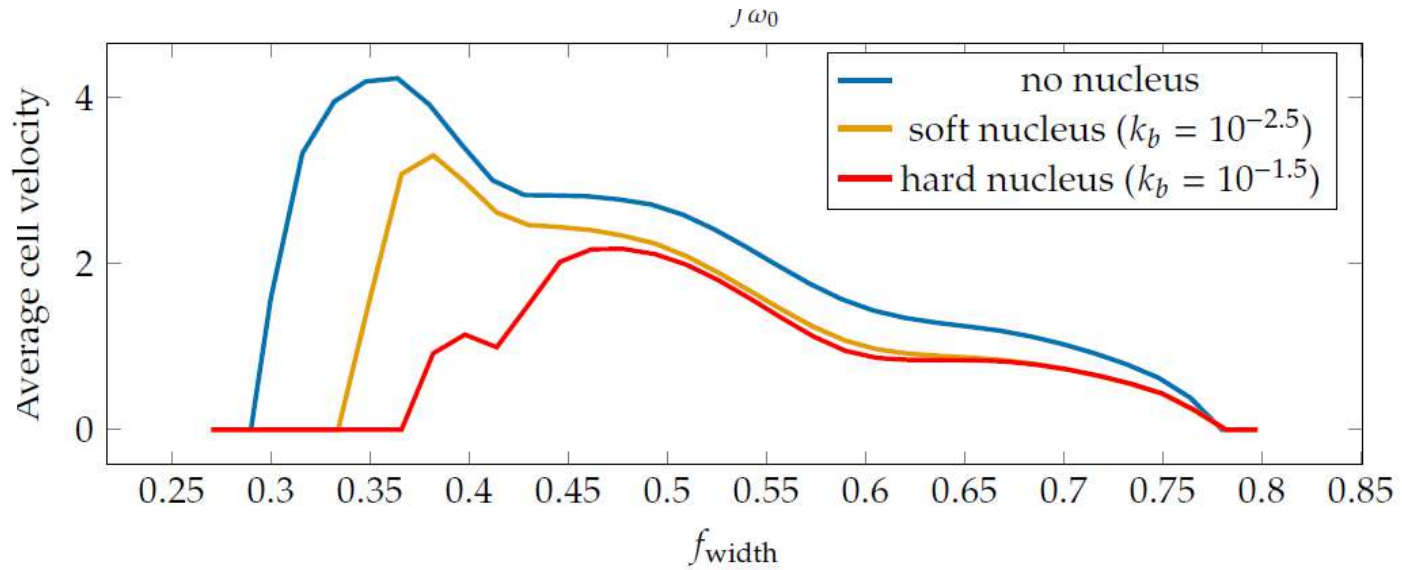
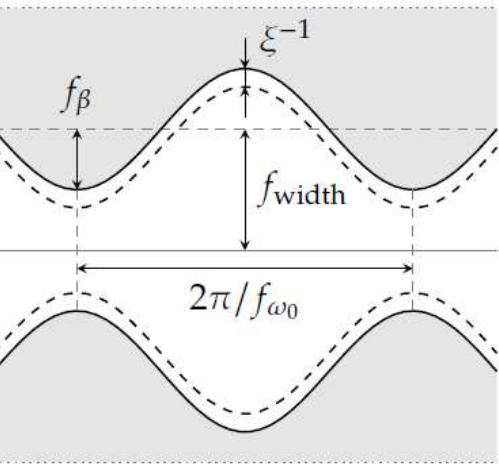




↓
Nucleus stiffness



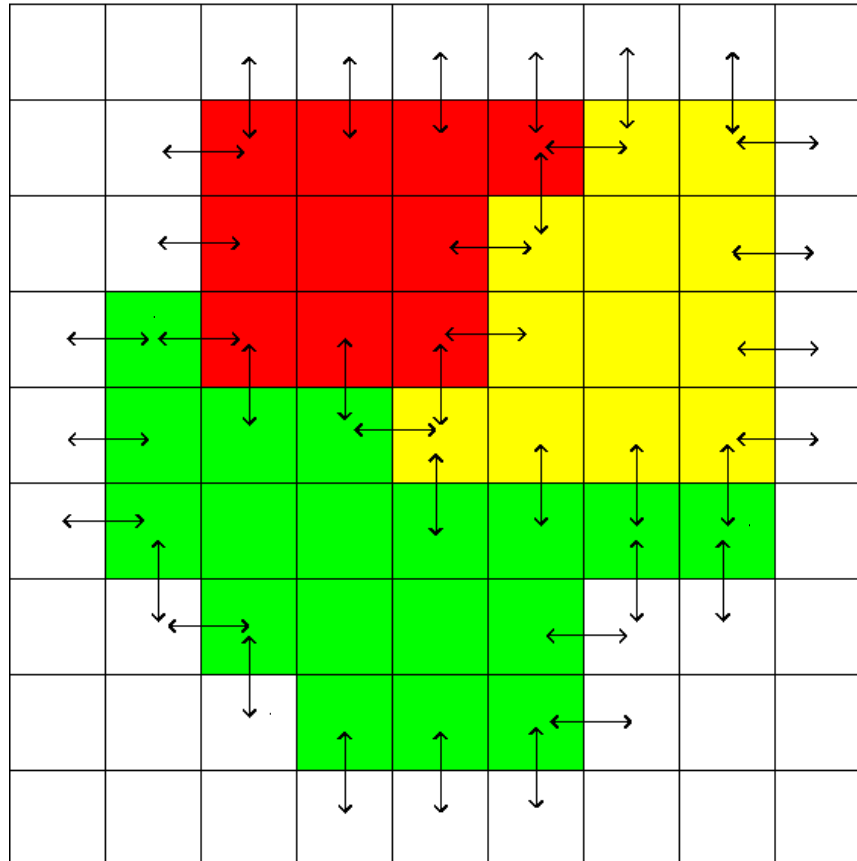
Simulations





The cellular Potts model

A cell is
represented by
several nodes



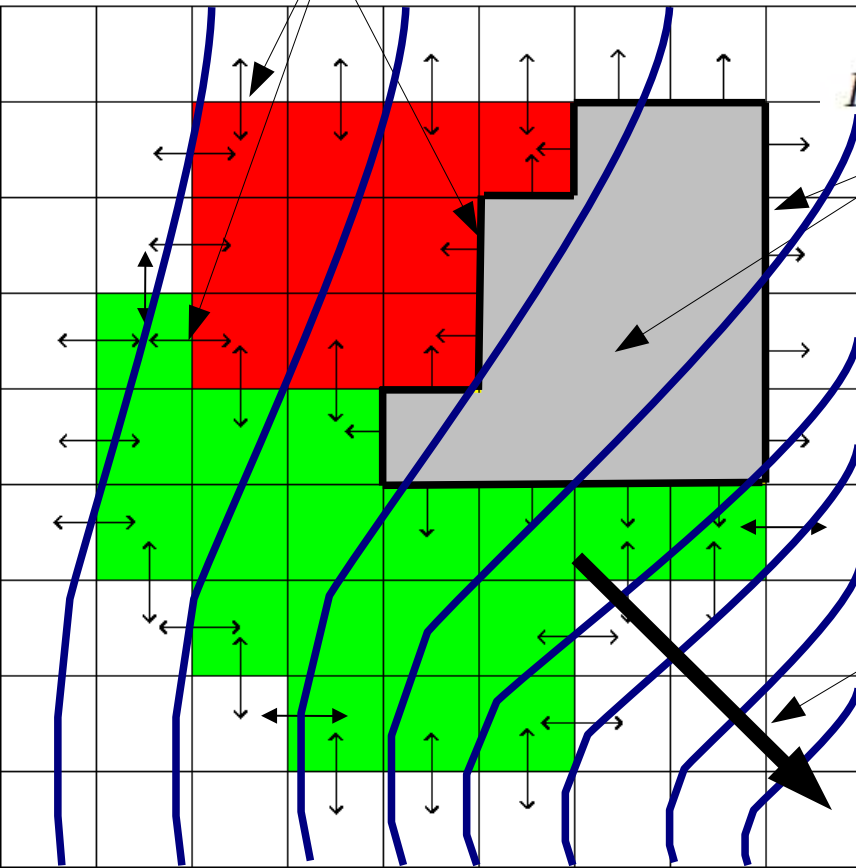
- Based on a generalized energy H
- Evolution stochastically tries to minimize the system energy



The cellular Potts model

$$H(t) = H_{adhesion}(t) + H_{attribute}(t) + H_{force}(t).$$

$$H_{adhesion}(t) = \sum_{\mathbf{x}, \mathbf{x}' \in \Omega} J_{\tau(\sigma(\mathbf{x})), \tau(\sigma(\mathbf{x}'))}(t) [1 - \delta_{\sigma(\mathbf{x}), \sigma(\mathbf{x}')} (t)],$$

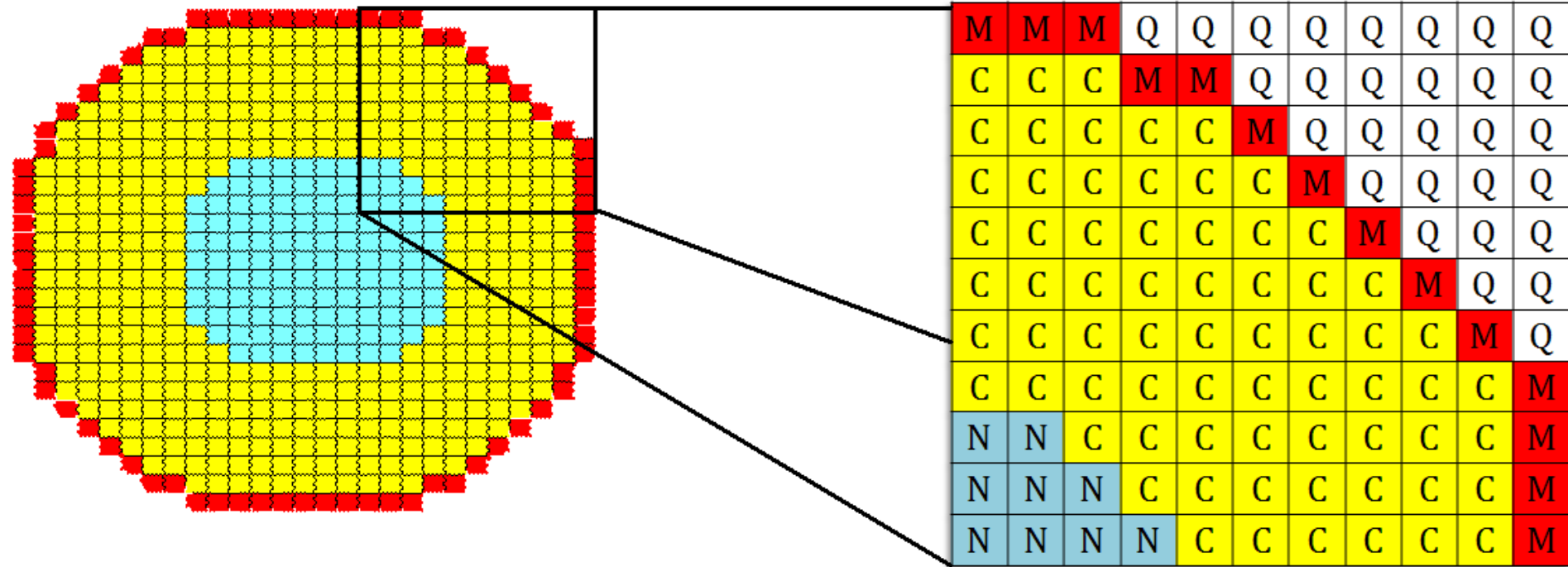


$$H_{attribute}(t) = \sum_{\eta, \sigma, i\text{-attribute}} \lambda_{\eta, \sigma}^i(t) \left| \frac{a_{\eta, \sigma}^i(t) - A_{\eta, \sigma}^i(t)}{a_{\eta, \sigma}^i(t)} \right|^p.$$

$$H_{force}^{chemical}(t) = - \sum_{\sigma} \sum_{\mathbf{x} \in \sigma} \mu_{\sigma}(t) c(\mathbf{x}, t),$$

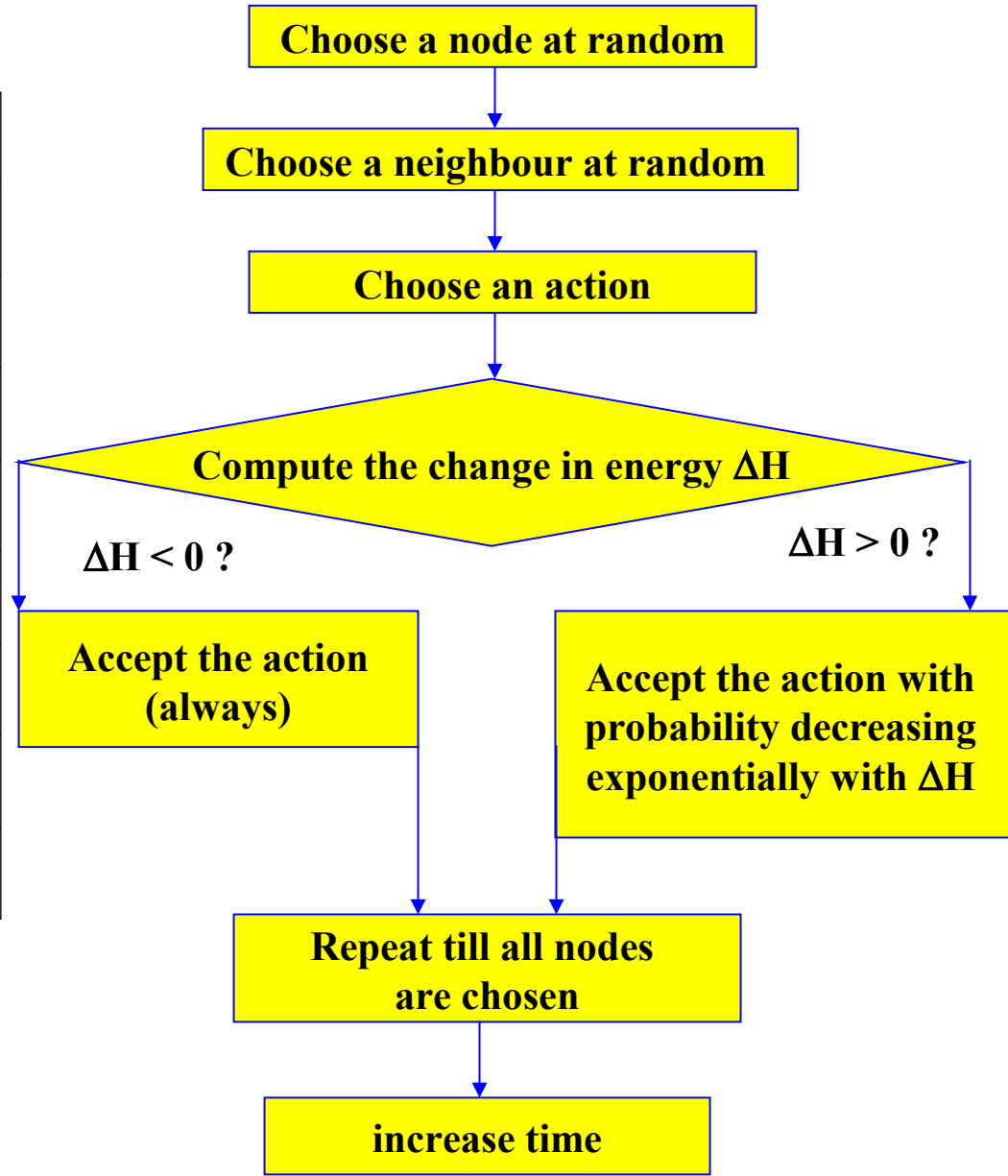
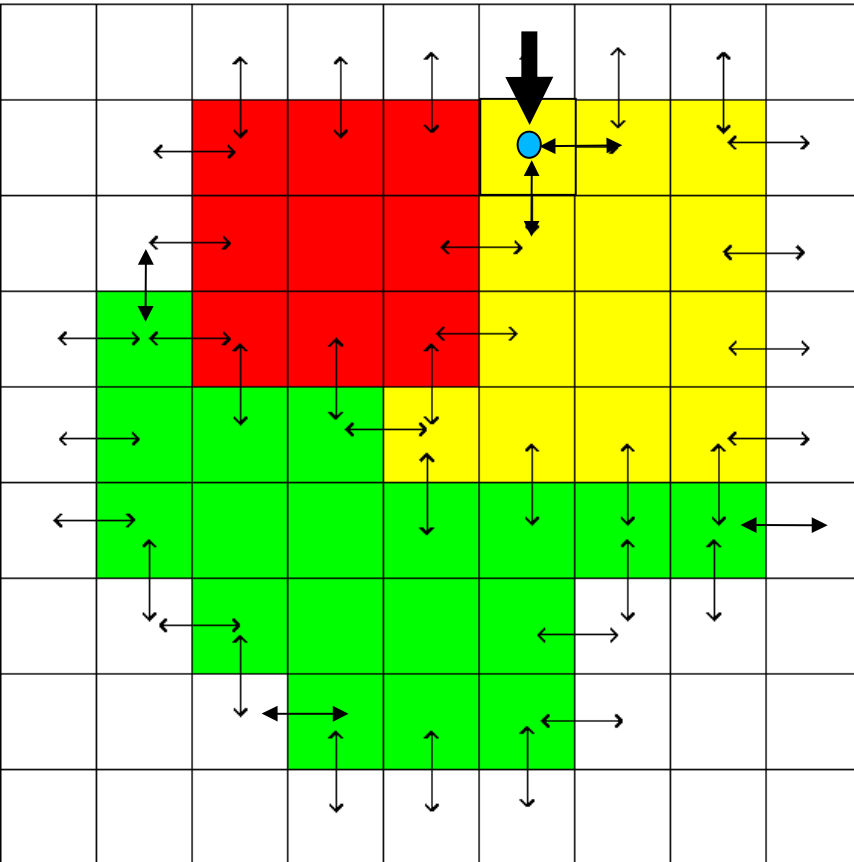
Generalized cellular Potts model

Taking into account of sub-cellular elements (e.g., nucleus)



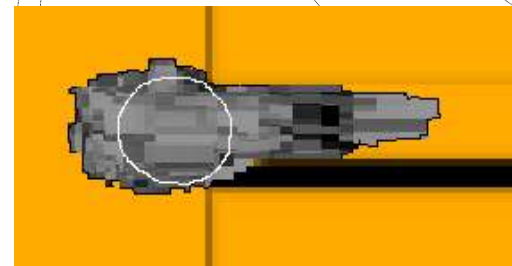
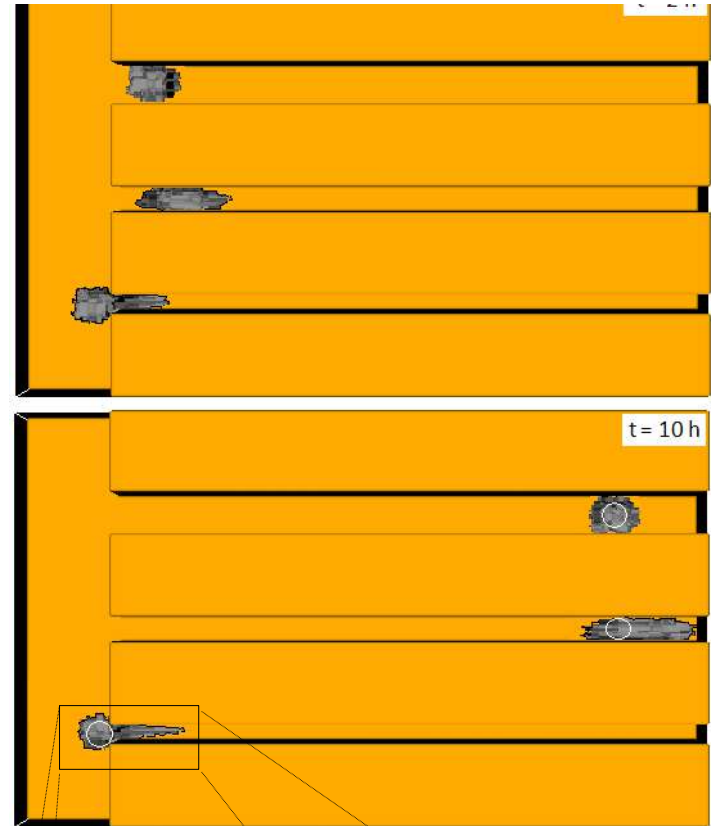
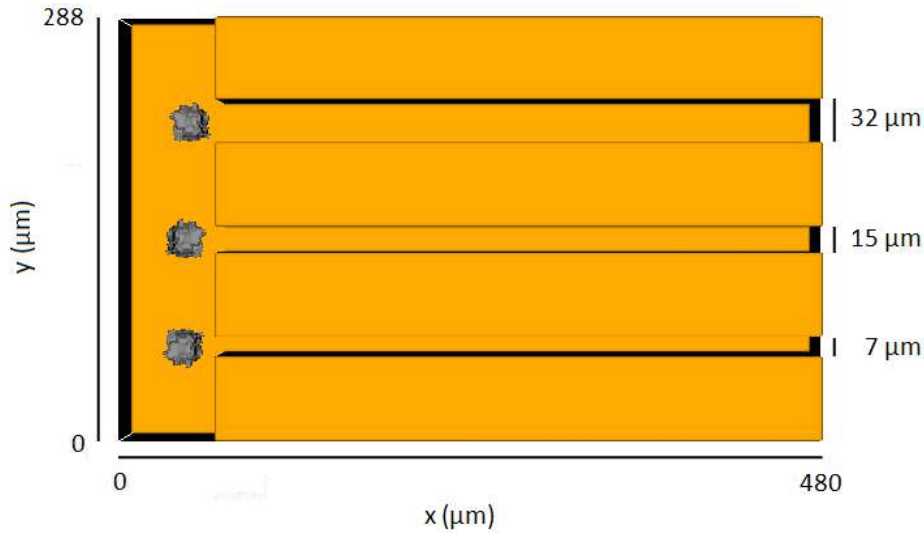


The cellular Potts model





Cells with stiff nuclei in microchannel

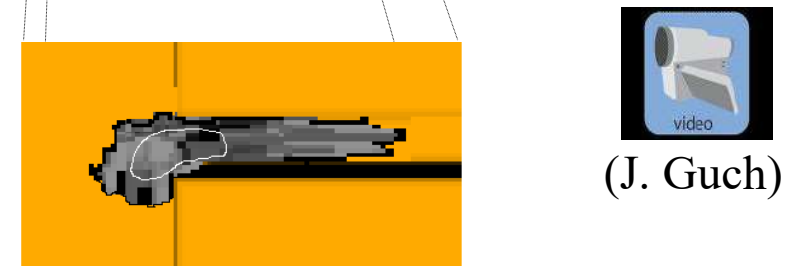
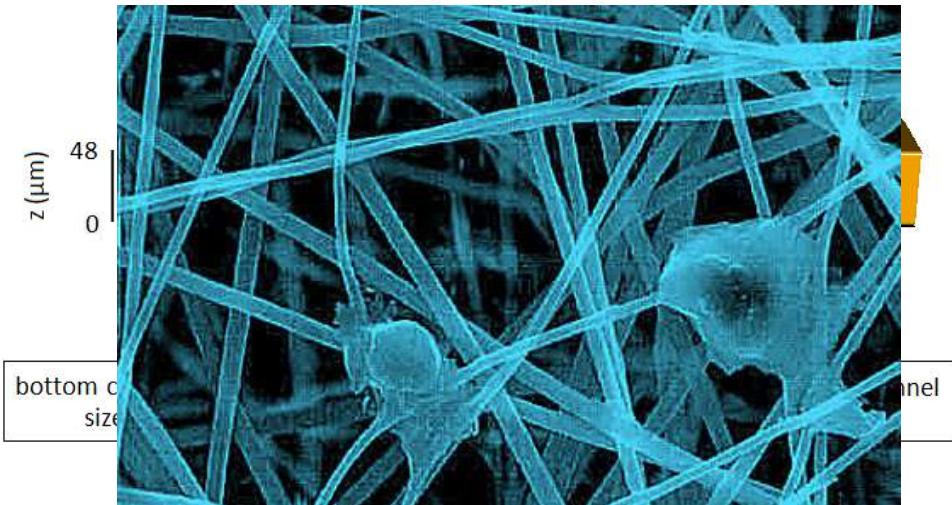
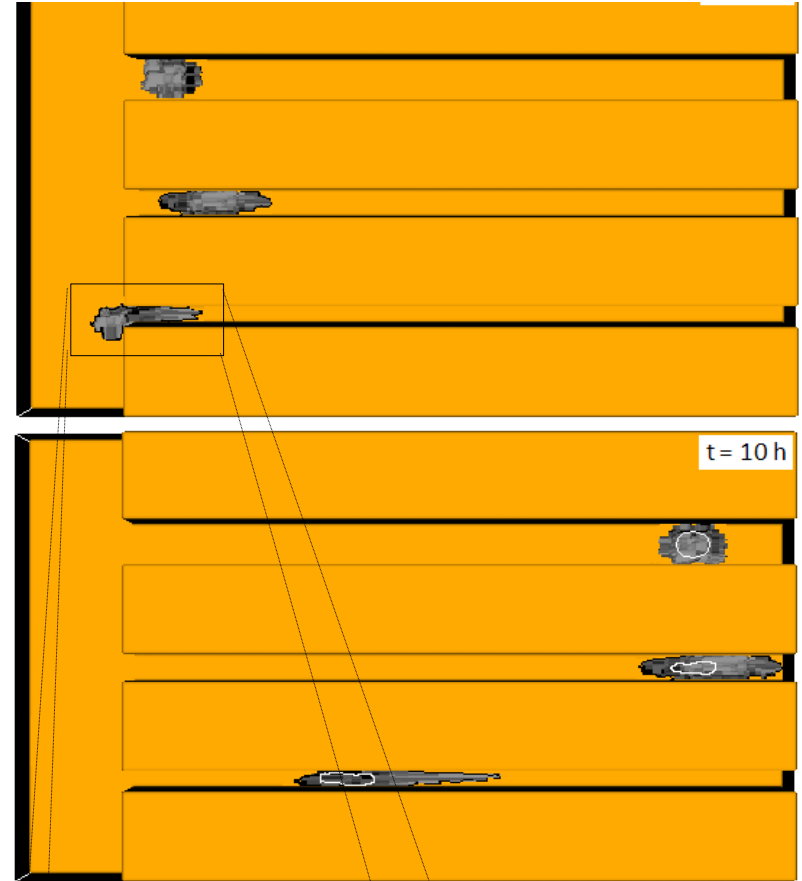
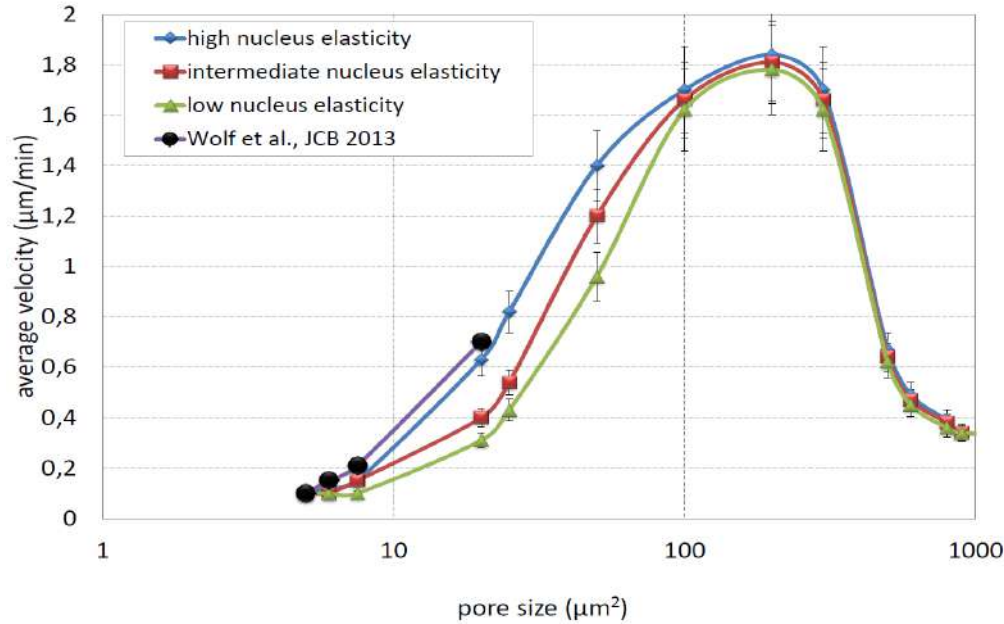


(J. Guch)

bottom channel size < nucleus diameter < middle channel size < cell diameter < top channel size

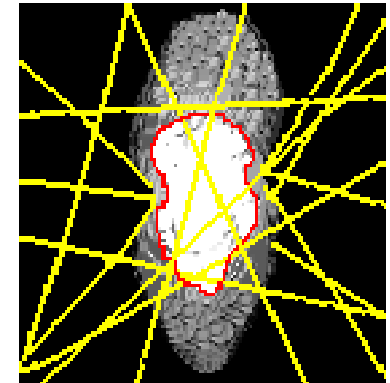
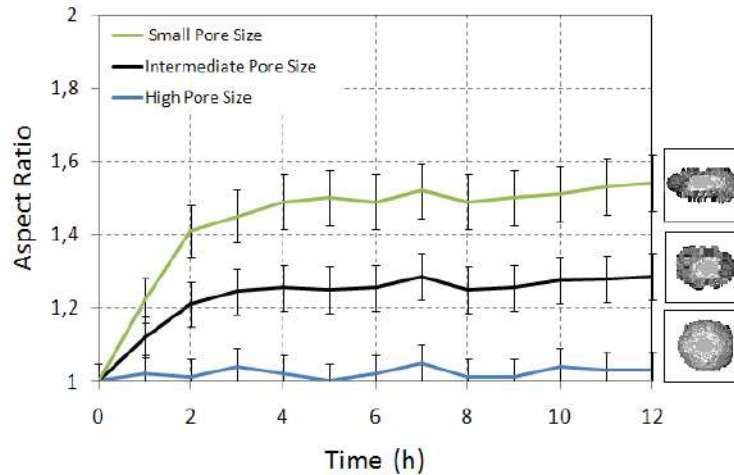
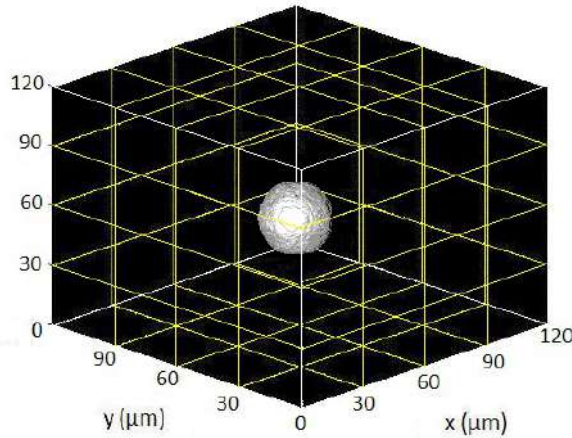
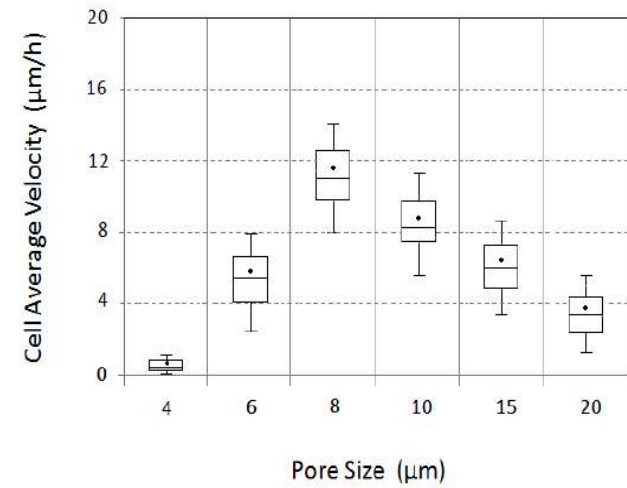
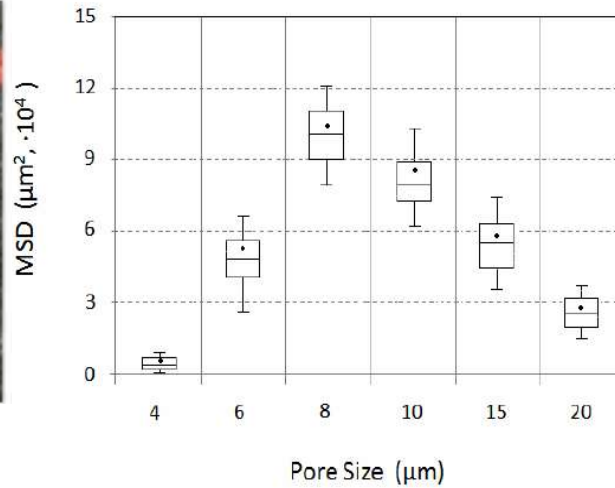
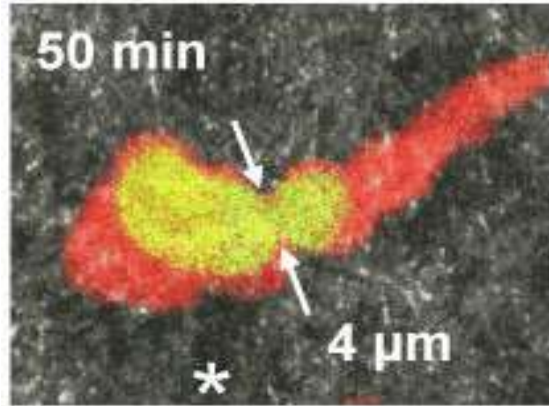


Cells with deformable nuclei in microchannel

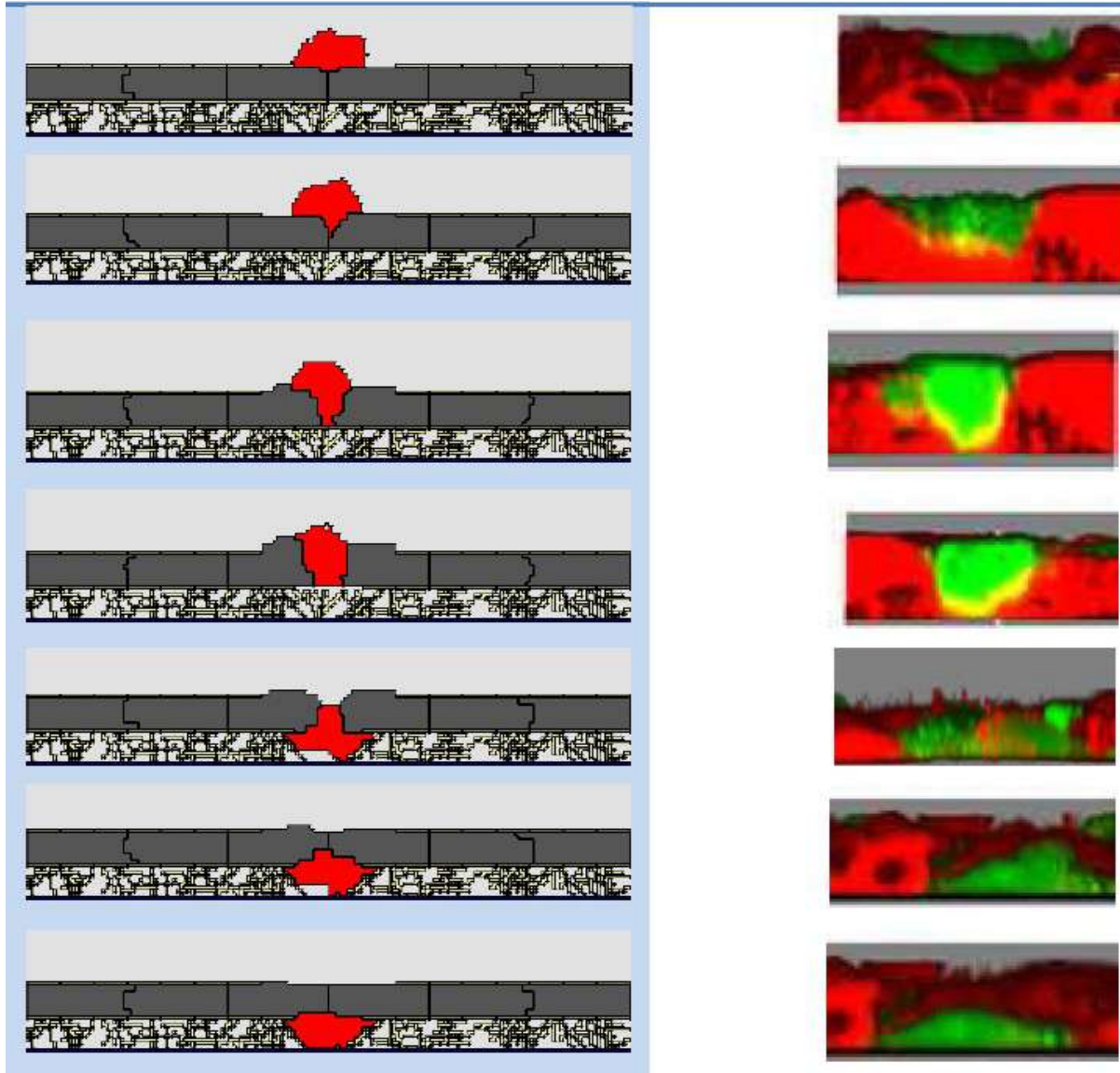


(J. Guch)

Effect of pore size in ECM

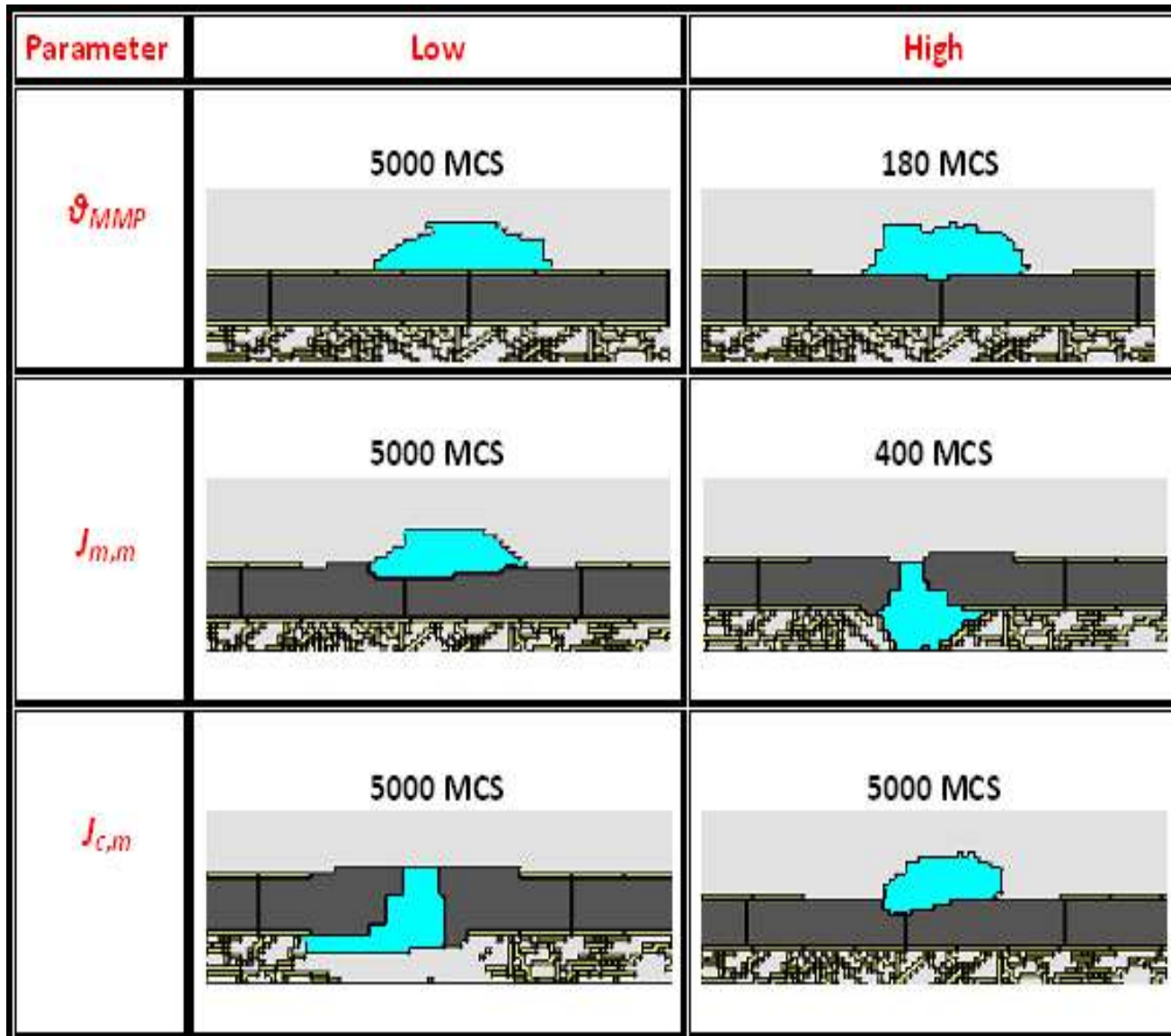


Invasion of single ovary cancer cell

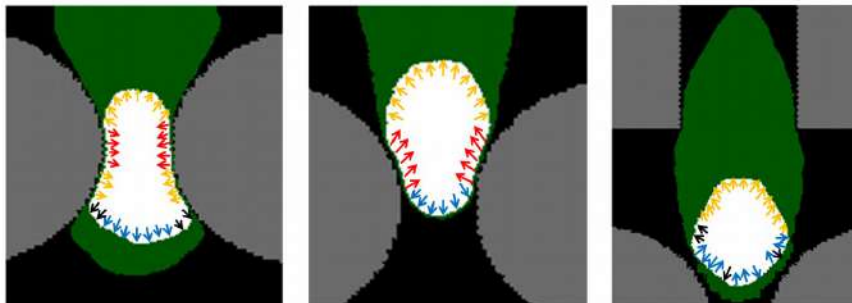
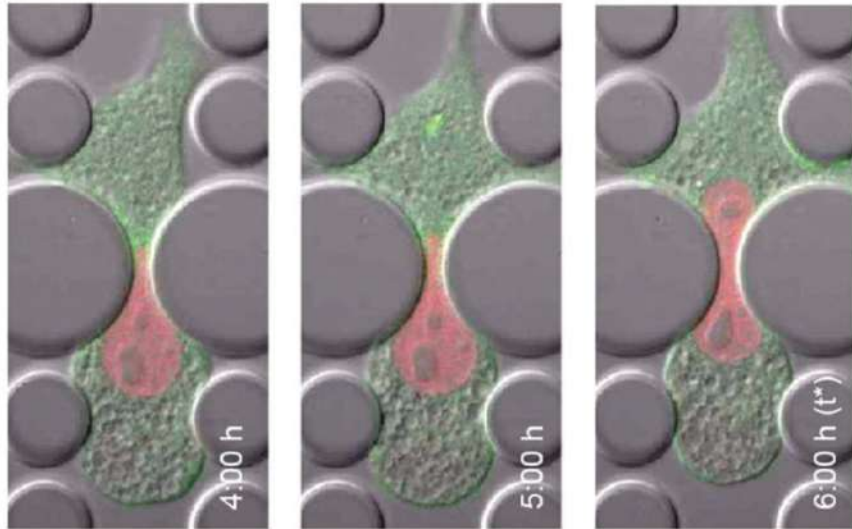




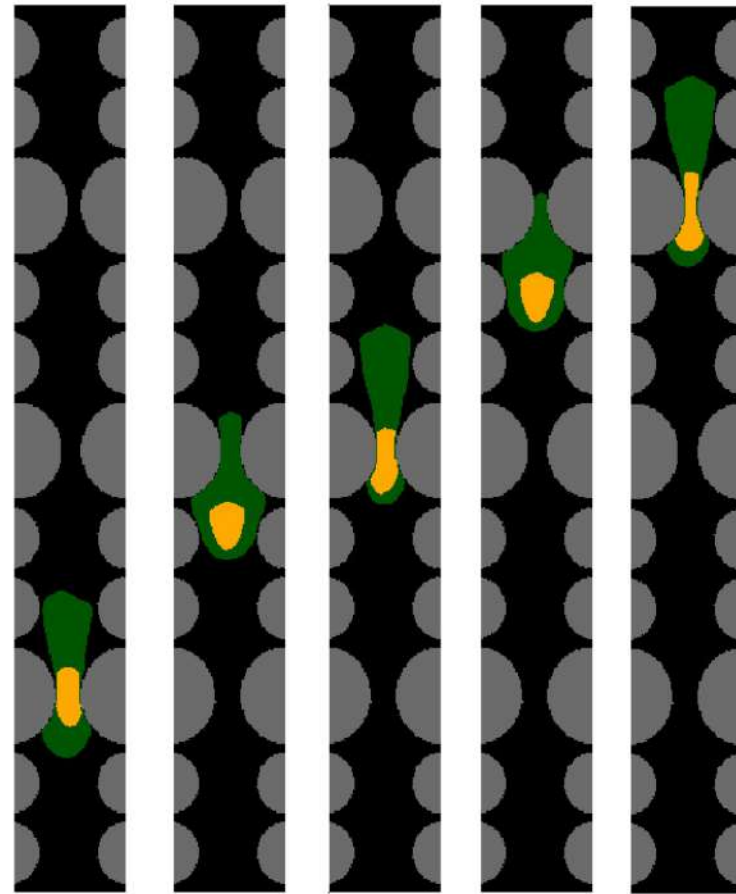
Invasion of single ovary cancer cell



Micropillar arrays



$$\frac{|F(x)|}{|F_{\max}(x)|}$$



3

7

9

14

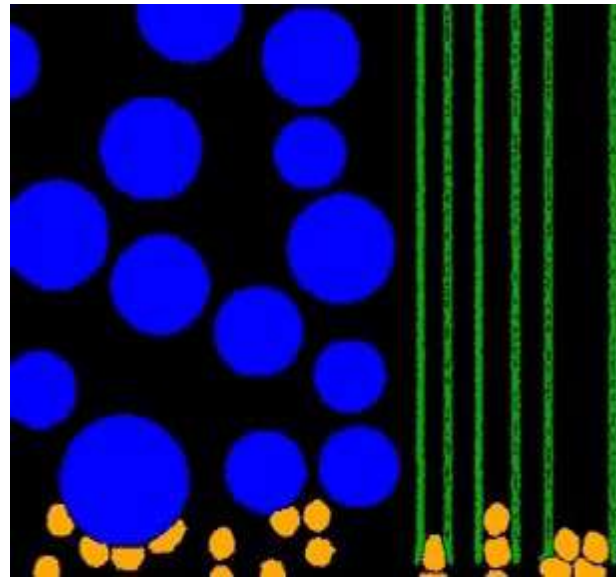
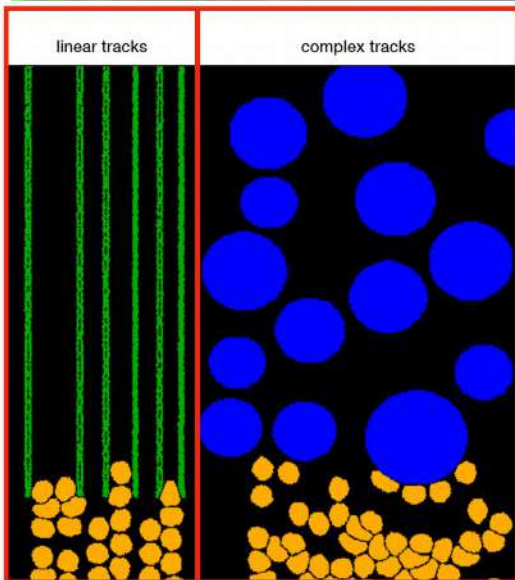
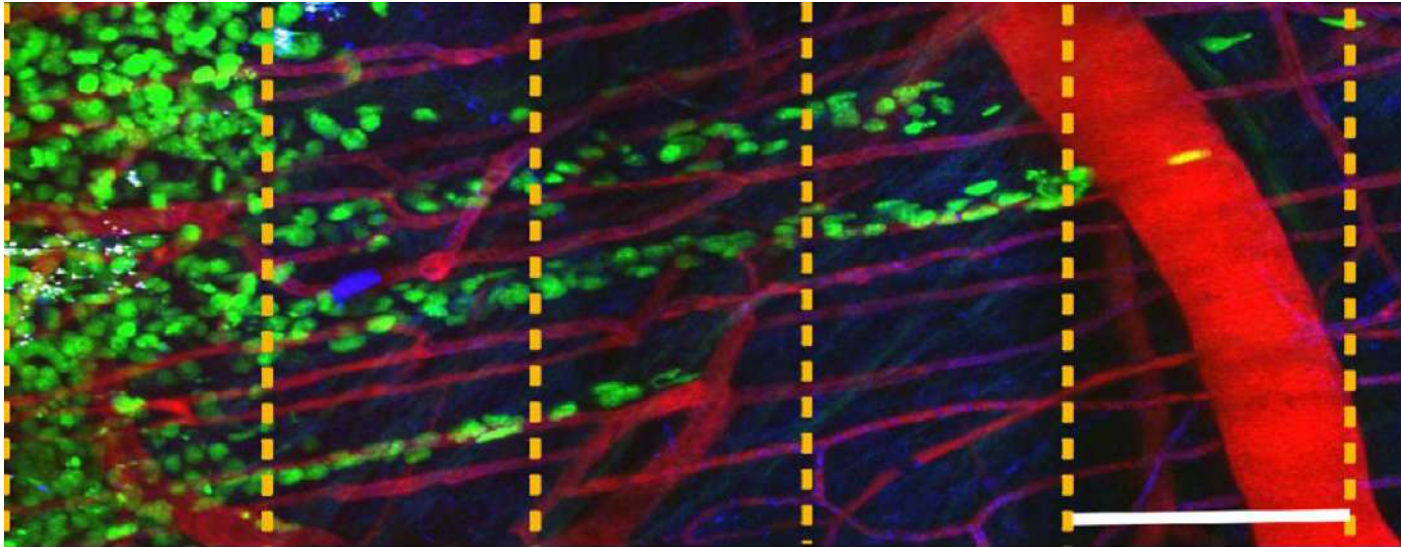
16

time [h]

PDGF



Cell invasion in tissue



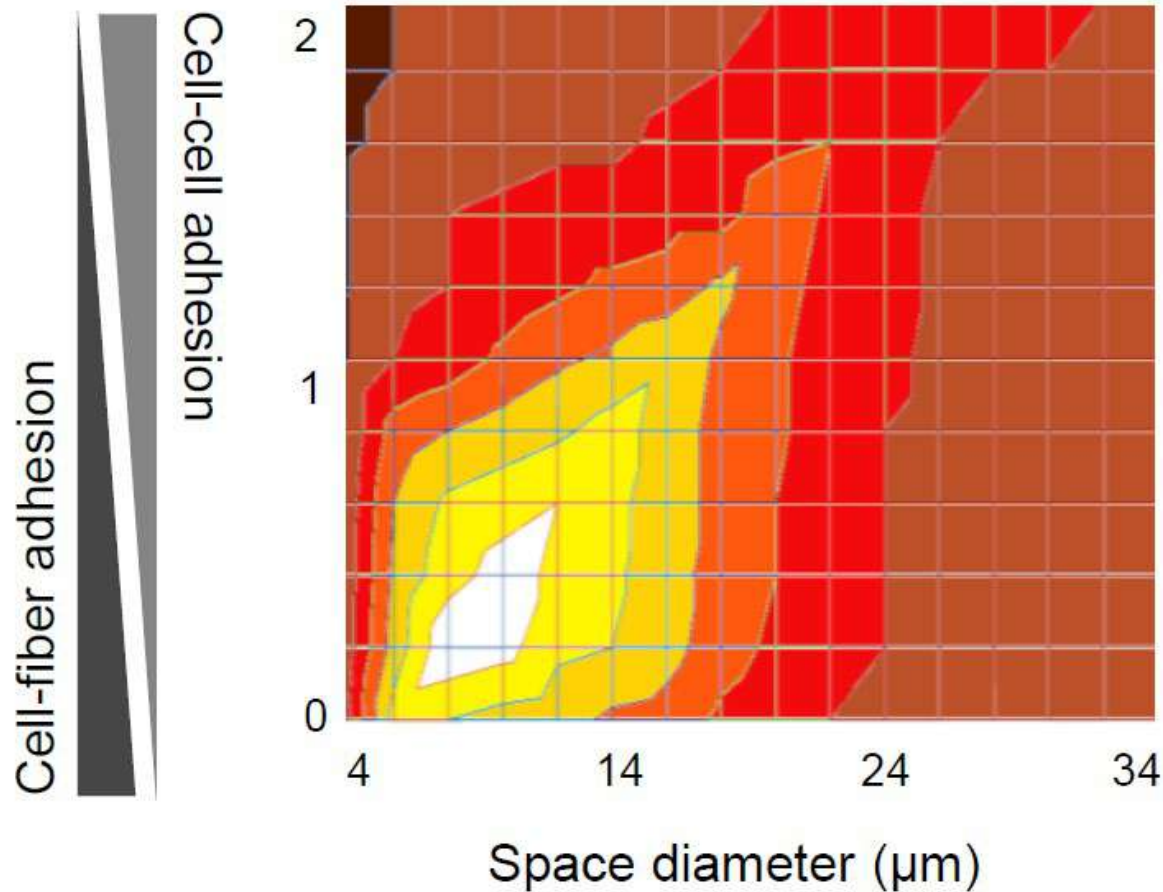
fibre_def_2 (1).mp4



adipociti_2 (1).mp4



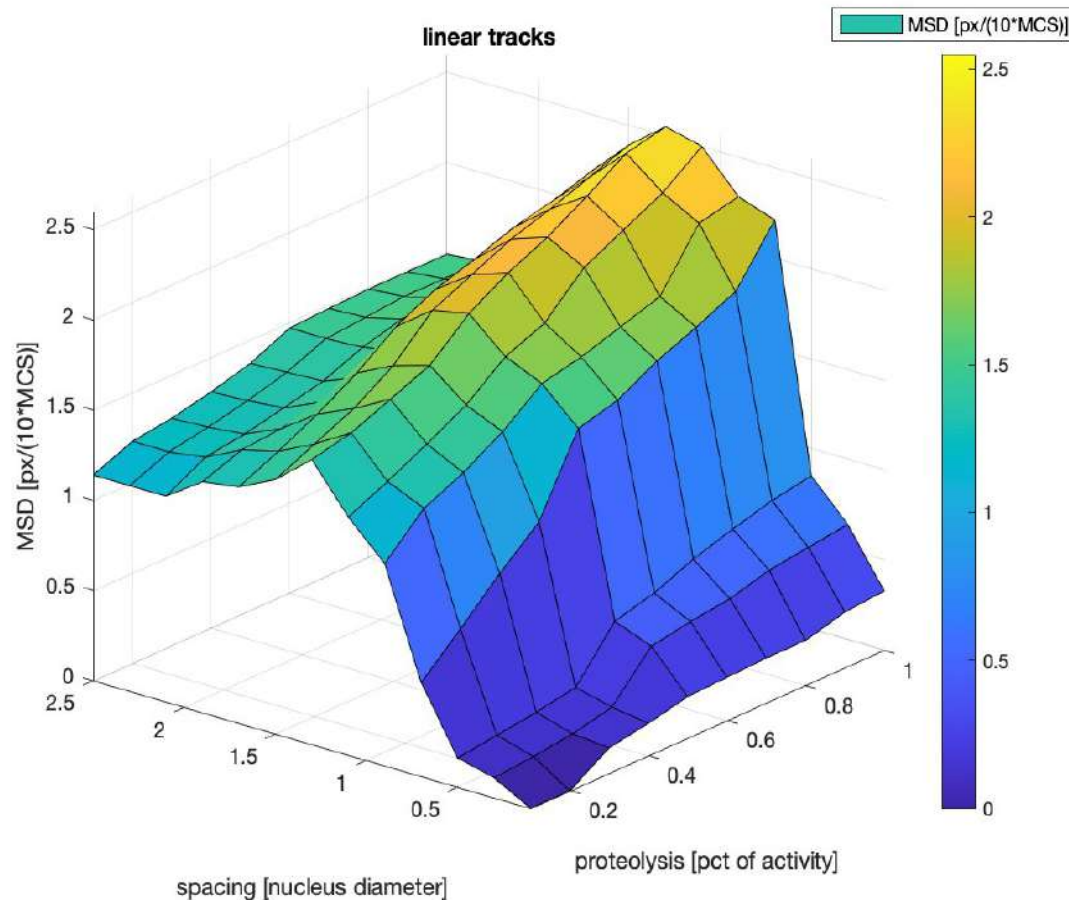
Cell invasion in tissue



Cell directional velocity ($\mu\text{m}/\text{MCS}$)

- 0-0,5
- 0,5-1
- 1-1,5
- 1,5-2
- 2-2,5
- 2,5-3
- 3-3,5

Cell invasion in tissue



Drawbacks:

- Limited number of cells
- Simulation-based statistics

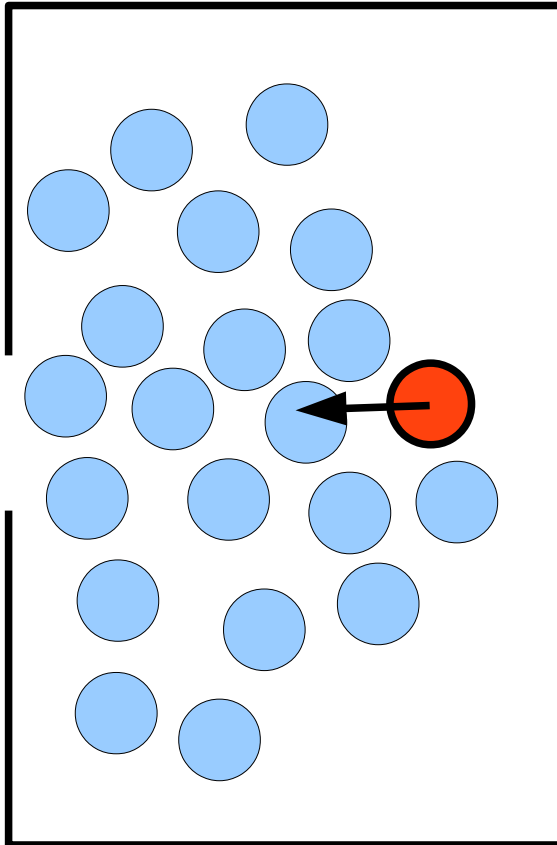
Advantages:

- Easier to insert sub-cellular mechanisms
- Looks closer to reality
- Intrinsic nonlocality



Non local sensing and motion

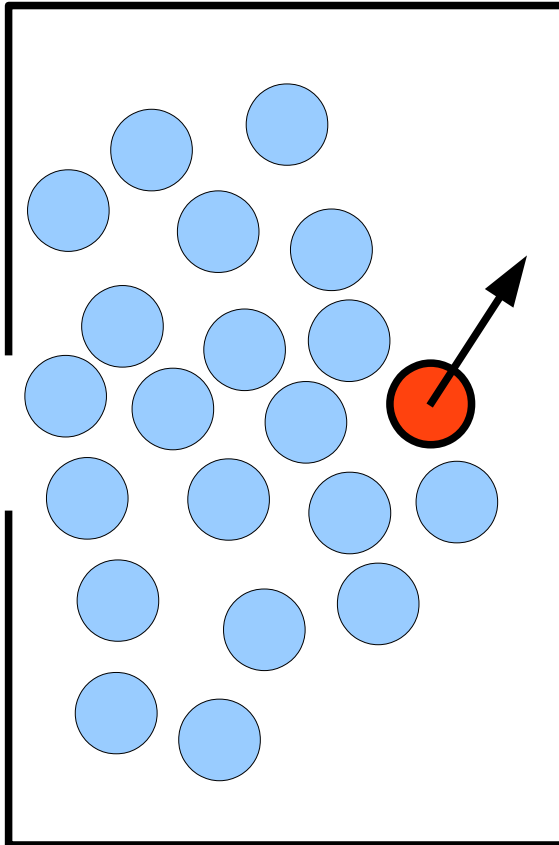
- Sense nonlocally the environment
- Choose where to go
- Move (or try to move)



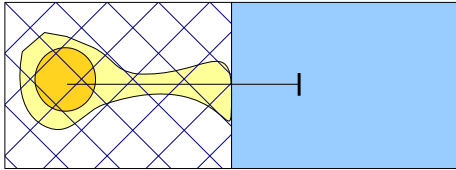


Nonlocal sensing and motion

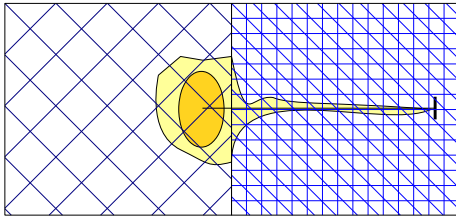
- Sense nonlocally the environment
- Choose where to go
- Move (or try to move)



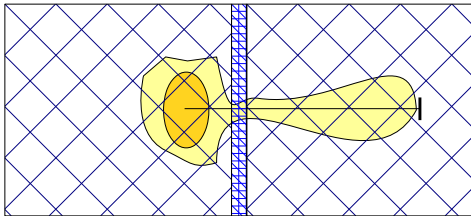
Physical limits of migration



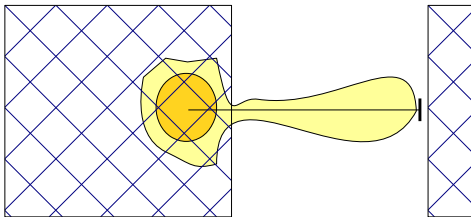
barriers



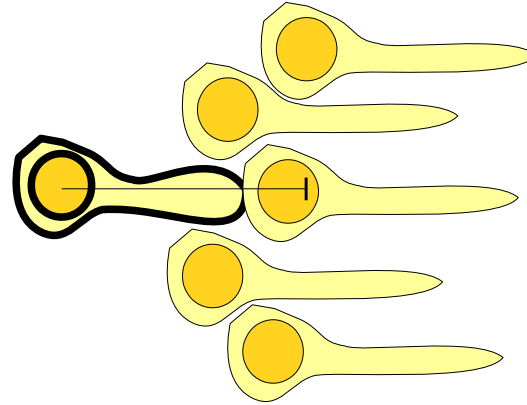
dense
ECM



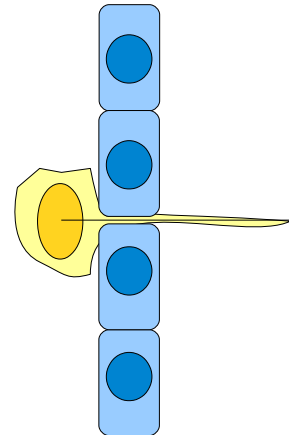
basal
membranes



lack of
adhesion
sites



volume
filling



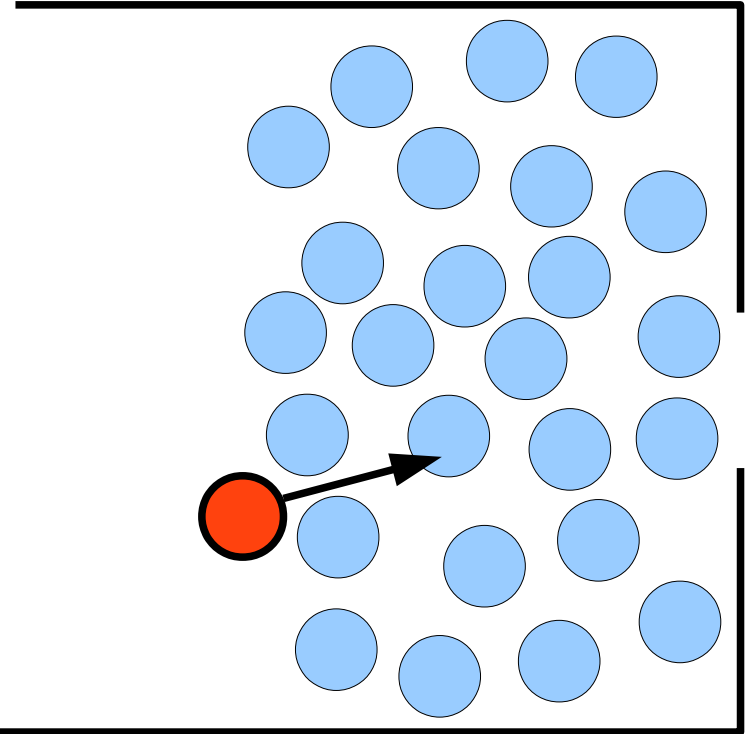
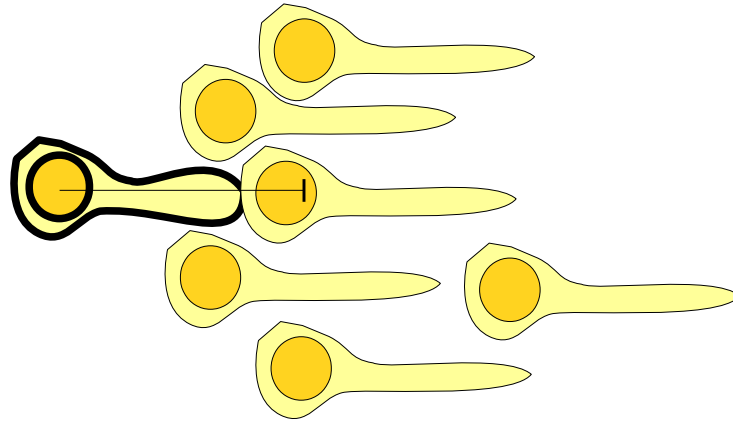
cell layers
epithelial sheets
endothelial walls
capillaries
lymphatics

$$\overline{R_S^M} \quad R_S^{max}$$



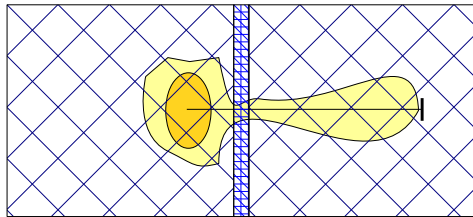
Physical limits of migration

volume
filling

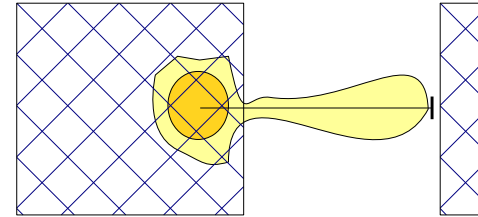




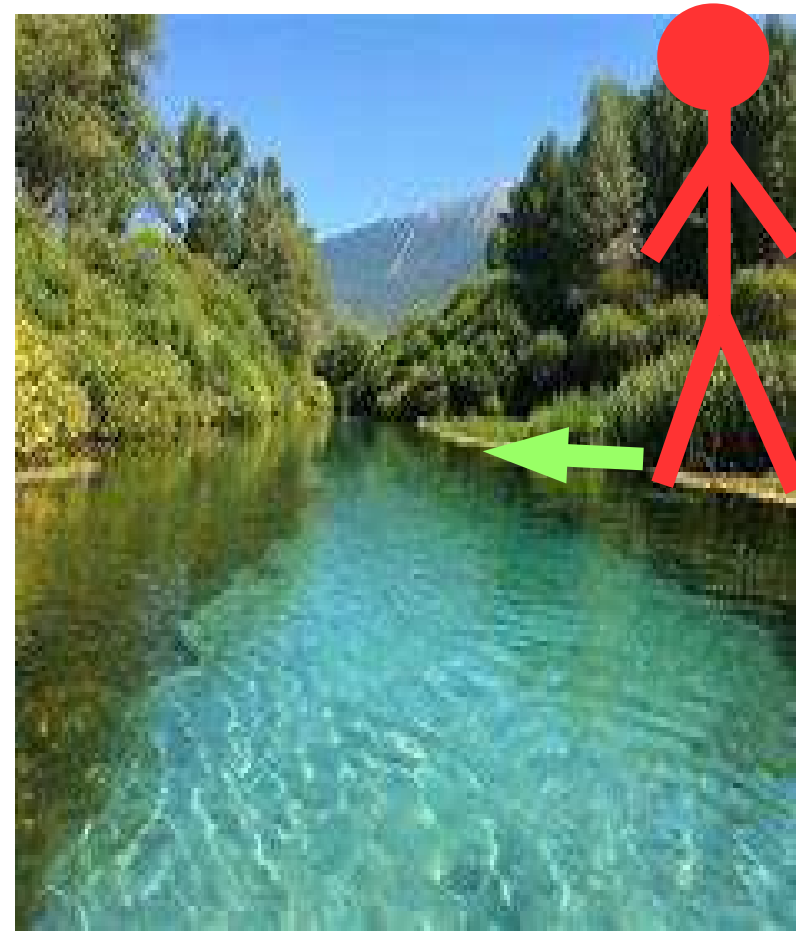
Physical limits of migration



- basal membranes
- cell layers
- intra/extravasation

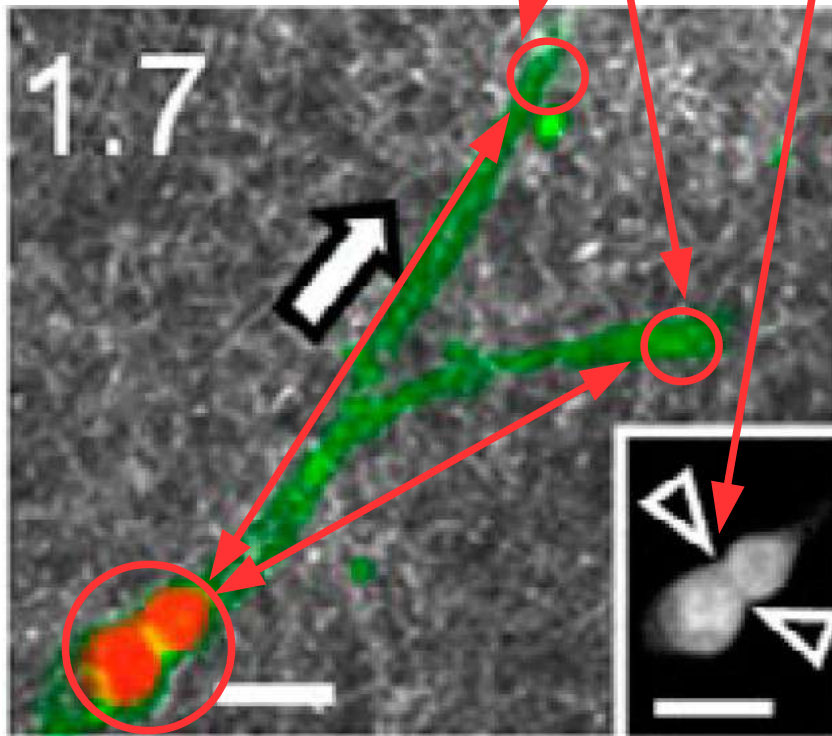


lack of
adhesion
sites

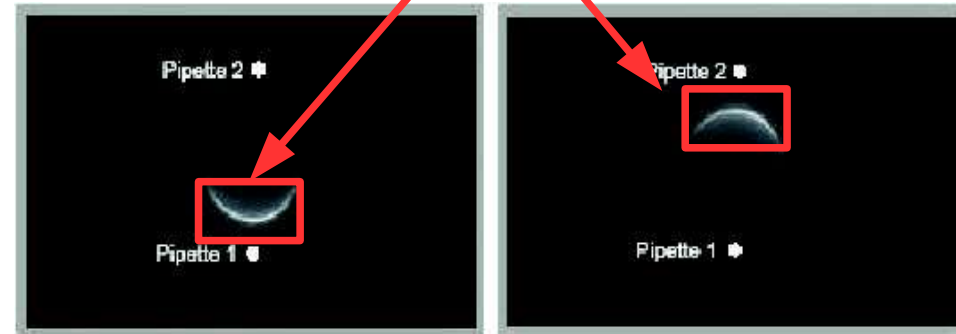


Sensing, polarization and motion

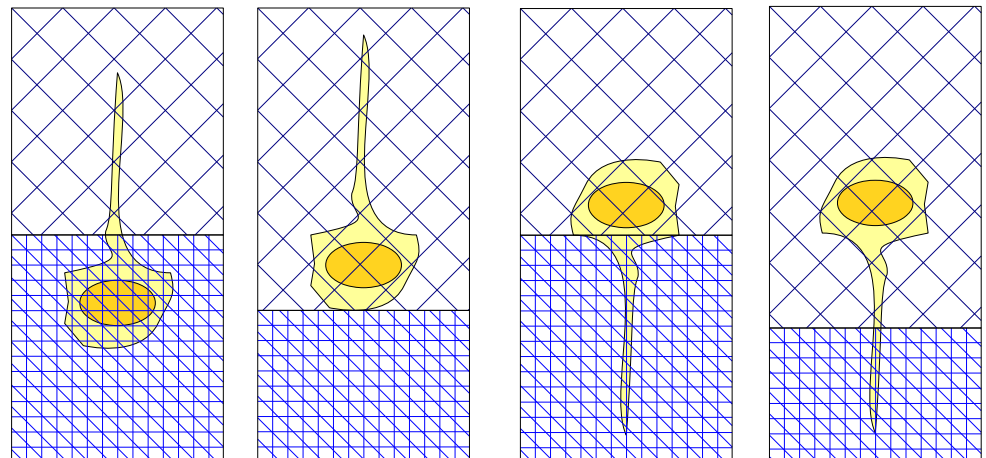
- Non local sensing
- Different cues for polarization and speed
- Physical limits of migration



Wolf, Friedl



Dense ECM





Non local models

- Othmer & Hillen (2002)
- Hillen, Painter & Schmeiser (2006)
- Armstrong, Painter & Sherratt (2007, 2010)

•

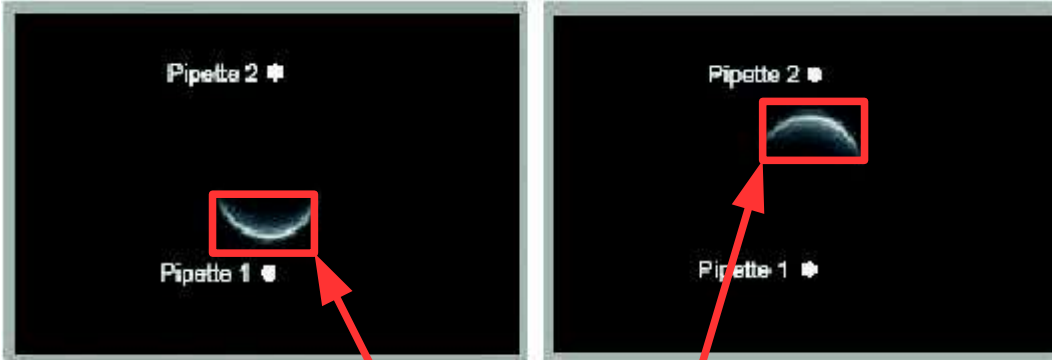
$$\overset{\circ}{\nabla}_{\rho} v(x, t) = \frac{n}{\omega\rho} \int_{S^{n-1}} \sigma v(x + \rho\sigma, t) d\sigma,$$

$$\overset{\circ}{\nabla}_{\rho} v(x, t) = \frac{1}{2\rho} (v(x + \rho, t) - v(x - \rho, t))$$

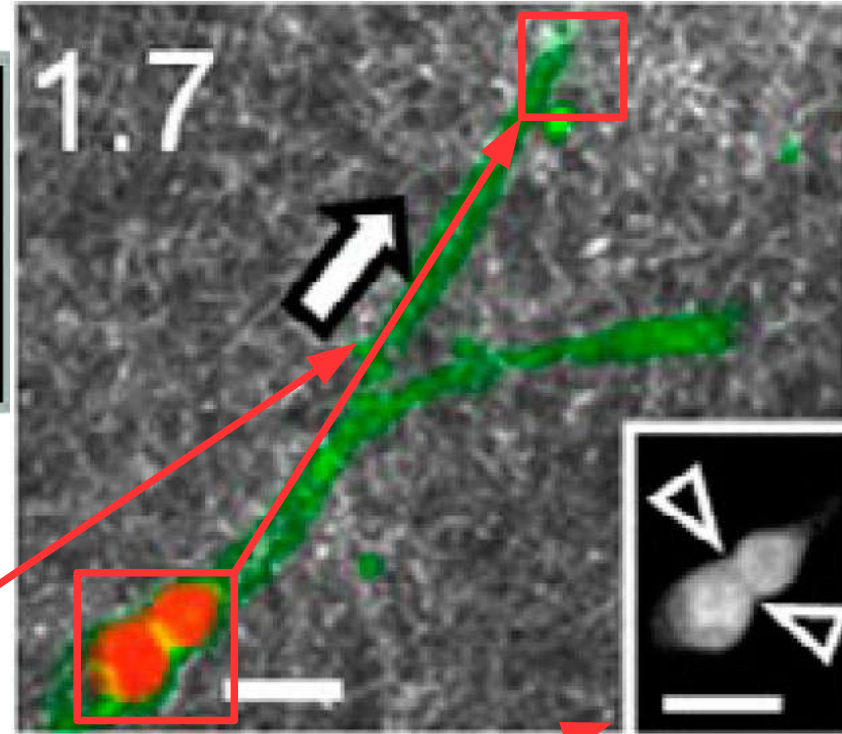
Kinetic model taking into account of

- Different cues for polarization and speed
- Non local sensing
- Physical limits of migration

Sensing, polarization and motion



De vreefotes, Janetopoulos

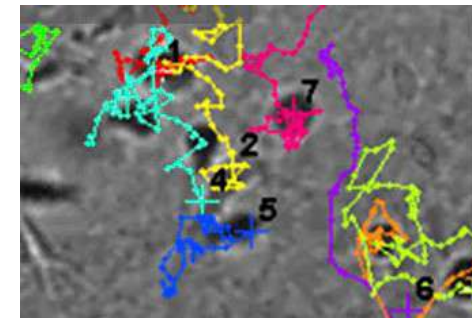


Wolf, Friedl

Summary

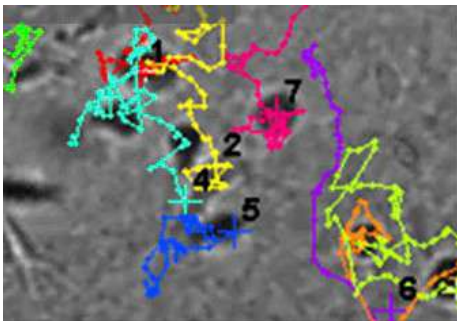
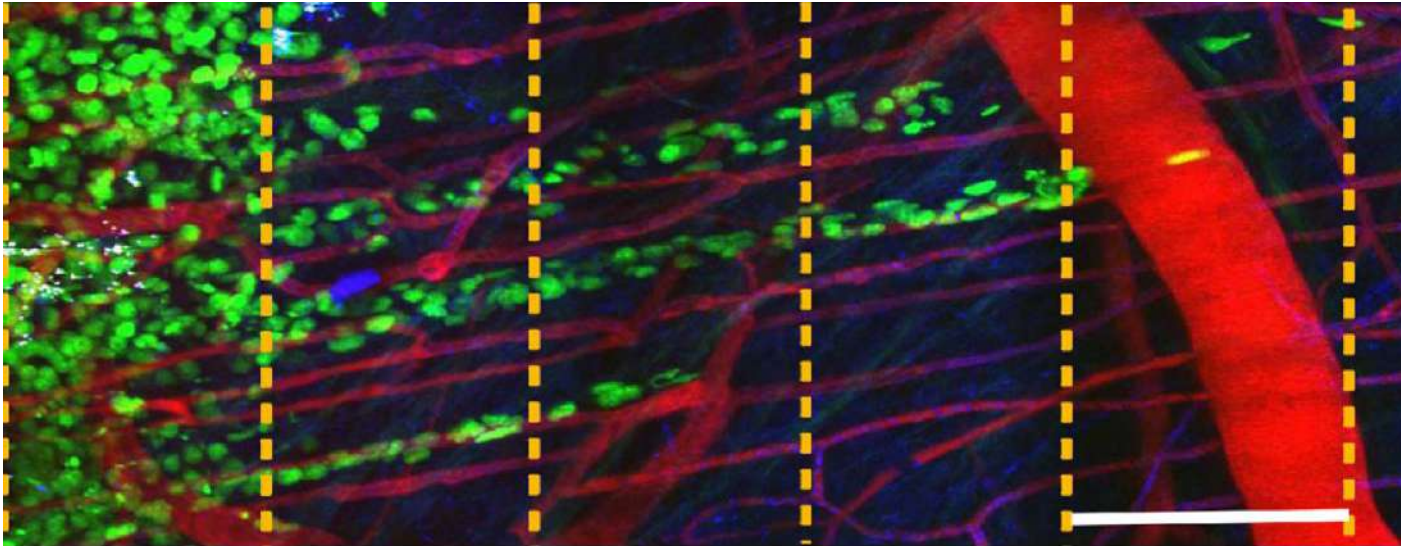
Kinetic model taking into account of

- Different cues for polarization and speed
- Non local sensing
- Physical limits of migration





Aims and summary



(P. Friedl, K. Wolf)



Distribution function

Distribution density for the cell population

$$p = p(t, \mathbf{x}, \mathbf{v}_p) \quad \mathbf{v}_p = (\hat{\mathbf{v}}, v) \in V_p = \mathbb{S}^{d-1} \times [0, U]$$

$$\rho(t, \mathbf{x}) = \int_{V_p} p(t, \mathbf{x}, \mathbf{v}_p) d\mathbf{v}_p$$

$$\mathbf{U}(t, \mathbf{x}) = \frac{1}{\rho(t, \mathbf{x})} \int_{V_p} p(t, \mathbf{x}, \mathbf{v}_p) \mathbf{v} d\mathbf{v}_p$$



Velocity jump process

Distribution density for the cell population

$$p = p(t, \mathbf{x}, \mathbf{v}_p) \quad \mathbf{v}_p = (\hat{\mathbf{v}}, v) \in V_p = \mathbb{S}^{d-1} \times [0, U]$$

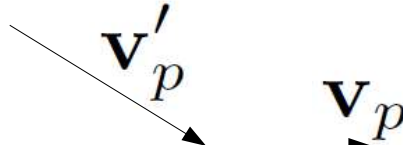
$$\frac{\partial p}{\partial t}(t, \mathbf{x}, \mathbf{v}_p) + \mathbf{v} \cdot \nabla p(t, \mathbf{x}, \mathbf{v}_p) = \mathcal{J}[p](t, \mathbf{x}, \mathbf{v}_p)$$

$$\mathcal{J}[p](\mathbf{x}, \mathbf{v}_p) = \mathcal{G}[p](\mathbf{x}, \mathbf{v}_p) - \mathcal{L}[p](\mathbf{x}, \mathbf{v}_p)$$



Velocity jump process

$$\mathcal{J}[p](\mathbf{x}, \mathbf{v}_p) = \mathcal{G}[p](\mathbf{x}, \mathbf{v}_p) - \mathcal{L}[p](\mathbf{x}, \mathbf{v}_p)$$

$$\mathcal{G}[p](\mathbf{x}, \mathbf{v}_p) = \int_{V_p} \overset{\text{Turning rate}}{\mu(\mathbf{x}, \mathbf{v}'_p)} \overset{\text{Turning frequency}}{T[\mathcal{S}, \mathcal{S}']}(\mathbf{x}, \mathbf{v}_p | \mathbf{v}'_p) p(t, \mathbf{x}, \mathbf{v}'_p) d\mathbf{v}'_p$$




Velocity jump process

$$\mathcal{J}[p](\mathbf{x}, \mathbf{v}_p) = \mathcal{G}[p](\mathbf{x}, \mathbf{v}_p) - \mathcal{L}[p](\mathbf{x}, \mathbf{v}_p)$$

$$\mathcal{G}[p](\mathbf{x}, \mathbf{v}_p) = \int_{V_p} \mu(\mathbf{x}, \mathbf{v}'_p) \overset{\text{Turning rate}}{T[\mathcal{S}, \mathcal{S}']}(\mathbf{x}, \mathbf{v}_p | \mathbf{v}'_p) \overset{\text{Turning frequency}}{p}(t, \mathbf{x}, \mathbf{v}'_p) d\mathbf{v}'_p$$

$$\mathcal{L}[p](\mathbf{x}, \mathbf{v}_p) = \int_{V_p} \mu(\mathbf{x}, \mathbf{v}_p) T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, \mathbf{v}''_p | \mathbf{v}_p) p(t, \mathbf{x}, \mathbf{v}_p) d\mathbf{v}''_p$$



Velocity jump process

T is a transition probability $\int_{V_p} T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, \mathbf{v}_p'' | \mathbf{v}_p) d\mathbf{v}_p'' = 1$

$$\mathcal{L}[p](\mathbf{x}, \mathbf{v}_p) = \int_{V_p} \mu(\mathbf{x}, \mathbf{v}_p) T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, \mathbf{v}_p'' | \mathbf{v}_p) p(t, \mathbf{x}, \mathbf{v}_p) d\mathbf{v}_p''$$



Velocity jump process

T is a transition probability $\int_{V_p} T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, \mathbf{v}_p'' | \mathbf{v}_p) d\mathbf{v}_p'' = 1$

$$\mathcal{L}[p](\mathbf{x}, \mathbf{v}_p) = \mu(\mathbf{x}, \mathbf{v}_p)p(t, \mathbf{x}, \mathbf{v}_p)$$

$$\begin{aligned} \frac{\partial p}{\partial t}(t, \mathbf{x}, \mathbf{v}_p) + \mathbf{v} \cdot \nabla p(t, \mathbf{x}, \mathbf{v}_p) \\ = \int_{V_p} \mu(\mathbf{x}, \mathbf{v}_p') T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, \mathbf{v}_p | \mathbf{v}_p') p(t, \mathbf{x}, \mathbf{v}_p') d\mathbf{v}_p' \\ - \mu(\mathbf{x}, \mathbf{v}_p) p(t, \mathbf{x}, \mathbf{v}_p) \end{aligned}$$



Mass conservation

Integrate over the velocity space

$$\begin{aligned} \int_{V_p} \frac{\partial p}{\partial t}(t, \mathbf{x}, \mathbf{v}_p) + \mathbf{v} \cdot \nabla p(t, \mathbf{x}, \mathbf{v}_p) d\mathbf{v}_p \\ = \int_{V_p} \int_{V_p} \mu(\mathbf{x}, \mathbf{v}'_p) T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, \mathbf{v}_p | \mathbf{v}'_p) p(t, \mathbf{x}, \mathbf{v}'_p) d\mathbf{v}'_p \\ - \mu(\mathbf{x}, \mathbf{v}_p) p(t, \mathbf{x}, \mathbf{v}_p) d\mathbf{v}_p \end{aligned}$$



Mass conservation

Integrate over the velocity space

$$\int_{V_p} \frac{\partial p}{\partial t}(t, \mathbf{x}, \mathbf{v}_p) + \mathbf{v} \cdot \nabla p(t, \mathbf{x}, \mathbf{v}_p) d\mathbf{v}_p$$
$$= \int_{V_p} \mu(\mathbf{x}, \mathbf{v}'_p) p(t, \mathbf{x}, \mathbf{v}'_p) d\mathbf{v}'_p$$
$$- \int_{V_p} \mu(\mathbf{x}, \mathbf{v}_p) p(t, \mathbf{x}, \mathbf{v}_p) d\mathbf{v}_p$$



Turning kernel

μ and T independent from the pre-tumbling velocity

$$\begin{aligned} \frac{\partial p}{\partial t}(t, \mathbf{x}, \mathbf{v}_p) + \mathbf{v} \cdot \nabla p(t, \mathbf{x}, \mathbf{v}_p) \\ = \int_{V_p} \mu(\mathbf{x}, \cancel{\mathbf{v}}_p) T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, \mathbf{v}_p | \cancel{\mathbf{v}}'_p) p(t, \mathbf{x}, \mathbf{v}'_p) d\mathbf{v}'_p \\ - \mu(\mathbf{x}, \cancel{\mathbf{v}}_p) p(t, \mathbf{x}, \mathbf{v}_p) \end{aligned}$$

$$\longrightarrow \mathcal{J}[p](t, \mathbf{x}, v, \hat{\mathbf{v}}) = \mu(\mathbf{x}) \left(\rho(t, \mathbf{x}) T(\mathbf{x}, v, \hat{\mathbf{v}}) - p(t, \mathbf{x}, v, \hat{\mathbf{v}}) \right)$$



Transport equation

$$p(t, \mathbf{x}, v, \hat{\mathbf{v}}) \quad \text{s.t.} \quad \rho(t, \mathbf{x}) = \int_{V_p} p(t, \mathbf{x}, v, \hat{\mathbf{v}}) dv d\hat{\mathbf{v}}$$

$$\frac{\partial p}{\partial t}(t, \mathbf{x}, v, \hat{\mathbf{v}}) + v \hat{\mathbf{v}} \cdot \nabla p(t, \mathbf{x}, v, \hat{\mathbf{v}}) = \mathcal{J}[p](t, \mathbf{x}, v, \hat{\mathbf{v}})$$

$$\mathcal{J}[p](t, \mathbf{x}, v, \hat{\mathbf{v}}) = \int_{V_p} \overset{\text{Turning rate}}{\mu(\mathbf{x}, v', \hat{\mathbf{v}}')} T(\mathbf{x}, v, \hat{\mathbf{v}} | v', \hat{\mathbf{v}}') p(t, \mathbf{x}, v', \hat{\mathbf{v}}') d'v d'\hat{\mathbf{v}} - \mu(\mathbf{x}, v, \hat{\mathbf{v}}) p(t, \mathbf{x}, v, \hat{\mathbf{v}})$$



Turning kernel

$$\frac{\partial p}{\partial t}(t, \mathbf{x}, v, \hat{\mathbf{v}}) + v \hat{\mathbf{v}} \cdot \nabla p(t, \mathbf{x}, v, \hat{\mathbf{v}}) = \mathcal{J}[p](t, \mathbf{x}, v, \hat{\mathbf{v}})$$

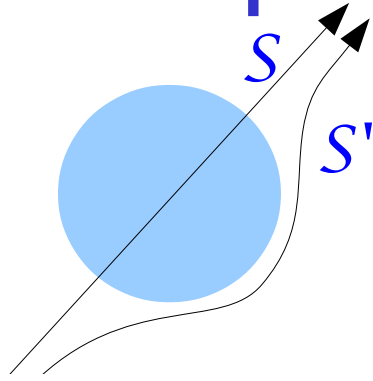
μ and T independent from the pre-tumbling velocity

$$\begin{aligned} \mathcal{J}[p](t, \mathbf{x}, v, \hat{\mathbf{v}}) = & \int_{V_p} \mu(\mathbf{x}, \cancel{v}, \cancel{\hat{\mathbf{v}}}) T(\mathbf{x}, v, \hat{\mathbf{v}} | \cancel{v}, \cancel{\hat{\mathbf{v}}}) p(t, \mathbf{x}, \cancel{v}, \cancel{\hat{\mathbf{v}}}) d'v d'\hat{\mathbf{v}} \\ & - \mu(\mathbf{x}, \cancel{v}, \cancel{\hat{\mathbf{v}}}) p(t, \mathbf{x}, v, \hat{\mathbf{v}}) \end{aligned}$$

$$\longrightarrow \mathcal{J}[p](t, \mathbf{x}, v, \hat{\mathbf{v}}) = \mu(\mathbf{x}) \left(\rho(t, \mathbf{x}) T(\mathbf{x}, v, \hat{\mathbf{v}}) - p(t, \mathbf{x}, v, \hat{\mathbf{v}}) \right)$$



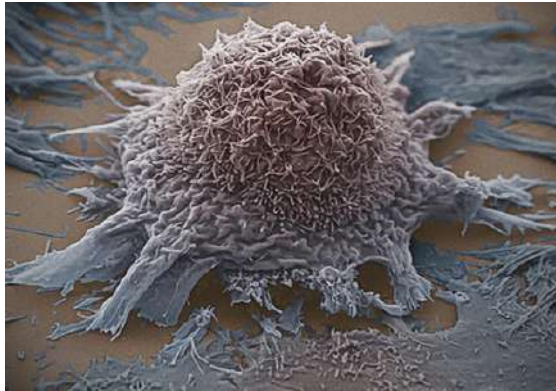
Nonlocal structure of the turning kernel



Given \hat{v}
measures S along \hat{v}
weighting the info

Given \hat{v}
measures S' along \hat{v}
evaluating possible speeds
weighting the info

$$T[S, S'] = c(\mathbf{x}) \int_{\mathbb{R}_+} \gamma_S(\lambda) b(S(\mathbf{x} + \lambda \hat{v})) d\lambda \int_{\mathbb{R}_+} \gamma_{S'}(\lambda') \psi(\mathbf{x}, v | S'(\mathbf{x} + \lambda' \hat{v})) d\lambda'$$



Compact support

Distribution function with
mean = $\bar{U}(\mathbf{x} | S'(\mathbf{x} + \lambda' \hat{v}))$
variance = $D(\mathbf{x} | S'(\mathbf{x} + \lambda' \hat{v}))$



Directional bias (only)

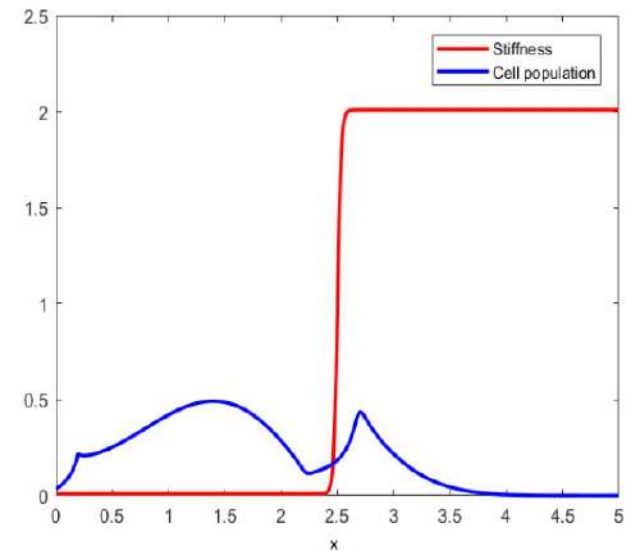
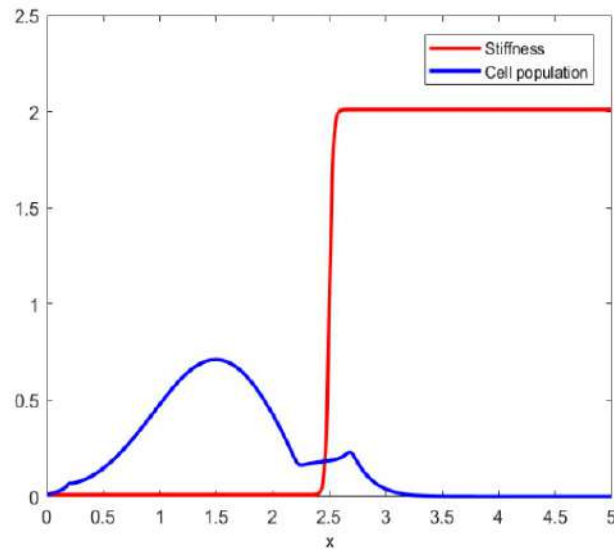
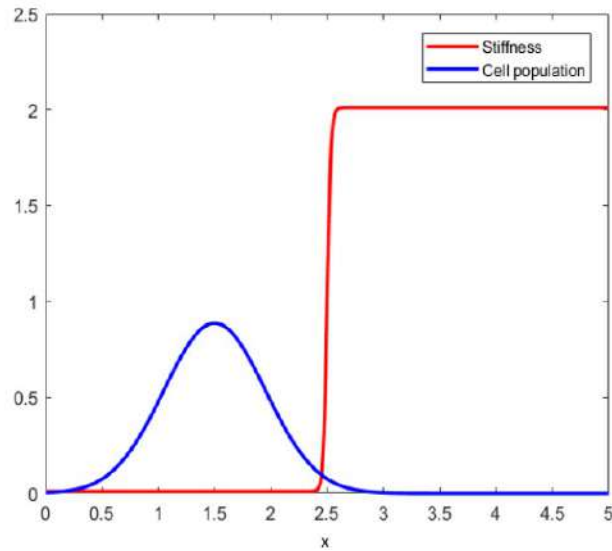
$$T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, v, \hat{\mathbf{v}}) = c(\mathbf{x}) \int_{\mathbb{R}_+} \gamma_{\mathcal{S}}(\lambda) \mathcal{T}_{\lambda}^{\hat{\mathbf{v}}}[\mathcal{S}](\mathbf{x}) d\lambda \int_{\mathbb{R}_+} \gamma_{\mathcal{S}'}(\lambda') \psi(\mathbf{x}, v | \mathcal{S}'(\mathbf{x} + \lambda' \hat{\mathbf{v}})) d\lambda'$$

$$T[\mathcal{S}](\mathbf{x}, \mathbf{v}_p) = \frac{\int_{\mathbb{R}_+} \gamma_{\mathcal{S}}(\lambda) \mathcal{T}_{\lambda}^{\hat{\mathbf{v}}}[\mathcal{S}](\mathbf{x}) d\lambda}{\int_{\mathbb{S}^{d-1}} \int_{\mathbb{R}_+} \gamma_{\mathcal{S}}(\lambda) \mathcal{T}_{\lambda}^{\hat{\mathbf{v}}}[\mathcal{S}](\mathbf{x}) d\lambda d\hat{\mathbf{v}}} \psi(\mathbf{x}, v)$$



Durotaxis

$$\gamma_S = \delta(\lambda - R_S)$$

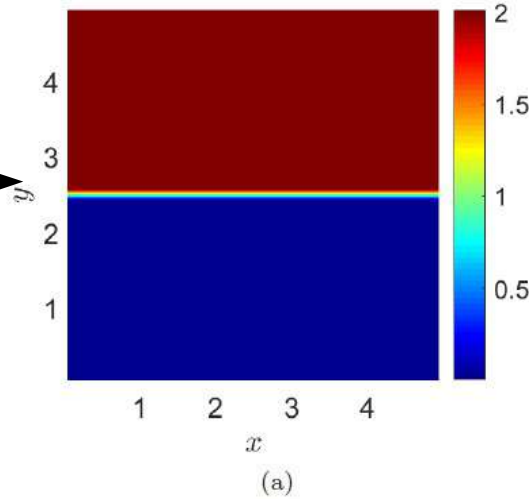
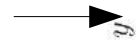




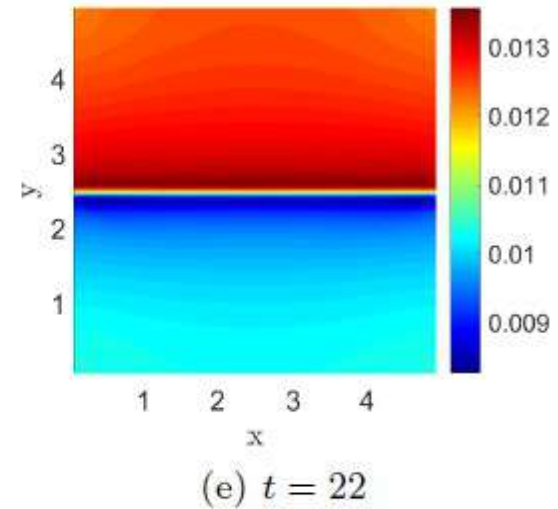
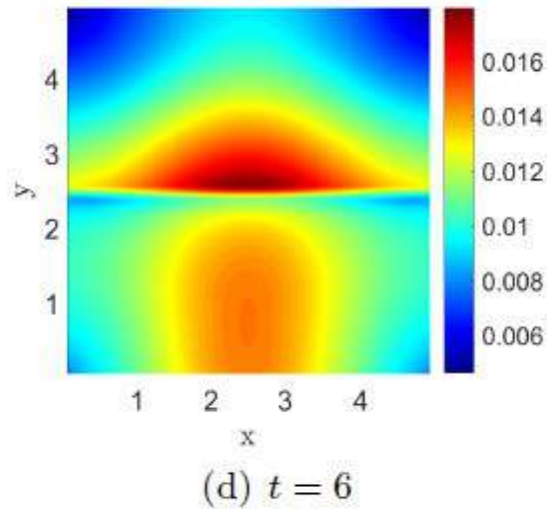
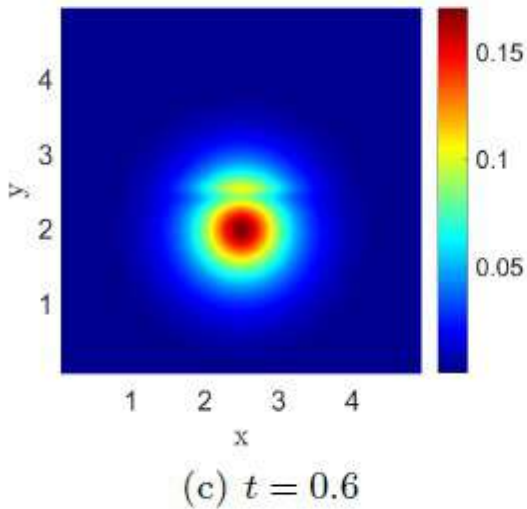
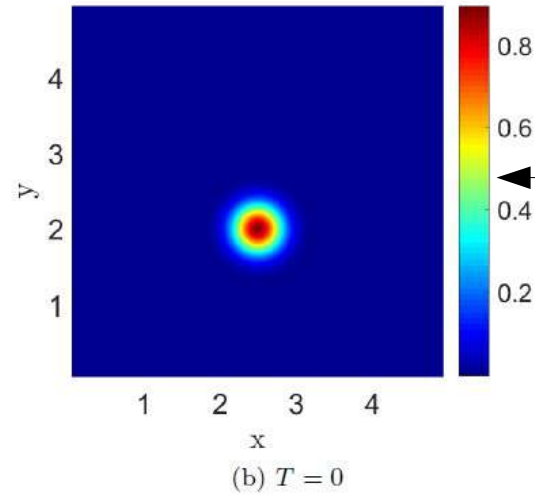
Durotaxis

$$\gamma_S = \delta(\lambda - R_S) \quad R_S = 0.02$$

stiffness

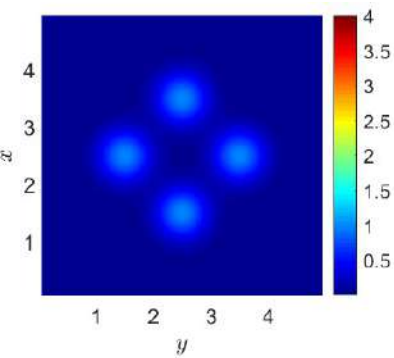


density

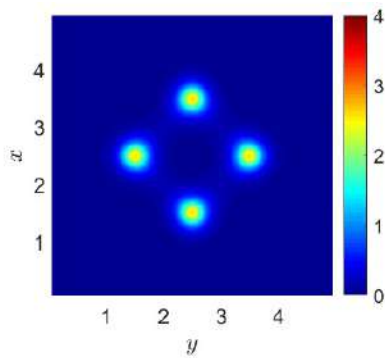


Cell-cell adhesion

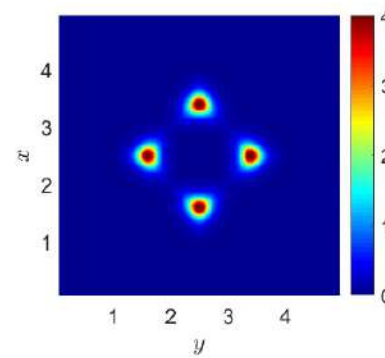
$$\frac{\partial p}{\partial t}(t, \mathbf{x}, v, \hat{\mathbf{v}}) + v \hat{\mathbf{v}} \cdot \nabla p(t, \mathbf{x}, v, \hat{\mathbf{v}}) = \mu \left[c(t, \mathbf{x}) \rho(t, \mathbf{x}) \int_{\mathbb{R}_+} \gamma_R(\lambda) b(\rho(t, \mathbf{x} + \lambda \hat{\mathbf{v}})) d\lambda \psi(v) - p(t, \mathbf{x}, v, \hat{\mathbf{v}}) \right]$$



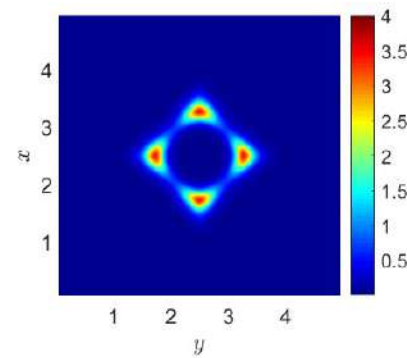
(a) $t = 0$



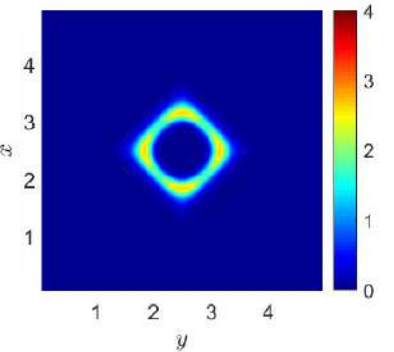
(b) $t = 2.5$



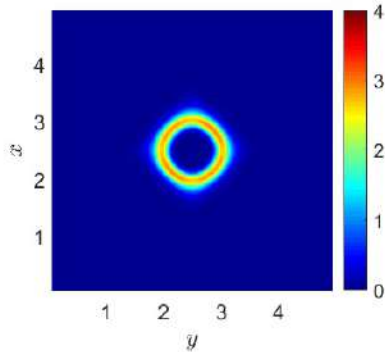
(c) $t = 7.5$



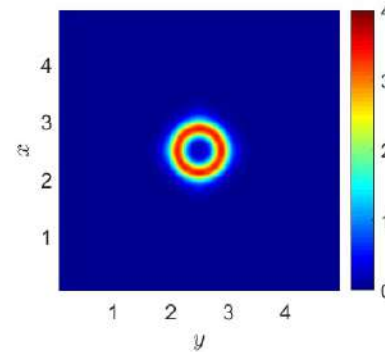
(d) $t = 12.5$



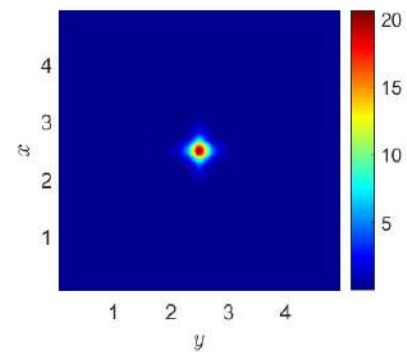
(e) $t = 15$



(f) $t = 17.5$



(g) $t = 20$



(h) $t = 50$

$$\gamma_\rho = \delta(\lambda - R_\rho)$$

$$R_\rho = 0.25$$



Macroscopic limits

Discriminating factor $\mathbf{U}_{S,S'}^0 = \int_{V_p} T[\mathcal{S}, \mathcal{S}']_0 \mathbf{v} d\mathbf{v}_p \stackrel{?}{=} \mathbf{0}$

Yes \rightarrow parabolic $\frac{\partial \rho}{\partial \tau} + \nabla \cdot (\rho \mathbf{U}_{S,S'}^1) = \nabla \cdot \left(\frac{1}{\mu} \nabla \cdot (\mathbb{D}_{S,S'}^0 \rho) \right)$

No \rightarrow hyperbolic $\frac{\partial \rho}{\partial \tau} + \nabla \cdot (\rho \mathbf{U}_{S,S'}^0) = 0$

$$\mathbf{U}_{S,S'}^i = \int_{V_p} T[\mathcal{S}, \mathcal{S}']_i \mathbf{v} d\mathbf{v}_p$$

$$\mathbb{D}_{S,S'}^i = \int_{V_p} T[\mathcal{S}, \mathcal{S}']_i (\mathbf{v} - \mathbf{U}_{S,S'}^i) \otimes (\mathbf{v} - \mathbf{U}_{S,S'}^i) d\mathbf{v}_p$$



Macroscopic limit

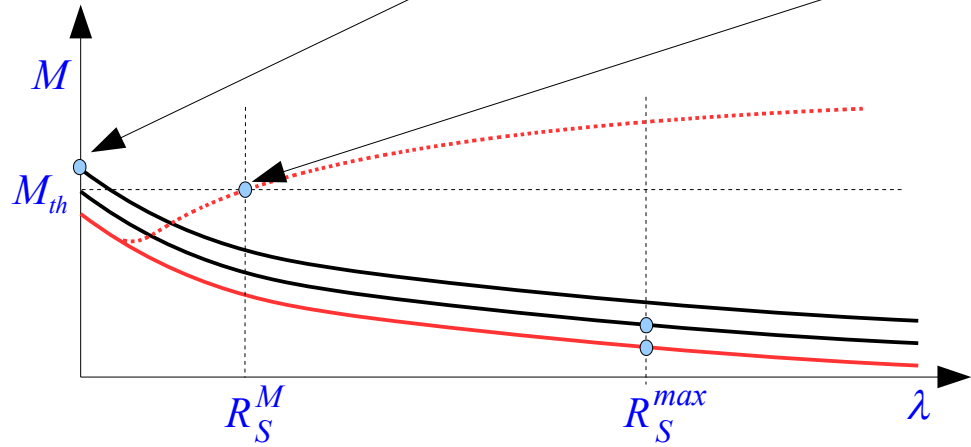
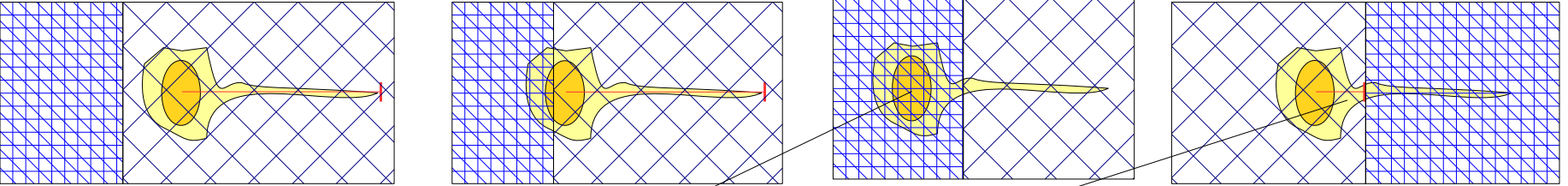
$$\frac{\partial \rho}{\partial \tau} + \nabla \cdot (\rho \mathbf{U}_S^0) = 0$$

For instance, if there is only a signal S affecting the orientation (no S')

$$\mathcal{J}[p](t, \mathbf{x}, \mathbf{v}_p) = \mu(\mathbf{x}) \left(\rho(t, \mathbf{x}) \psi(\mathbf{x}, v) c(\mathbf{x}) \int_{\mathbb{R}_+} b(\mathcal{S}(\mathbf{x} + \lambda \hat{\mathbf{v}})) \gamma_S(\lambda) d\lambda - p(t, \mathbf{x}, \mathbf{v}_p) \right)$$

$$\mathbf{U}_S^0(\boldsymbol{\xi}) = \bar{U}(\boldsymbol{\xi}) \frac{\int_{\mathbb{S}^{d-1}} \left(\int_{\mathbb{R}_+} b(\mathcal{S}(\boldsymbol{\xi} + \lambda \hat{\mathbf{v}})) \gamma_S(\lambda) d\lambda \right) \hat{\mathbf{v}} d\hat{\mathbf{v}}}{\int_{\mathbb{S}^{d-1}} \left(\int_{\mathbb{R}_+} b(\mathcal{S}(\boldsymbol{\xi} + \lambda \hat{\mathbf{v}})) \gamma_S(\lambda) d\lambda \right) d\hat{\mathbf{v}}}$$

Physical limits of migration



limits
their ranges

$$T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, v, \hat{\mathbf{v}}) = c(\mathbf{x}) \int_{\mathbb{R}_+} \gamma_{\mathcal{S}}(\lambda) \mathcal{T}_{\lambda}^{\hat{\mathbf{v}}}[\mathcal{S}](\mathbf{x}) d\lambda \int_{\mathbb{R}_+} \gamma_{\mathcal{S}'}(\lambda') \psi(\mathbf{x}, v | \mathcal{S}'(\mathbf{x} + \lambda' \hat{\mathbf{v}})) d\lambda'$$

Random polarization + volume filling

Random polarization

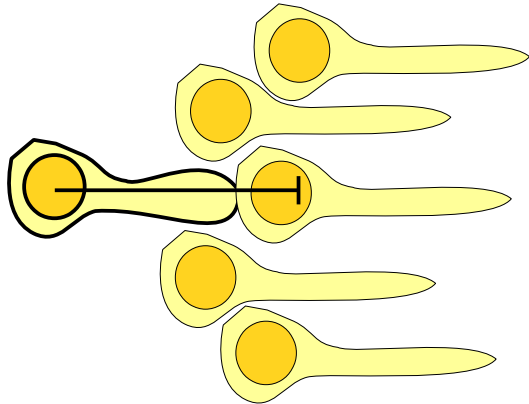
$$T[\mathcal{S}, \mathcal{S}'](\mathbf{x}, \mathbf{v}_p) = c(\mathbf{x}) \int_{\mathbb{R}_+} \gamma_{\mathcal{S}}(\lambda) \cancel{T_{\lambda}^{\hat{\mathbf{v}}}}[\mathcal{S}](\mathbf{x}) d\lambda \int_{\mathbb{R}_+} \gamma_{\mathcal{S}'}(\lambda') \psi(\mathbf{x}, v | \mathcal{S}'(\mathbf{x} + \lambda' \hat{\mathbf{v}})) d\lambda'$$

Volume filling

$$T[\rho](t, \mathbf{x}, v, \hat{\mathbf{v}}) = \frac{1}{\int_{\mathbb{S}^{d-1}} \Gamma'_0(t, \mathbf{x}, \hat{\mathbf{v}}) d\hat{\mathbf{v}}} \int_0^{R_{\rho}(t, \mathbf{x}, \hat{\mathbf{v}})} \gamma_{\rho}(\lambda') \psi(\mathbf{x}, v | \rho(t, \mathbf{x} + \lambda' \hat{\mathbf{v}})) d\lambda'$$

$$\inf \left\{ \lambda' \in [0, R_{\rho}^{max}] \mid \rho(t, \mathbf{x} + \lambda' \hat{\mathbf{v}}) > \rho_{th} \right\}$$

$$\bar{v}(t, \mathbf{x} | \rho(t, \mathbf{x} + \lambda' \hat{\mathbf{v}})) = \bar{v}_M \left(1 - \frac{\rho(t, \mathbf{x} + \lambda' \hat{\mathbf{v}})}{\rho_{th}} \right)_+$$



Nonlocality and radius limitation
tend to prevent overcrowding



Random polarization + volume filling

$$T[\rho](t, \mathbf{x}, v, \hat{\mathbf{v}}) = \frac{1}{\int_{\mathbb{S}^{d-1}} \Gamma'_0(t, \mathbf{x}, \hat{\mathbf{v}}) d\hat{\mathbf{v}}} \int_0^{R_\rho(t, \mathbf{x}, \hat{\mathbf{v}})} \gamma_\rho(\lambda') \psi(\mathbf{x}, v | \rho(t, \mathbf{x} + \lambda' \hat{\mathbf{v}})) d\lambda'$$

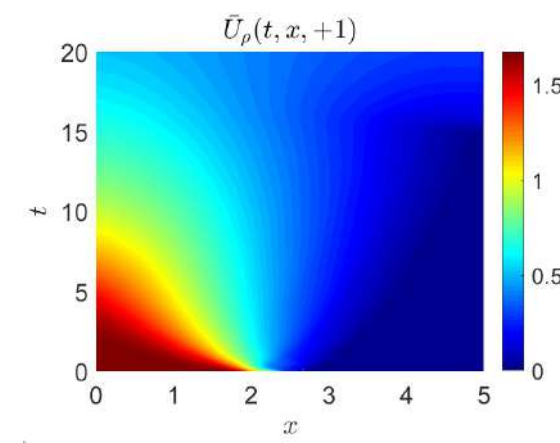
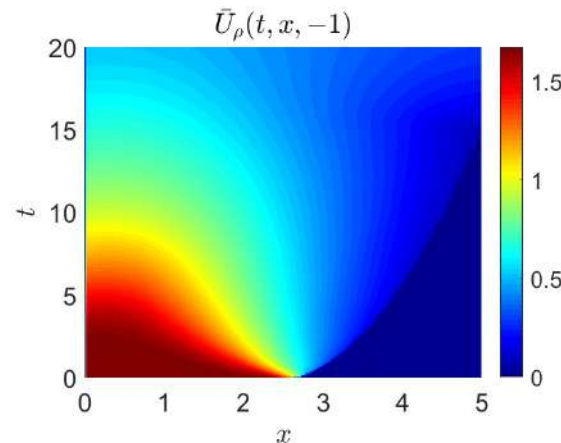
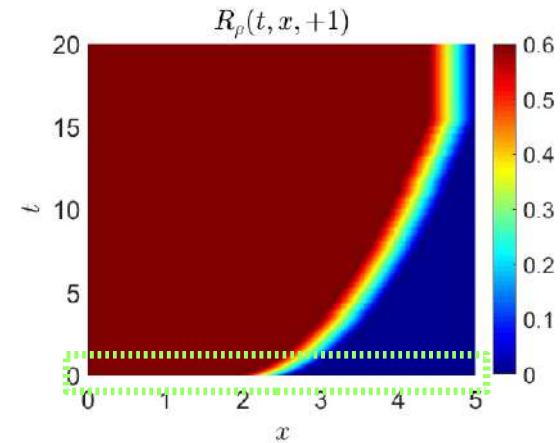
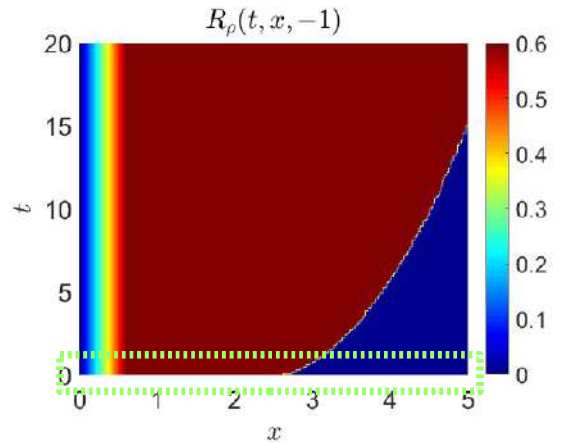
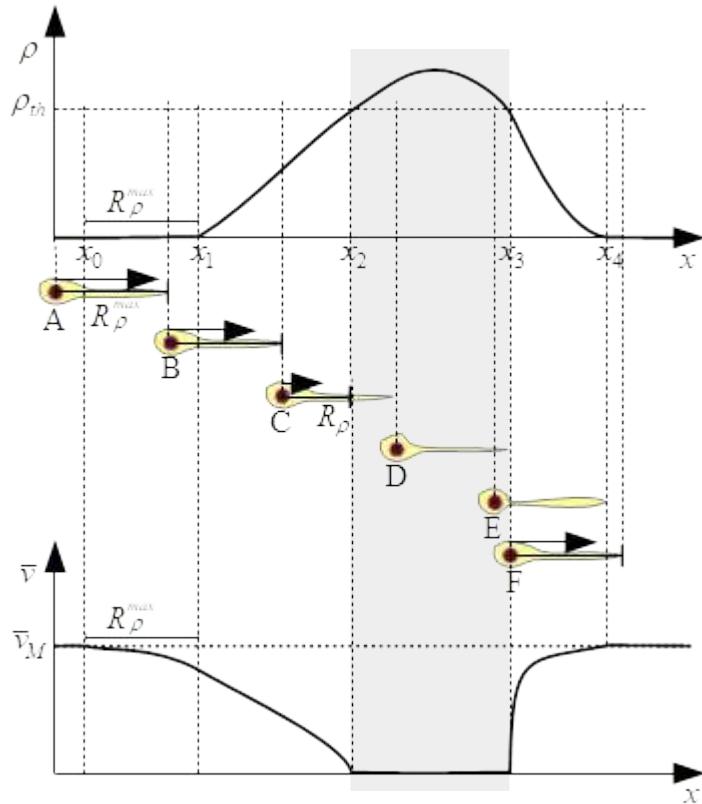
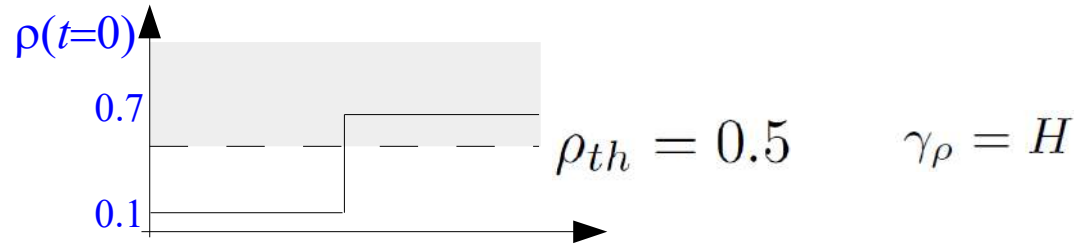
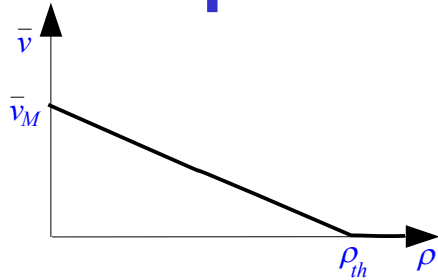
$$\bar{v}(t, \mathbf{x} | \rho(t, \mathbf{x} + \lambda' \hat{\mathbf{v}})) = \bar{v}_M \left(1 - \frac{\rho(t, \mathbf{x} + \lambda' \hat{\mathbf{v}})}{\rho_{th}} \right)_+$$

Hyperbolic scaling

$$\frac{\partial \rho}{\partial \tau} + \nabla \cdot \left\{ \rho \bar{v}_M \frac{1}{\int_{\mathbb{S}^{d-1}} \Gamma'_0 d\hat{\mathbf{v}}} \left[\int_{\mathbb{S}^{d-1}} \Gamma'_0 \hat{\mathbf{v}} d\hat{\mathbf{v}} - \frac{1}{\rho_{th}} \int_{\mathbb{S}^{d-1}} \left(\int_0^{R_\rho(\hat{\mathbf{v}})} \rho(\mathbf{x} + \lambda' \hat{\mathbf{v}}) \gamma_\rho(\lambda') d\lambda' \right) \hat{\mathbf{v}} d\hat{\mathbf{v}} \right]_+ \right\} = 0$$



Volume filling as a Physical Limit of Migration

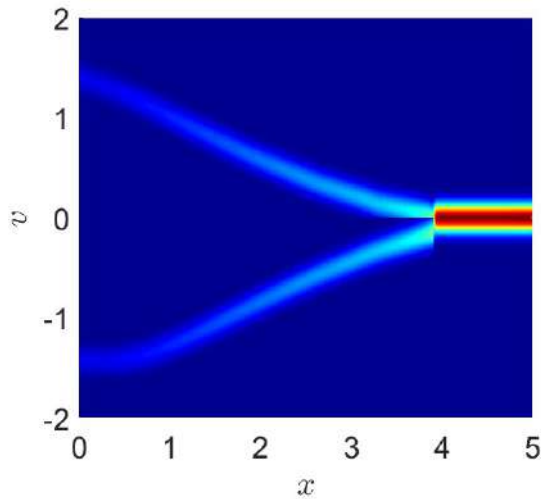
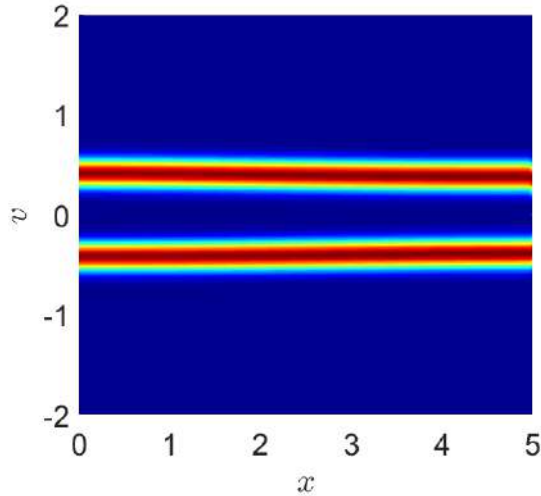




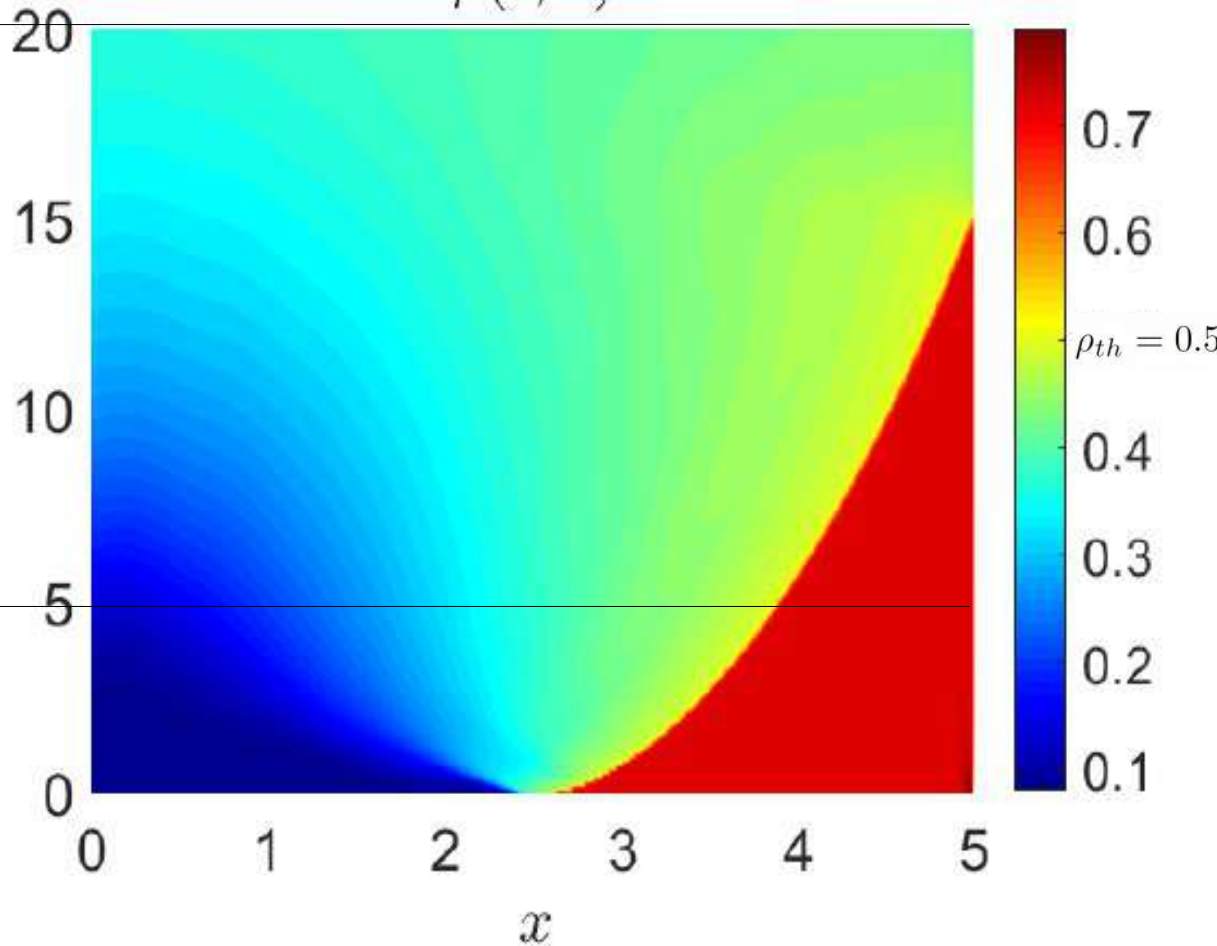
Volume filling as a Physical Limit of Migration

No motion in the overcrowded region due to limited radius

Nonlocality drives dispersal from the interface



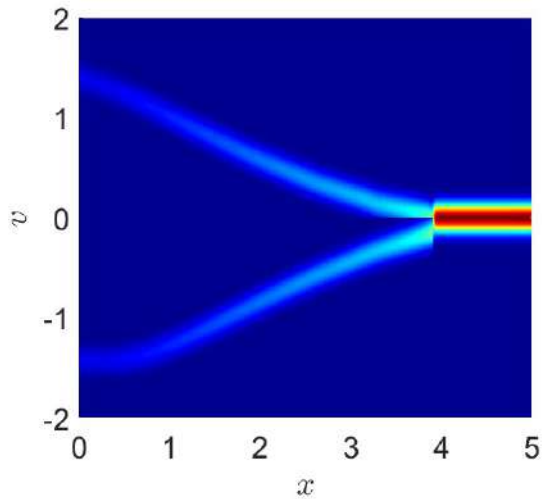
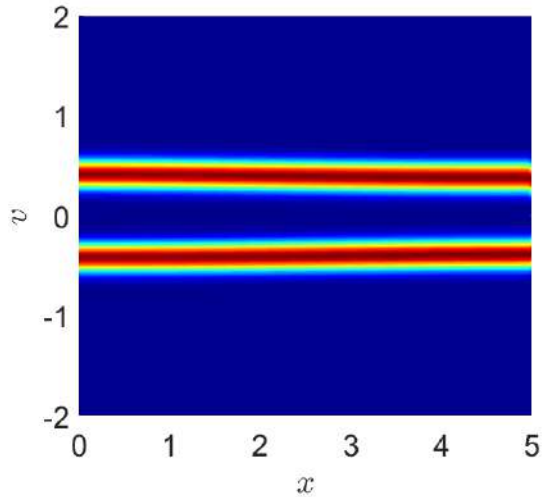
$\rho(t, x)$



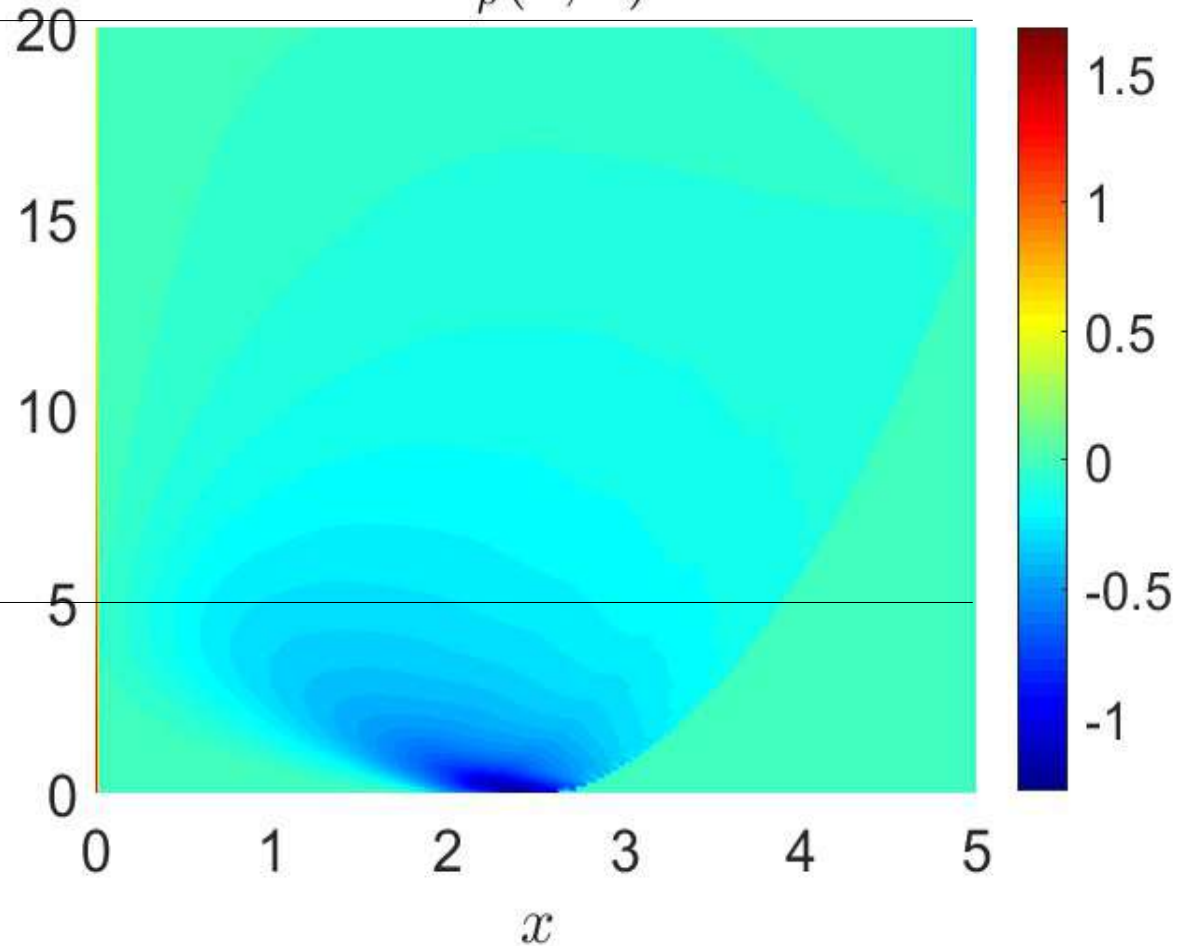


Volume filling as a Physical Limit of Migration

Cells do not move in the overcrowded region due to limited radius

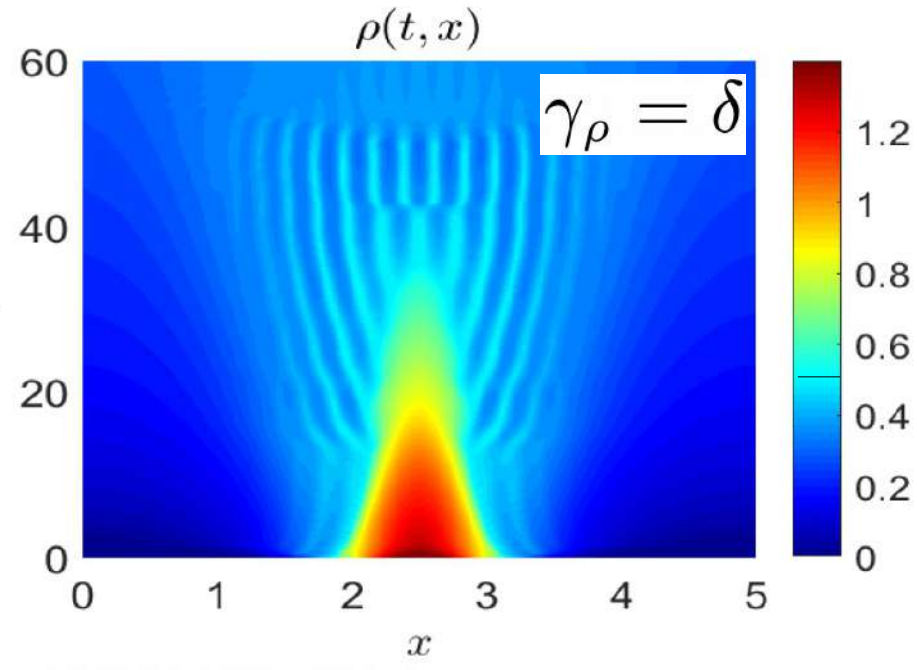
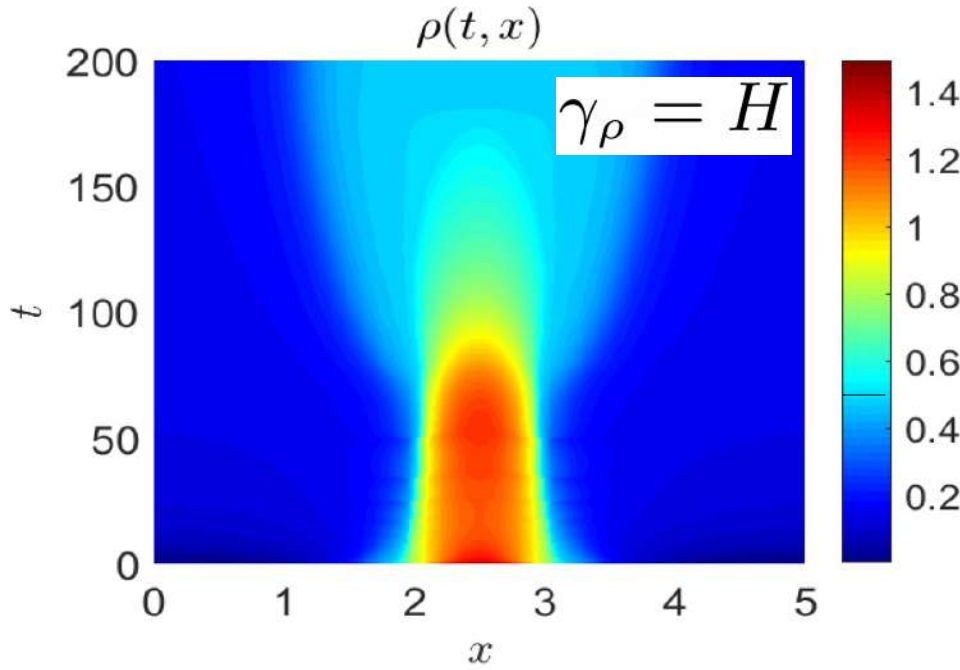


$$U_{\rho}(x, \hat{\mathbf{v}})$$





Volume filling



$$R_\rho^{max} = 0.2 \quad \rho_{th} = 0.5.$$



$$\frac{\partial p}{\partial t}(t, \mathbf{x}, v, \hat{\mathbf{v}}) + v \hat{\mathbf{v}} \cdot \nabla p(t, \mathbf{x}, v, \hat{\mathbf{v}}) = \frac{1}{\gamma} \left[\frac{\rho(t, \mathbf{x})}{\Gamma |\mathbb{S}^{d-1}|} \int_{\mathbb{R}_+} \gamma(\lambda) \psi(v | \rho(t, \mathbf{x} + \lambda \hat{\mathbf{v}})) d\lambda - p(t, \mathbf{x}, v, \hat{\mathbf{v}}) \right]$$

Homogeneous equilibrium:
$$p_\infty = \frac{1}{|\mathbb{S}^{d-1}|} \psi(v | 1)$$

Perturbation equation

$$\frac{\partial p}{\partial t}(t, \mathbf{x}, v, \hat{\mathbf{v}}) + v \hat{\mathbf{v}} \cdot \nabla p(t, \mathbf{x}, v, \hat{\mathbf{v}}) = \frac{1}{\gamma} \left[\frac{\rho(t, \mathbf{x})}{|\mathbb{S}^{d-1}|} \psi(v | 1) + \frac{1}{\Gamma |\mathbb{S}^{d-1}|} \int_{\mathbb{R}_+} \gamma(\lambda) \rho(t, \mathbf{x} + \lambda \hat{\mathbf{v}}) d\lambda \frac{\partial \psi}{\partial \rho}(v | 1) - p(t, \mathbf{x}, v, \hat{\mathbf{v}}) \right]$$



Stability

$$p(t, \mathbf{x}, v, \hat{\mathbf{v}}) = g(v, \hat{\mathbf{v}}) e^{i\mathbf{k} \cdot \mathbf{x} + \sigma t}$$

$$\rho_g = \int_{\mathbb{S}^{d-1}} \int_0^U g(v, \hat{\mathbf{v}}) dv d\hat{\mathbf{v}}$$

$$\longrightarrow \left(\sigma + i\mathbf{k} \cdot \hat{\mathbf{v}}v + \frac{1}{\mathcal{V}} \right) g = \frac{1}{\mathcal{V} |\mathbb{S}^{d-1}|} \left[\psi(v|1) + \hat{\gamma}(\mathbf{k} \cdot \hat{\mathbf{v}}) \frac{\partial \psi}{\partial \rho}(v|1) \right] \rho_g$$

$$\mathcal{V} = \frac{\bar{v}(\rho_\infty)}{R\mu}$$

$$\frac{1}{\Gamma} \int_{\mathbb{R}_+} \gamma(\lambda) e^{i\mathbf{k} \cdot \hat{\mathbf{v}}\lambda} d\lambda$$

$$\left\{ \begin{array}{l} \int_{\mathbb{S}^{d-1}} \int_0^U \frac{\left[\psi(v|1) + \frac{\partial \psi}{\partial \rho}(v|1) \hat{\gamma}_c(\mathbf{k} \cdot \hat{\mathbf{v}}) \right] \left(\sigma_r + \frac{1}{\mathcal{V}} \right) + \frac{\partial \psi}{\partial \rho}(v|1) \hat{\gamma}_s(\mathbf{k} \cdot \hat{\mathbf{v}}) (\sigma_i + \mathbf{k} \cdot \hat{\mathbf{v}}v)}{\left(\sigma_r + \frac{1}{\mathcal{V}} \right)^2 + (\sigma_i + \mathbf{k} \cdot \hat{\mathbf{v}}v)^2} dv d\hat{\mathbf{v}} = |\mathbb{S}^{d-1}| \mathcal{V} \\ \int_{\mathbb{S}^{d-1}} \int_0^U \frac{\left[\psi(v|1) + \frac{\partial \psi}{\partial \rho}(v|1) \hat{\gamma}_c(\mathbf{k} \cdot \hat{\mathbf{v}}) \right] (\sigma_i + \mathbf{k} \cdot \hat{\mathbf{v}}v) - \frac{\partial \psi}{\partial \rho}(v|1) \hat{\gamma}_s(\mathbf{k} \cdot \hat{\mathbf{v}}) \mathbf{k} \cdot \hat{\mathbf{v}}v \left(\sigma + \frac{1}{\mathcal{V}} \right)}{\left(\sigma_r + \frac{1}{\mathcal{V}} \right)^2 + (\sigma_i + \mathbf{k} \cdot \hat{\mathbf{v}}v)^2} dv d\hat{\mathbf{v}} = 0. \end{array} \right.$$

$$\hat{\gamma}_c(\mathbf{k} \cdot \hat{\mathbf{v}}) = \frac{1}{\Gamma} \int_{\mathbb{R}_+} \gamma(\lambda) \cos(\lambda \mathbf{k} \cdot \hat{\mathbf{v}}) d\lambda, \quad \text{and} \quad \hat{\gamma}_s(\mathbf{k} \cdot \hat{\mathbf{v}}) = \frac{1}{\Gamma} \int_{\mathbb{R}_+} \gamma(\lambda) \sin(\lambda \mathbf{k} \cdot \hat{\mathbf{v}}) d\lambda$$



Stability

$$p(t, \mathbf{x}, v, \hat{\mathbf{v}}) = g(v, \hat{\mathbf{v}}) e^{i\mathbf{k} \cdot \mathbf{x} + \sigma t}$$

$$\rho_g = \int_{\mathbb{S}^{d-1}} \int_0^U g(v, \hat{\mathbf{v}}) dv d\hat{\mathbf{v}}$$

$$\longrightarrow \left(\sigma + i\mathbf{k} \cdot \hat{\mathbf{v}}v + \frac{1}{\gamma} \right) g = \frac{1}{\gamma |\mathbb{S}^{d-1}|} \left[\psi(v|1) + \hat{\gamma}(\mathbf{k} \cdot \hat{\mathbf{v}}) \frac{\partial \psi}{\partial \rho}(v|1) \right] \rho_g$$

$$\gamma = \frac{\bar{v}(\rho_\infty)}{R\mu}$$

$$\frac{1}{\Gamma} \int_{\mathbb{R}_+} \gamma(\lambda) e^{i\mathbf{k} \cdot \hat{\mathbf{v}}\lambda} d\lambda$$

In 1D

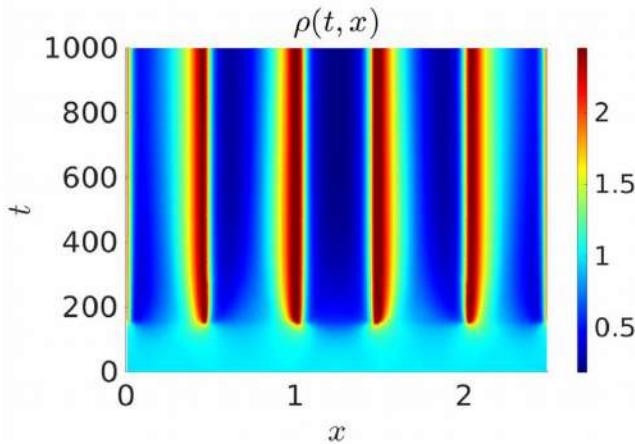
$$\begin{cases} \left[\psi(v|1) + \frac{\partial \psi}{\partial \rho}(v|1) \hat{\gamma}_c(k) \right] \sigma_i = 0 \\ \cdot 1 = \frac{1}{\gamma} \int_0^U \frac{\left(\psi(v|1) + \frac{\partial \psi}{\partial \rho}(v|1) \hat{\gamma}_c(k) \right) \left(\sigma + \frac{1}{\gamma} \right) + \frac{\partial \psi}{\partial \rho}(v|1) \hat{\gamma}_s(k) kv}{\left(\sigma + \frac{1}{\gamma} \right)^2 + k^2 v^2} dv \end{cases}$$

$$\hat{\gamma}_c(\mathbf{k} \cdot \hat{\mathbf{v}}) = \frac{1}{\Gamma} \int_{\mathbb{R}_+} \gamma(\lambda) \cos(\lambda \mathbf{k} \cdot \hat{\mathbf{v}}) d\lambda, \quad \text{and} \quad \hat{\gamma}_s(\mathbf{k} \cdot \hat{\mathbf{v}}) = \frac{1}{\Gamma} \int_{\mathbb{R}_+} \gamma(\lambda) \sin(\lambda \mathbf{k} \cdot \hat{\mathbf{v}}) d\lambda$$

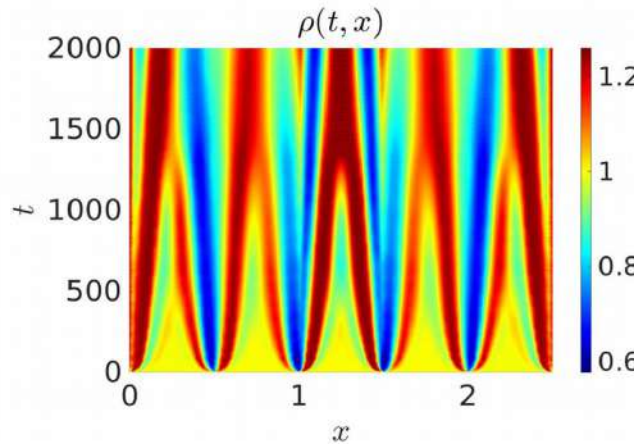


Stability: Dirac sensing

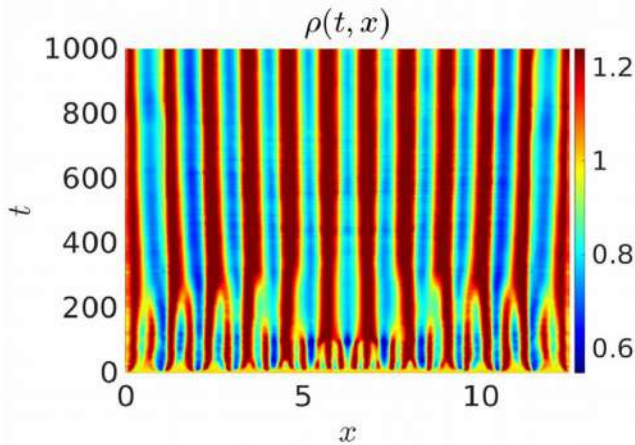
Always unstable to short waves



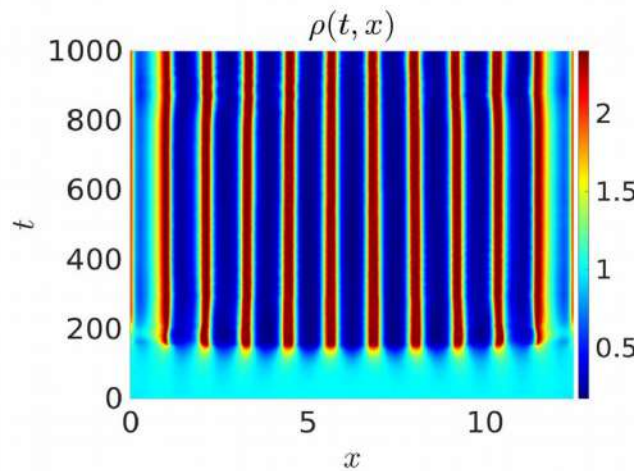
(a) $M = 4.13, \mathcal{V} = 0.015$



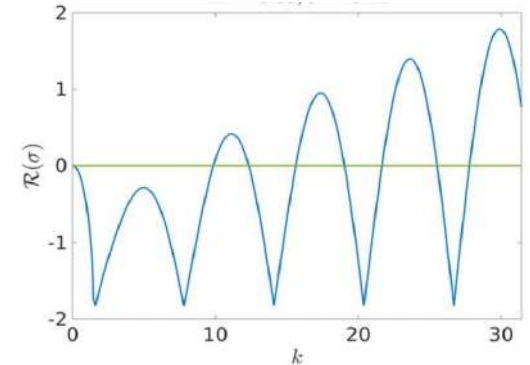
(b) $M = 4.13, \mathcal{V} = 0.005$



(c) $M = 0.68, \mathcal{V} = 0.02$



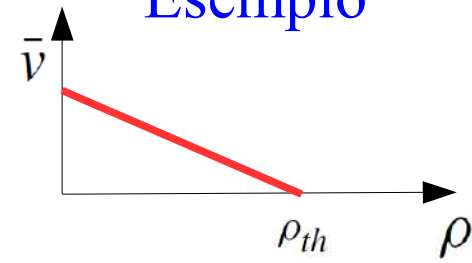
(d) $M = 0.68, \mathcal{V} = 0.008$



$$\mathcal{V} = \frac{\bar{v}(\rho_\infty)}{R\mu}$$

$$M = -\frac{\bar{v}'(\rho_\infty)R}{\bar{v}(\rho_\infty)}$$

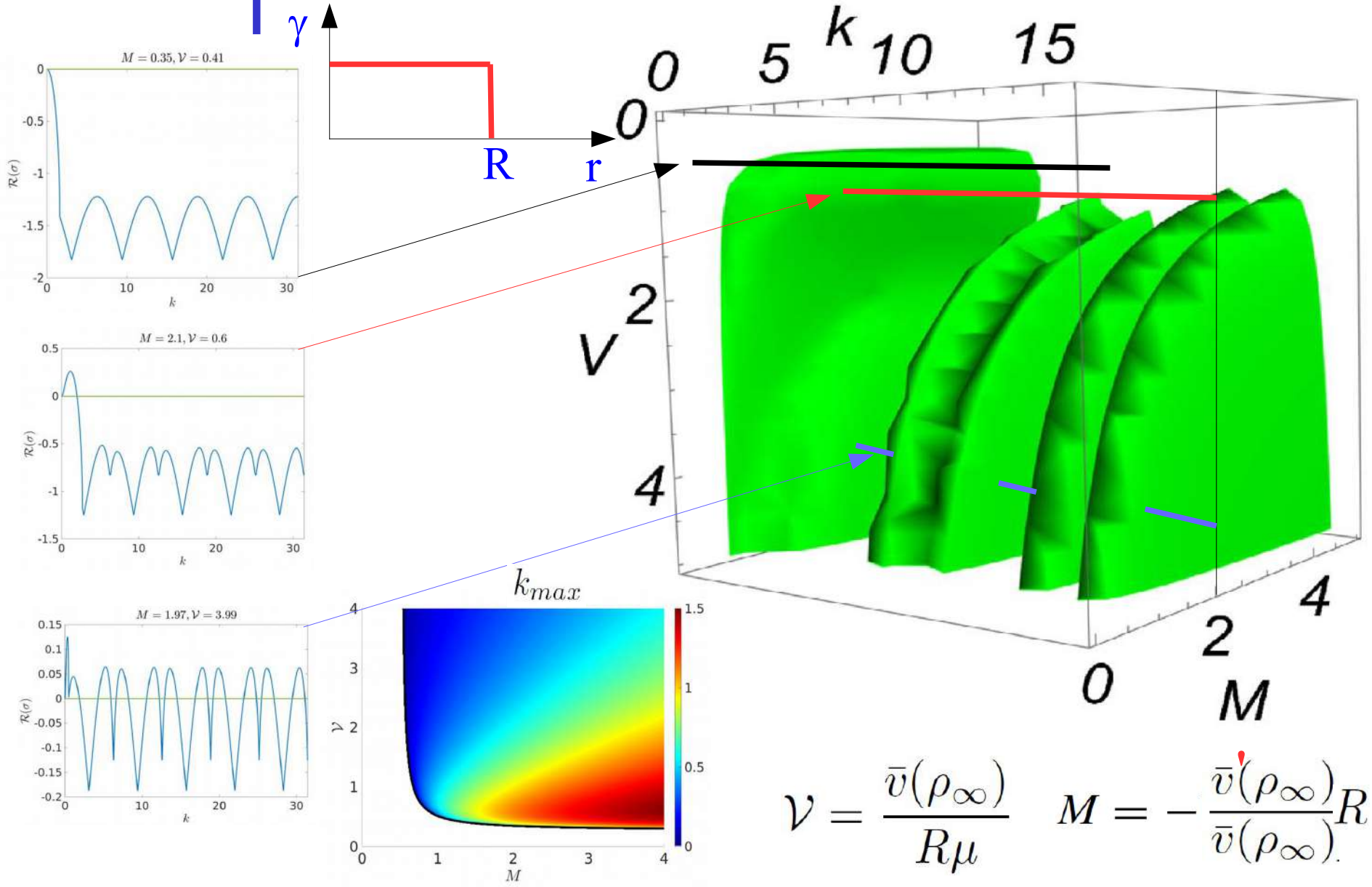
Esempio



$$M = \frac{\rho_\infty}{\rho_{th} - \rho_\infty}$$



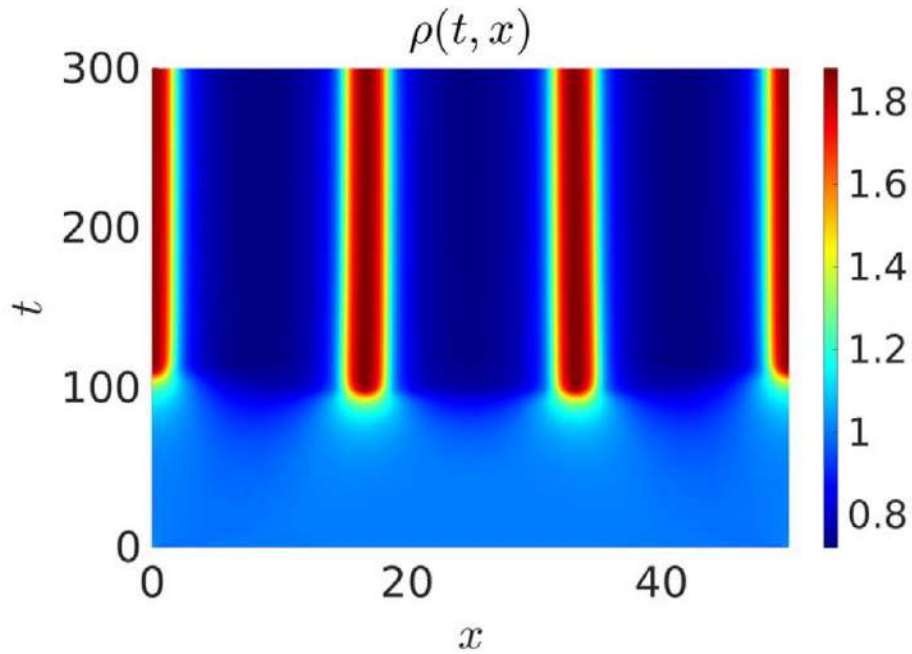
Stability: Uniform sensing



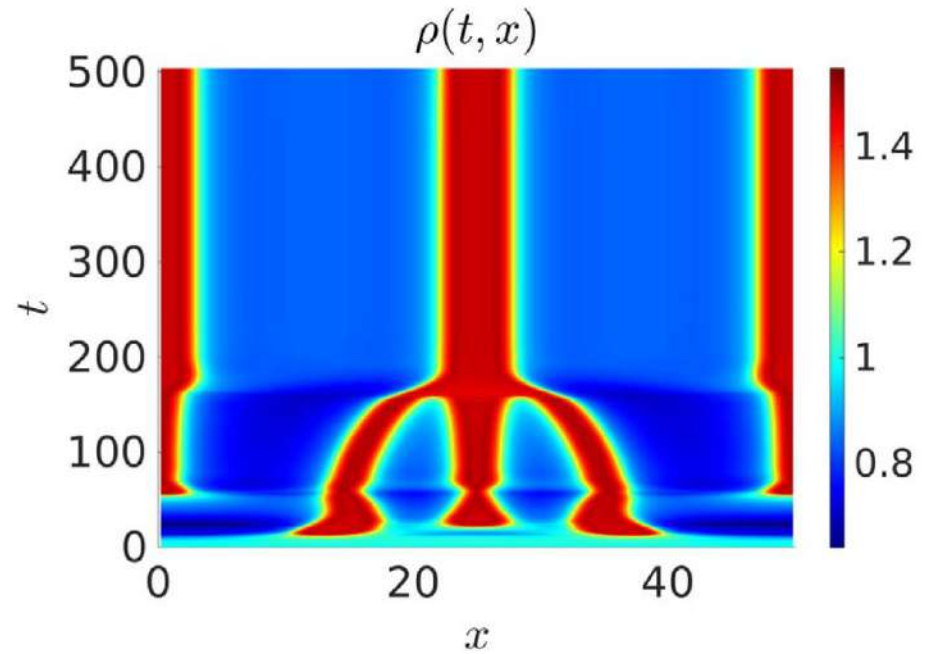
$$\mathcal{V} = \frac{\bar{v}(\rho_\infty)}{R\mu} \quad M = -\frac{\bar{v}'(\rho_\infty)}{\bar{v}(\rho_\infty)}R$$



Stability: Uniform sensing



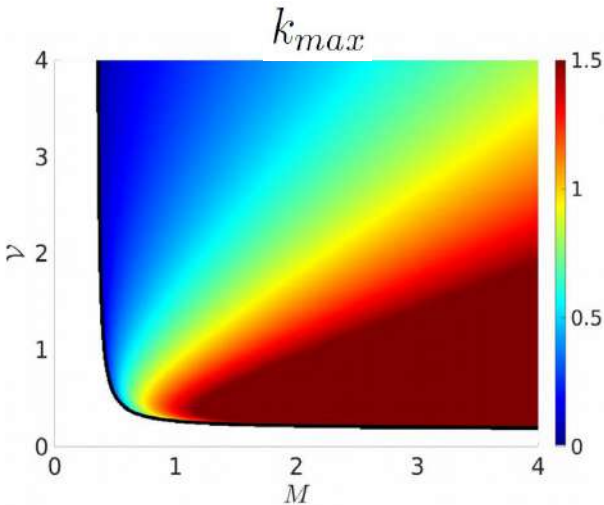
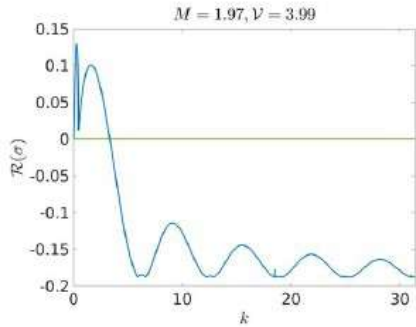
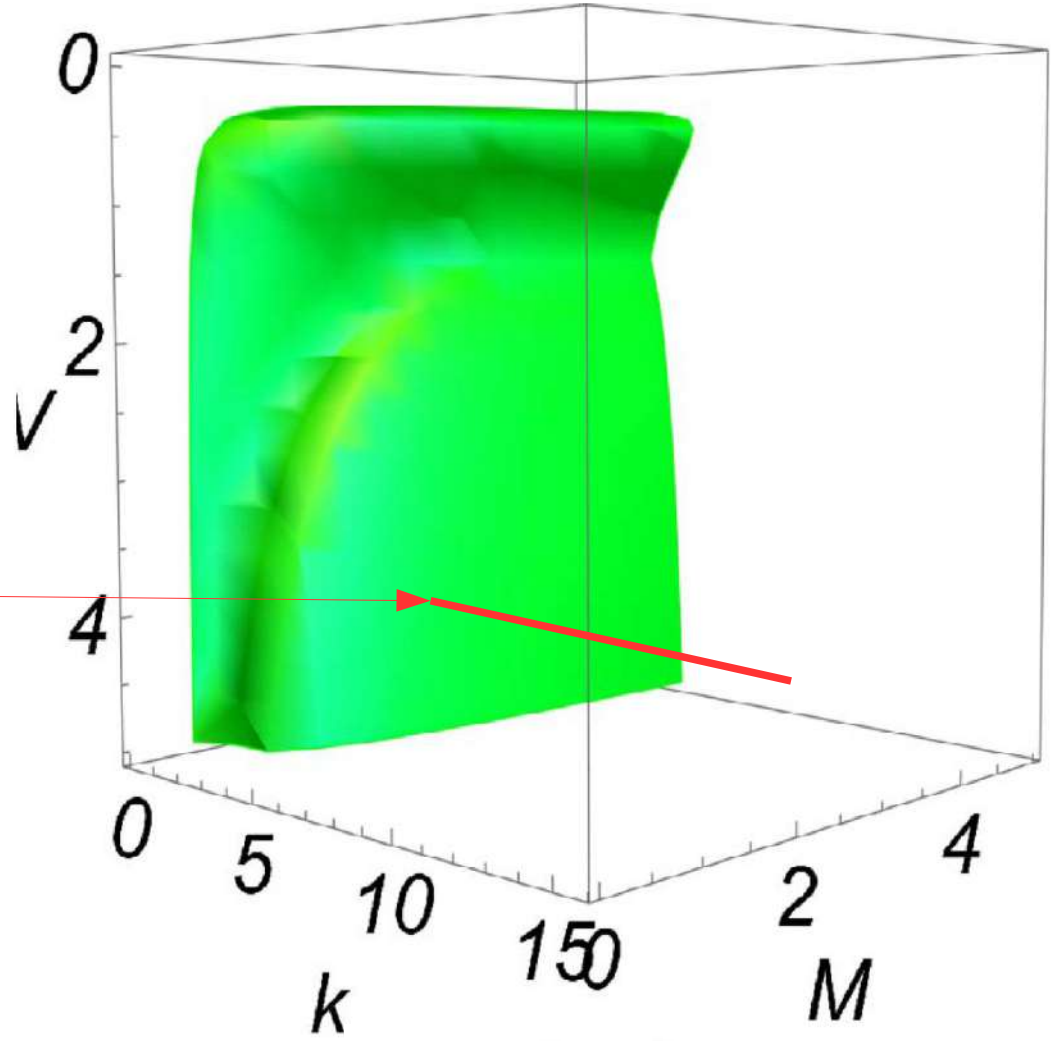
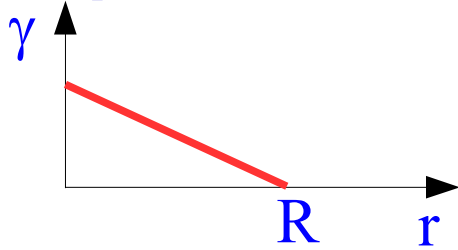
(a) $M = 1.01, \mathcal{V} = 1.65, k_{max} = 0.35, \Lambda = 17.95$



(b) $M = 1.97, \mathcal{V} = 3.99, k_{max} = 0.28, \Lambda = 22.42$



Stability: Decreasing sensing

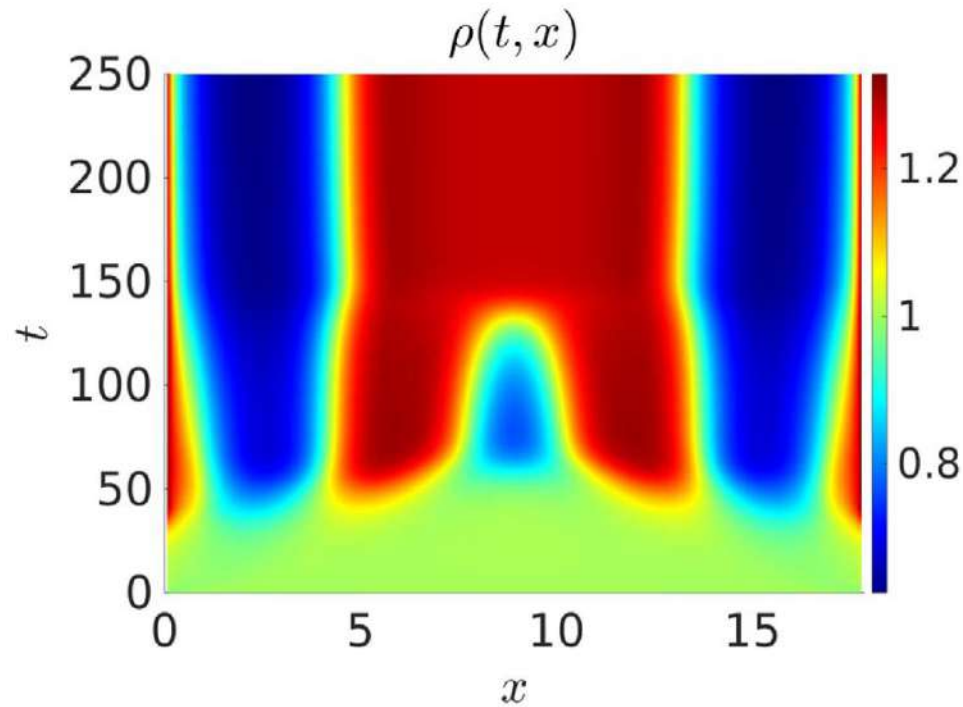


$$\psi = \frac{\bar{v}(\rho_\infty)}{R\mu}$$

$$M = -V'(1)$$



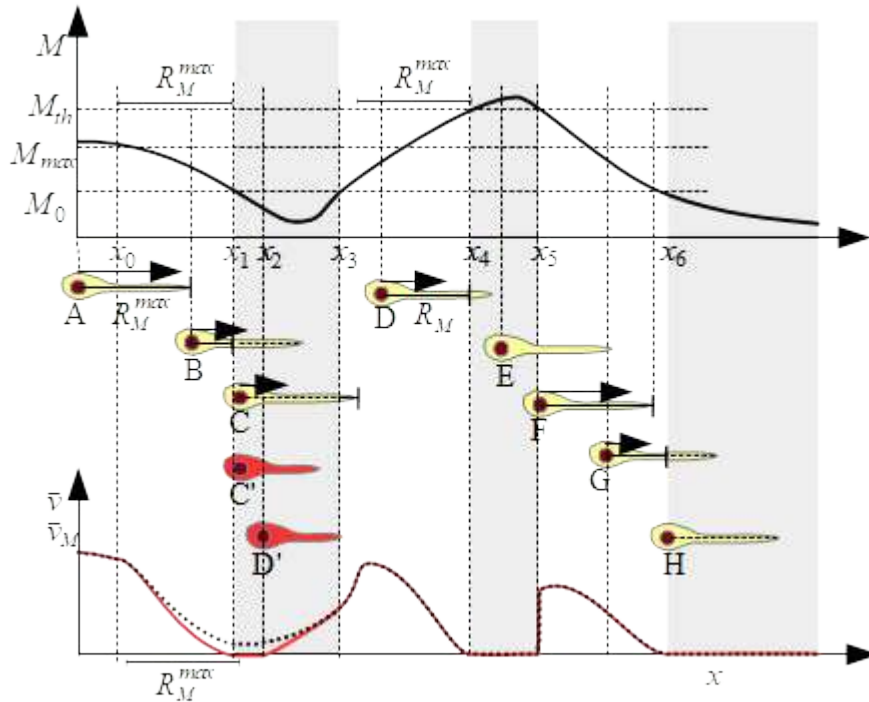
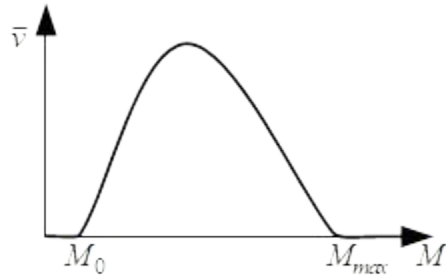
Stability: Decreasing sensing



(c) $M = 2.1$, $\mathcal{V} = 0.6$, $k_{max} = 1$, $\Lambda = 6.3$

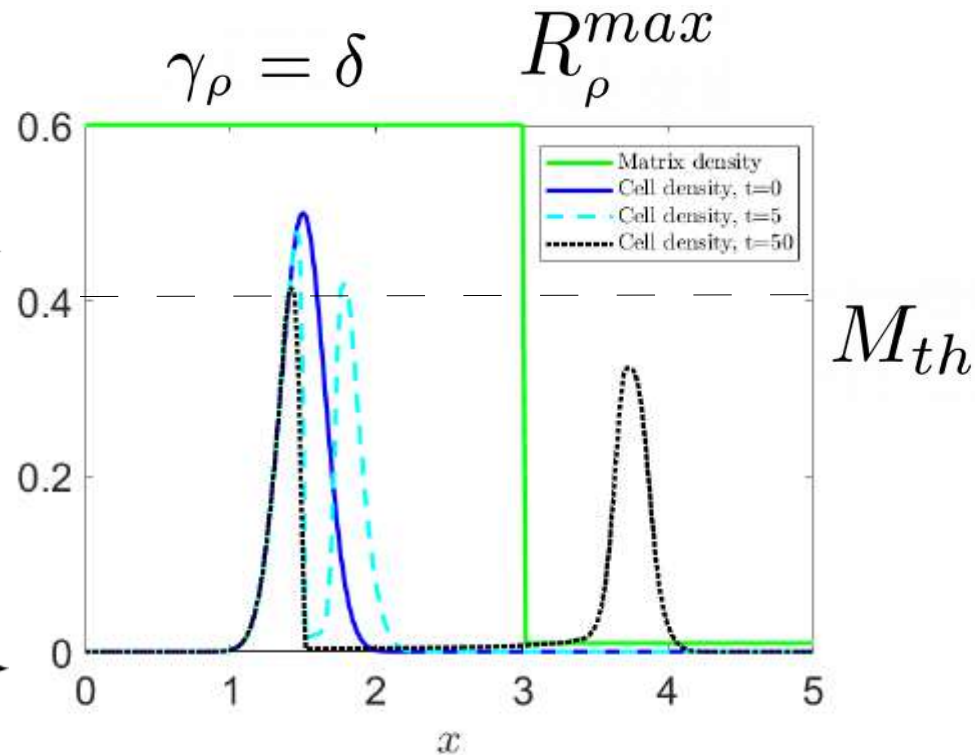
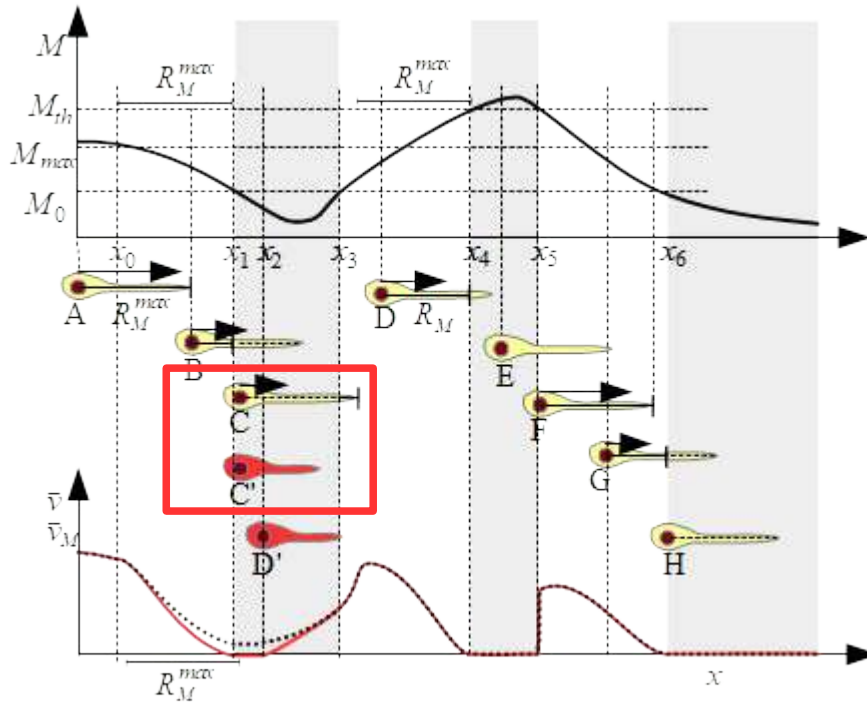
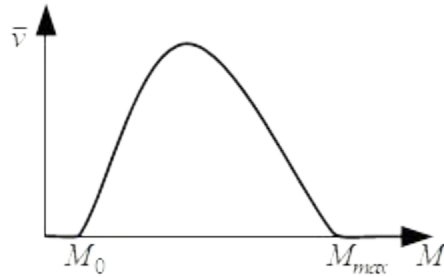


ECM as a Physical Limit of Migration



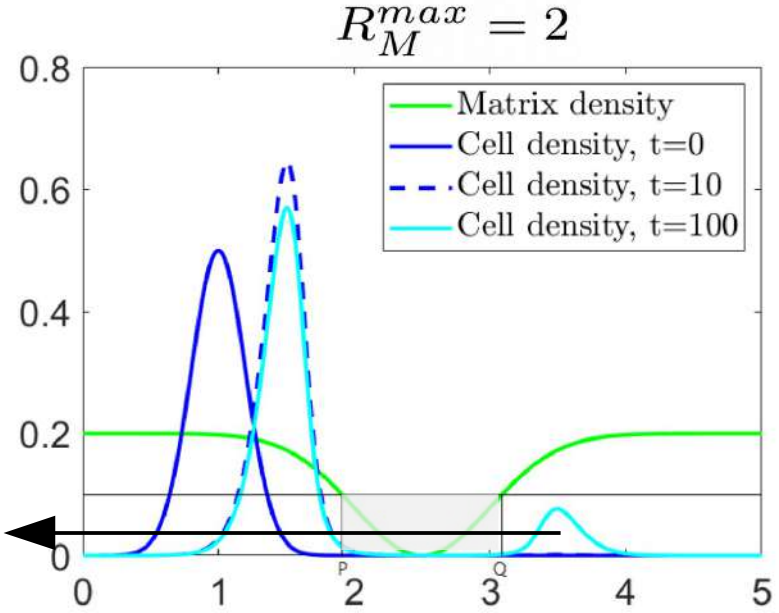
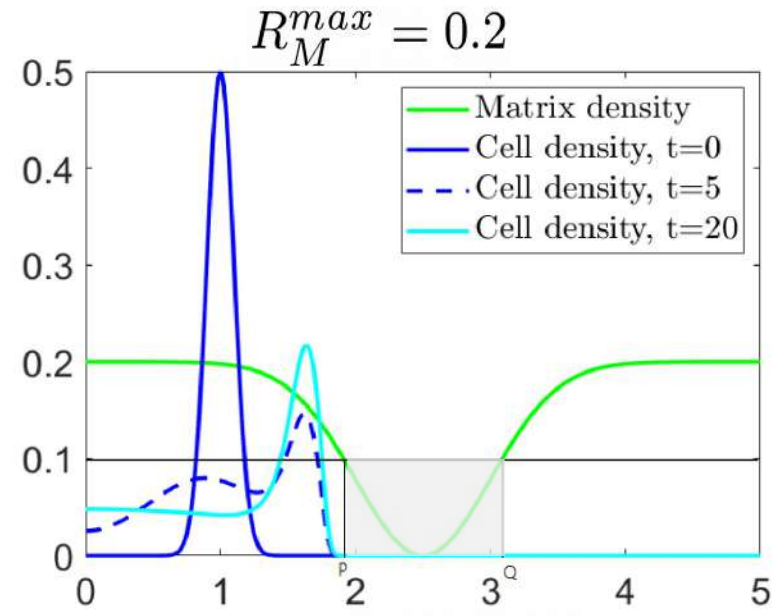
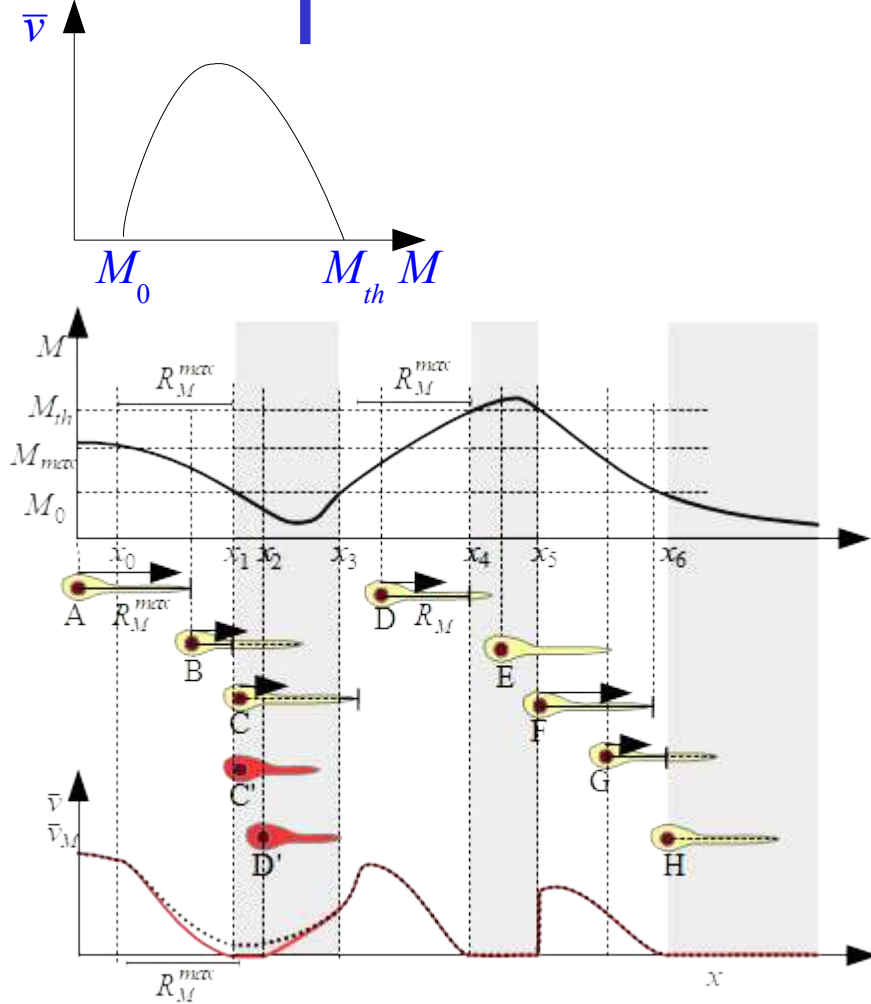


ECM barrier





Lack of adhesion sites

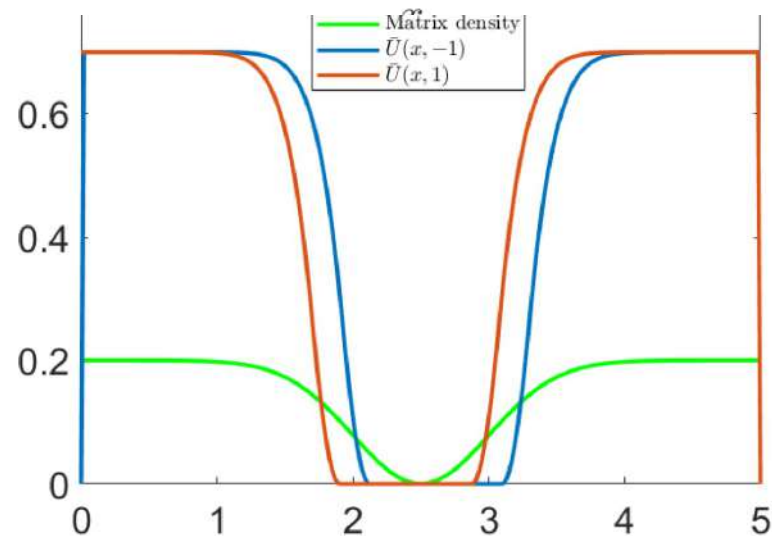
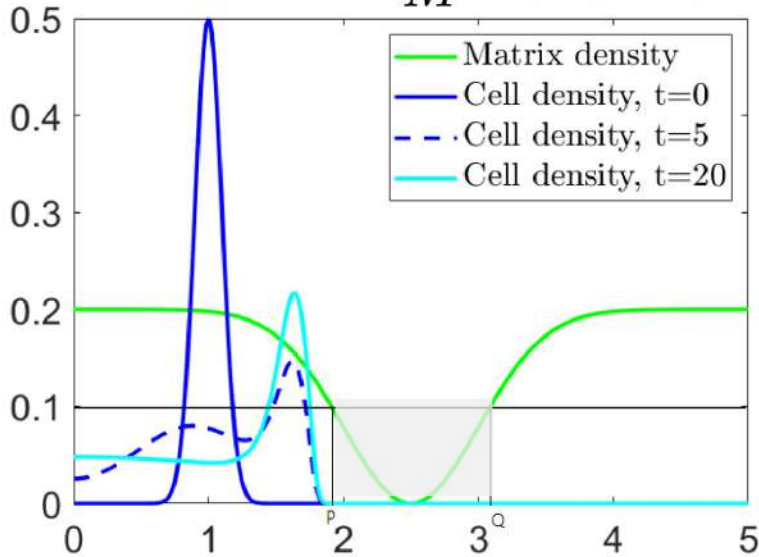


Nonlocality allows jumping beyond the pit

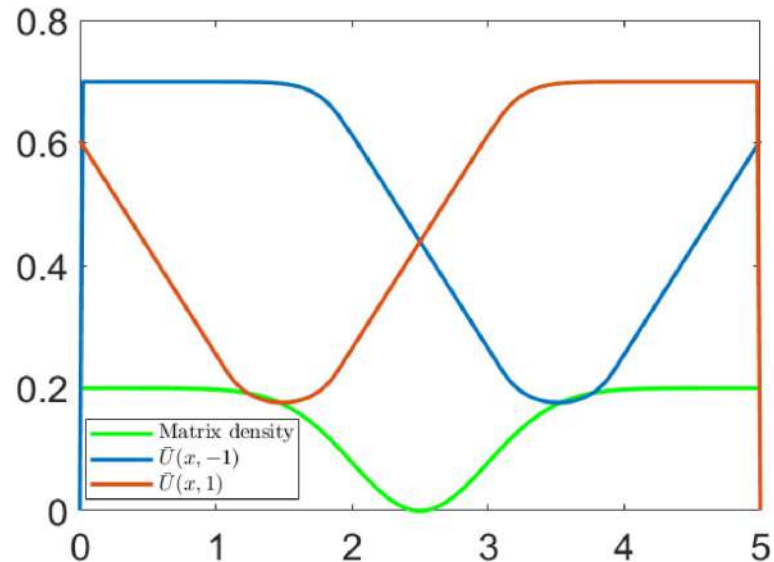
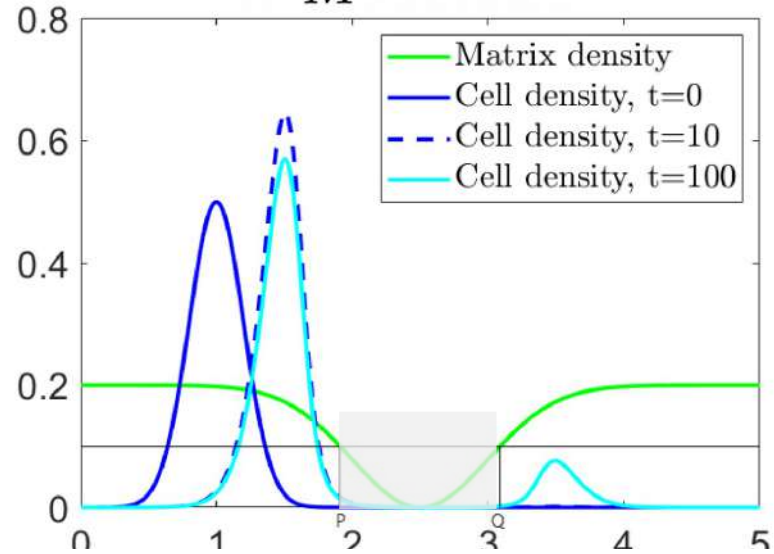


Poor ECM

$$R_M^{max} = 0.2$$



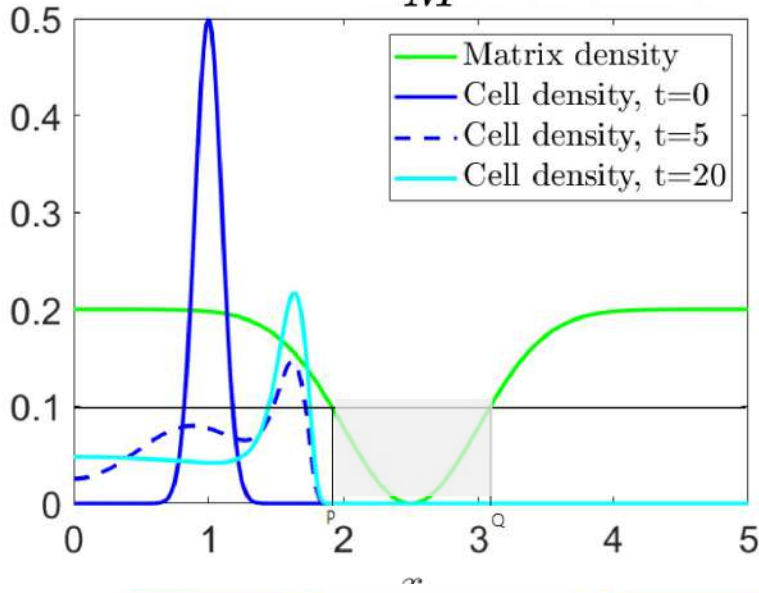
$$R_M^{max} = 2$$



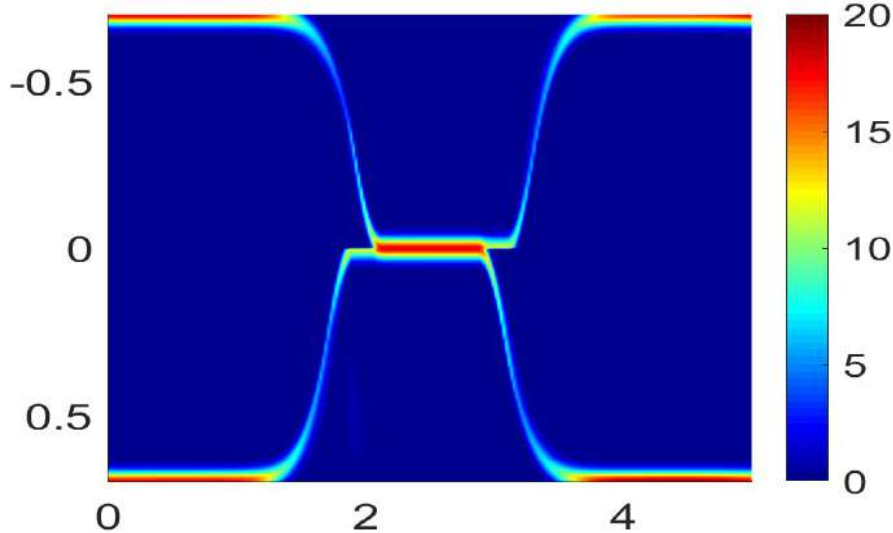
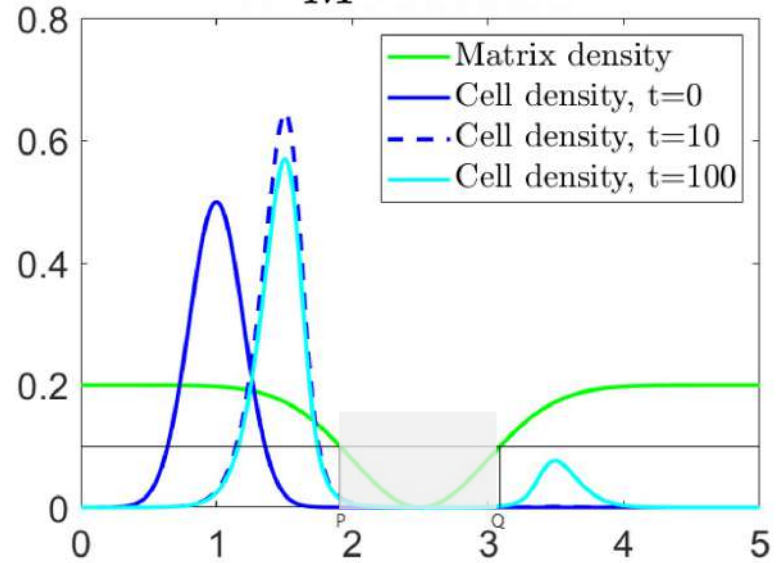


Poor ECM

$$R_M^{max} = 0.2$$



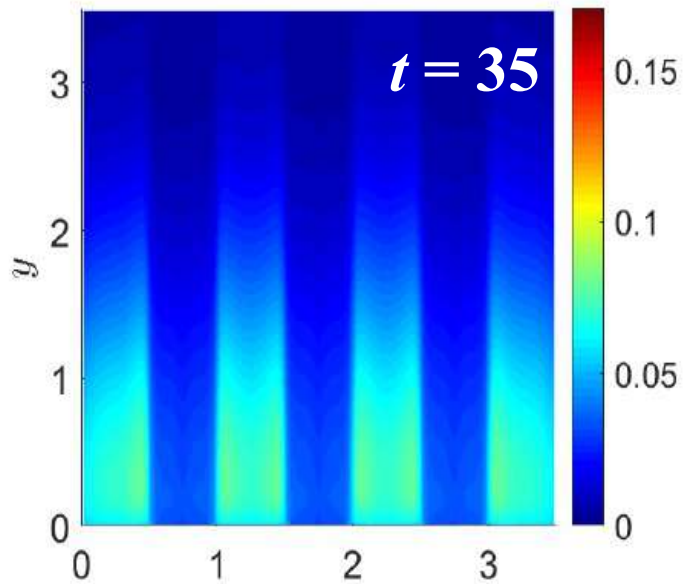
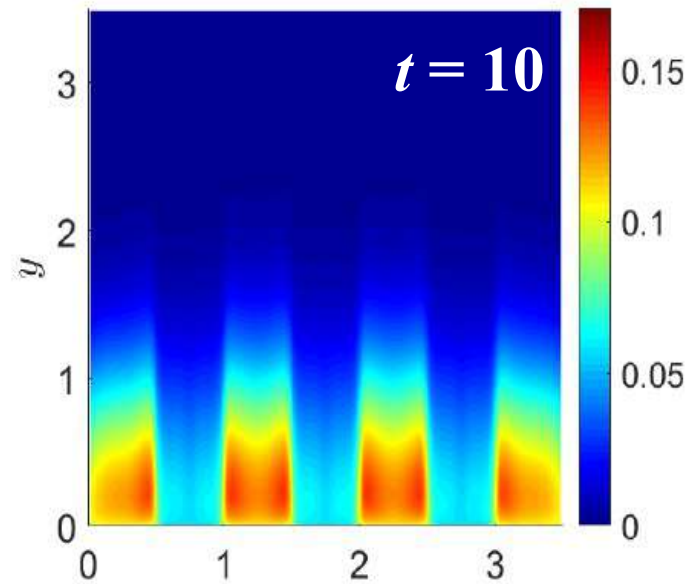
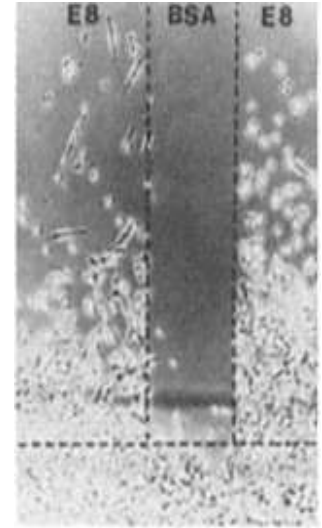
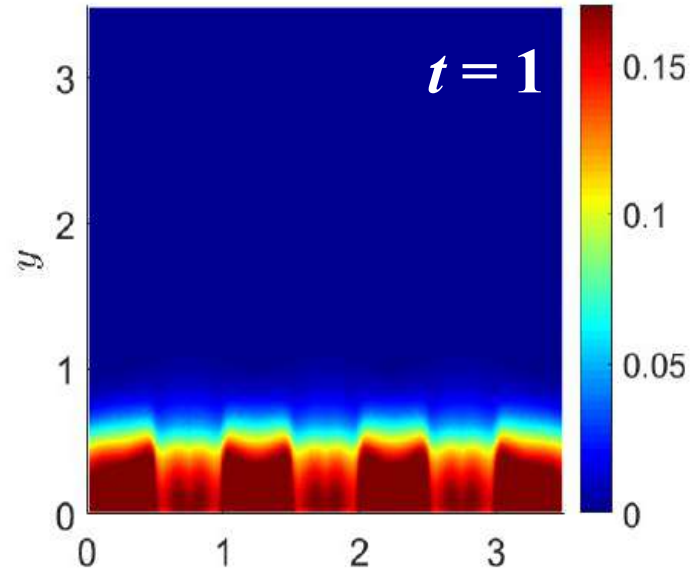
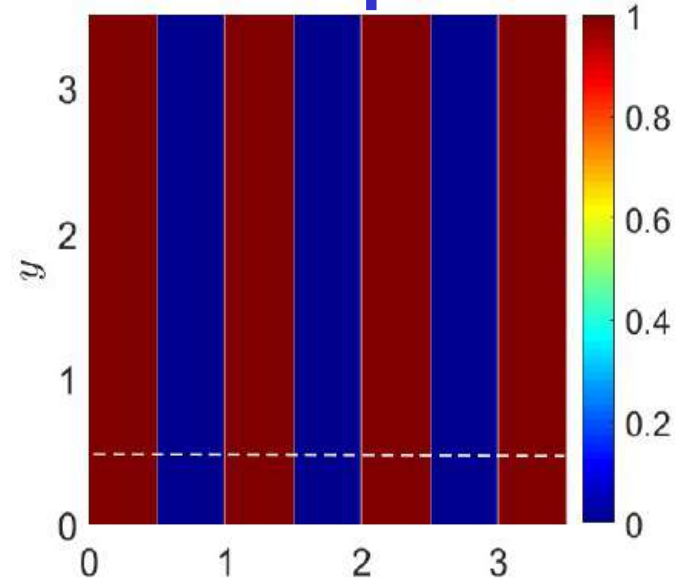
$$R_M^{max} = 2$$



Nonlocality allows
jumping beyond the pit



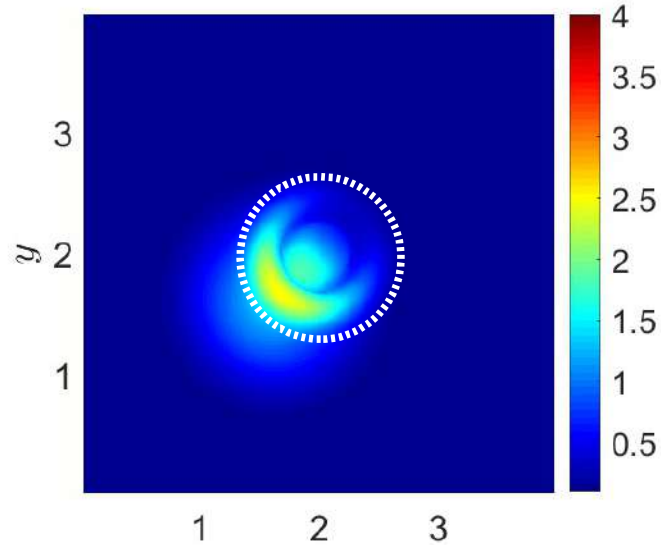
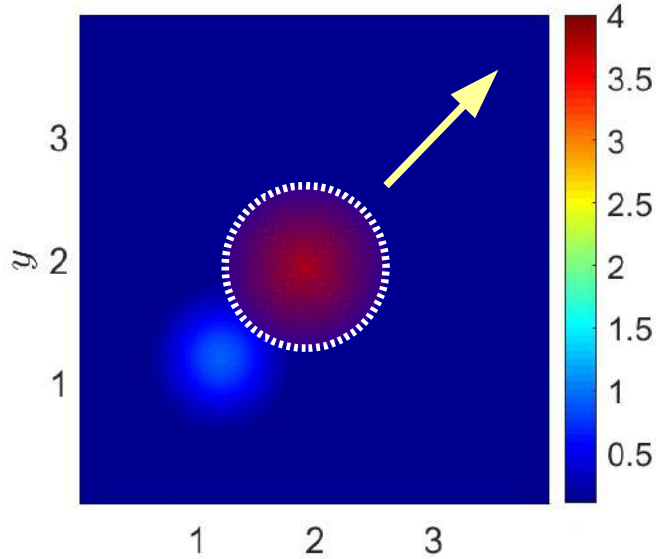
Stripes



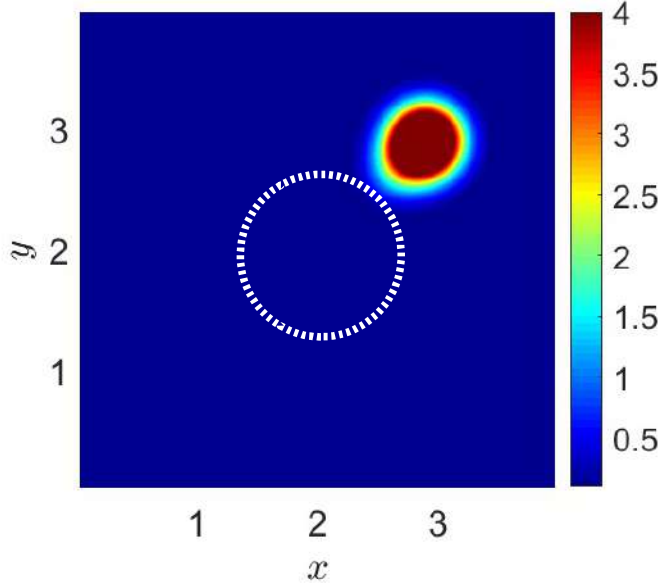
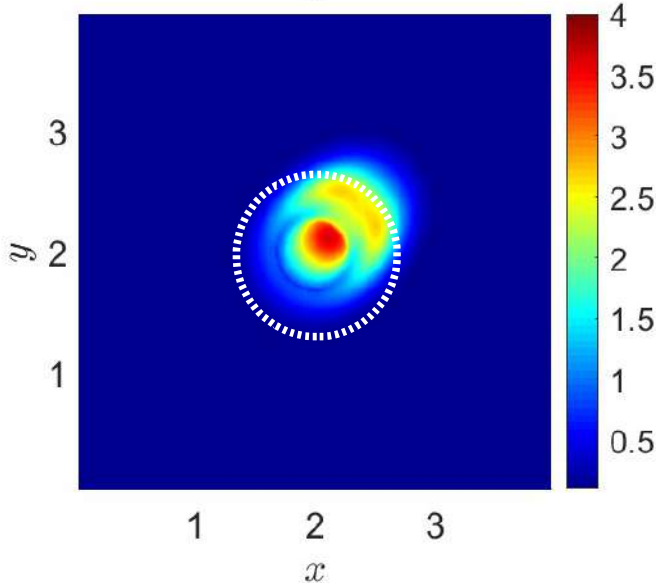
Nonlocality
allows
jumping on
the stripes



Double bias: Effect of independent cues

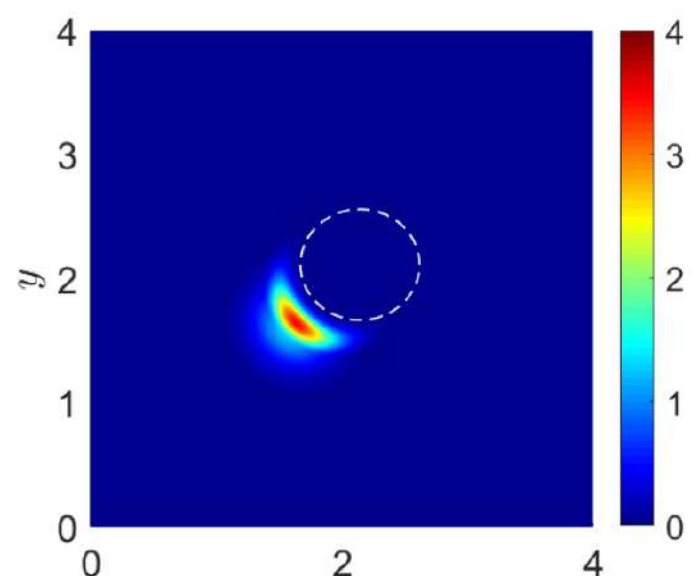
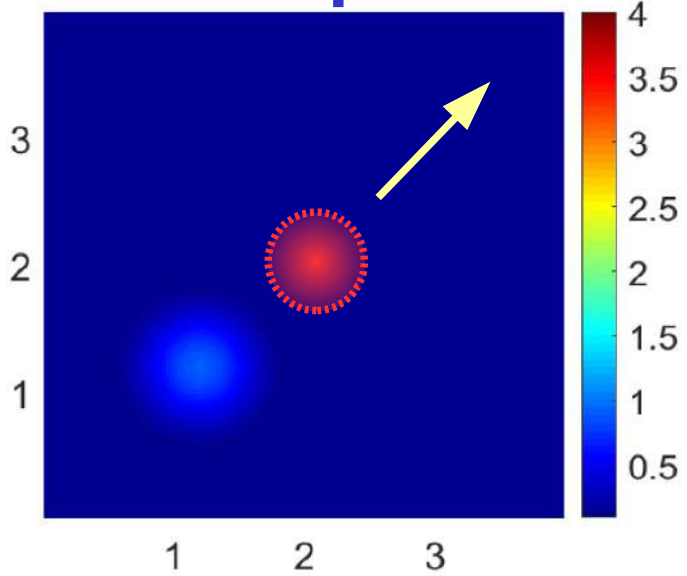


Denser ECM
in circle
(but still OK)

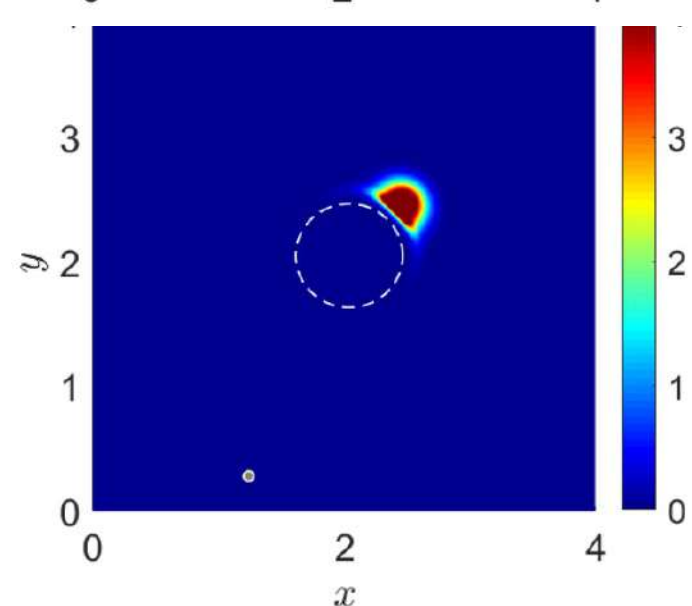
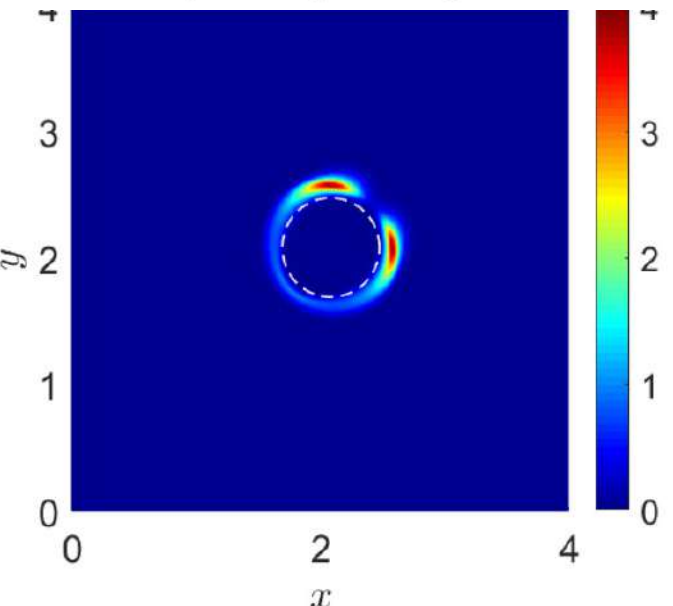




Double bias: Effect of independent cues

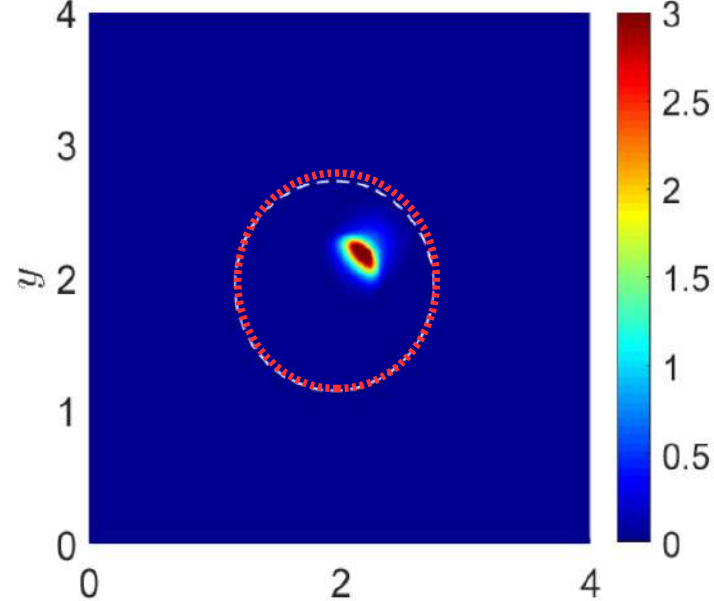
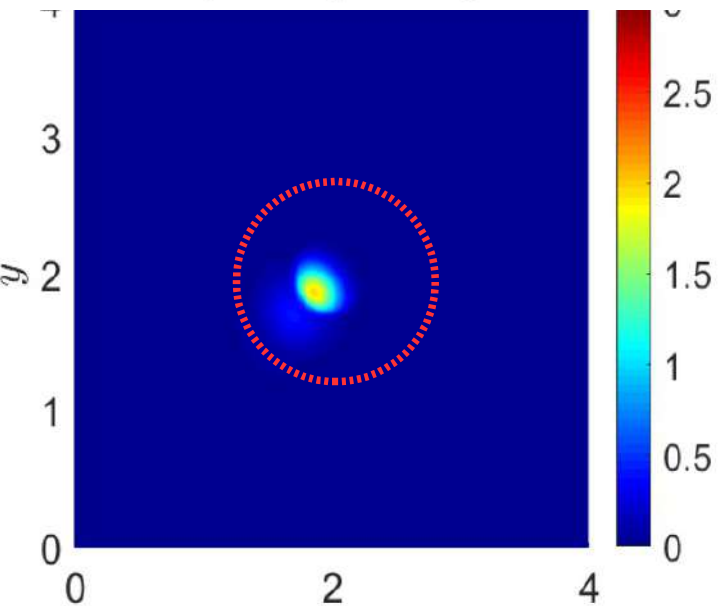
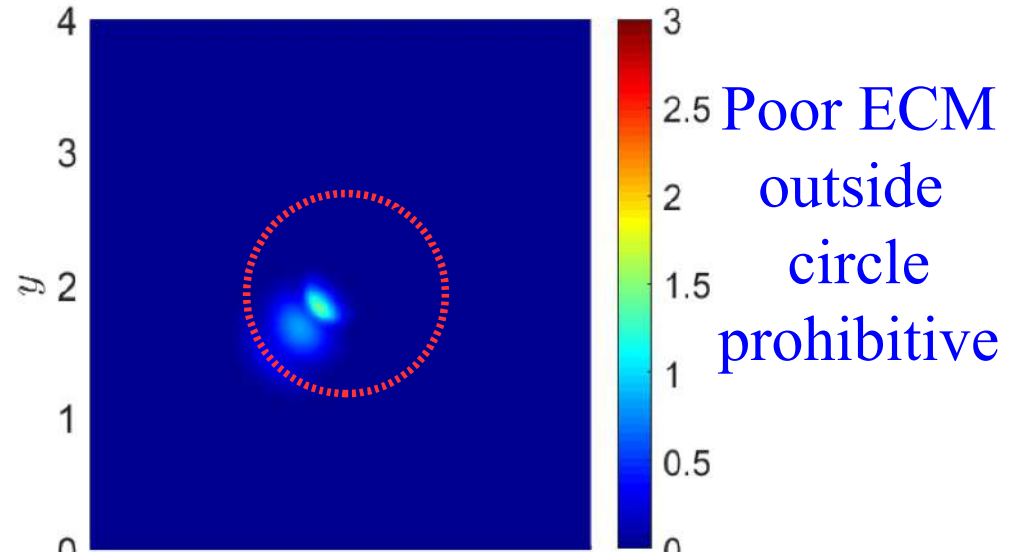
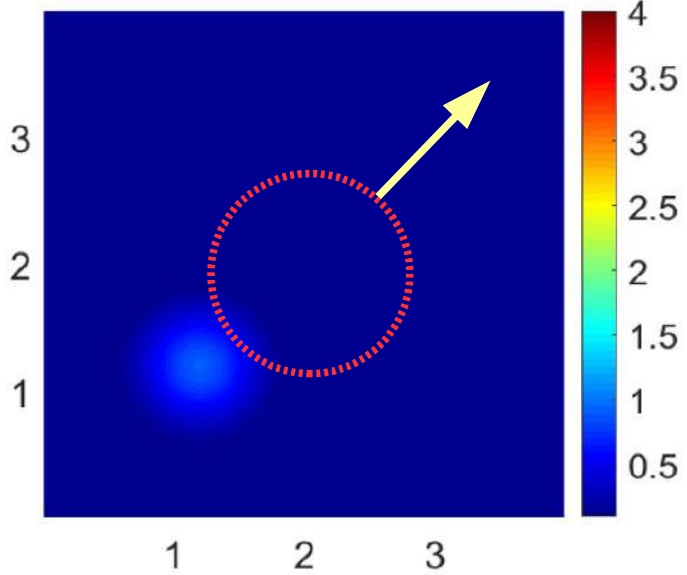


Denser ECM
in circle
(Too dense)





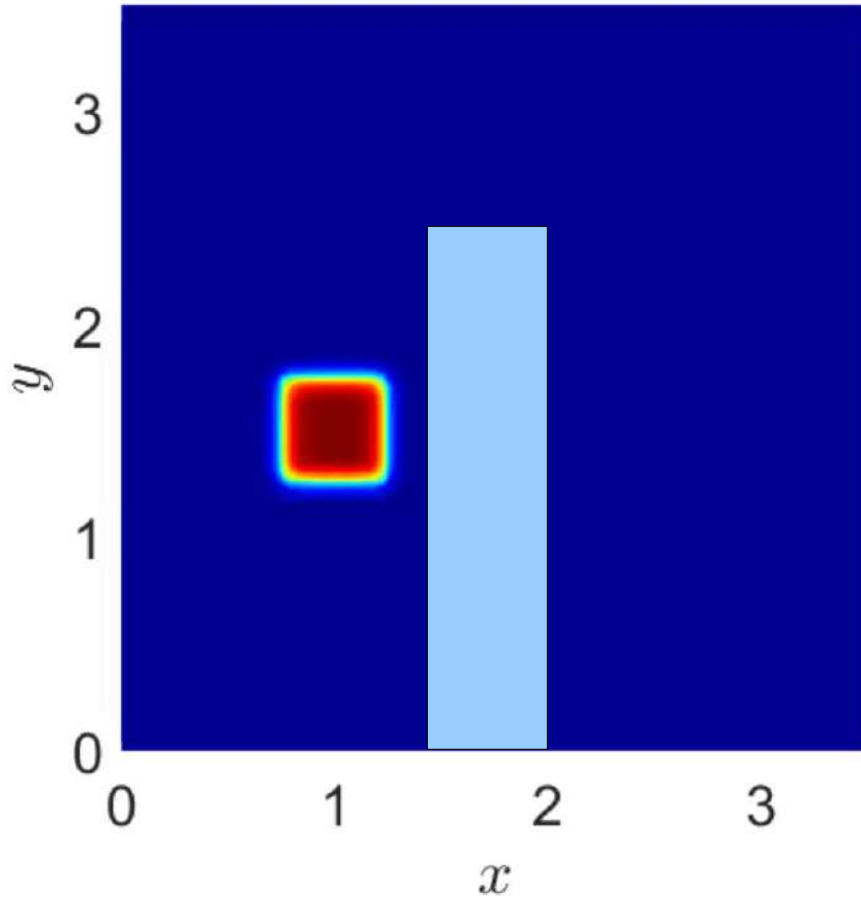
Double bias: Effect of independent cues



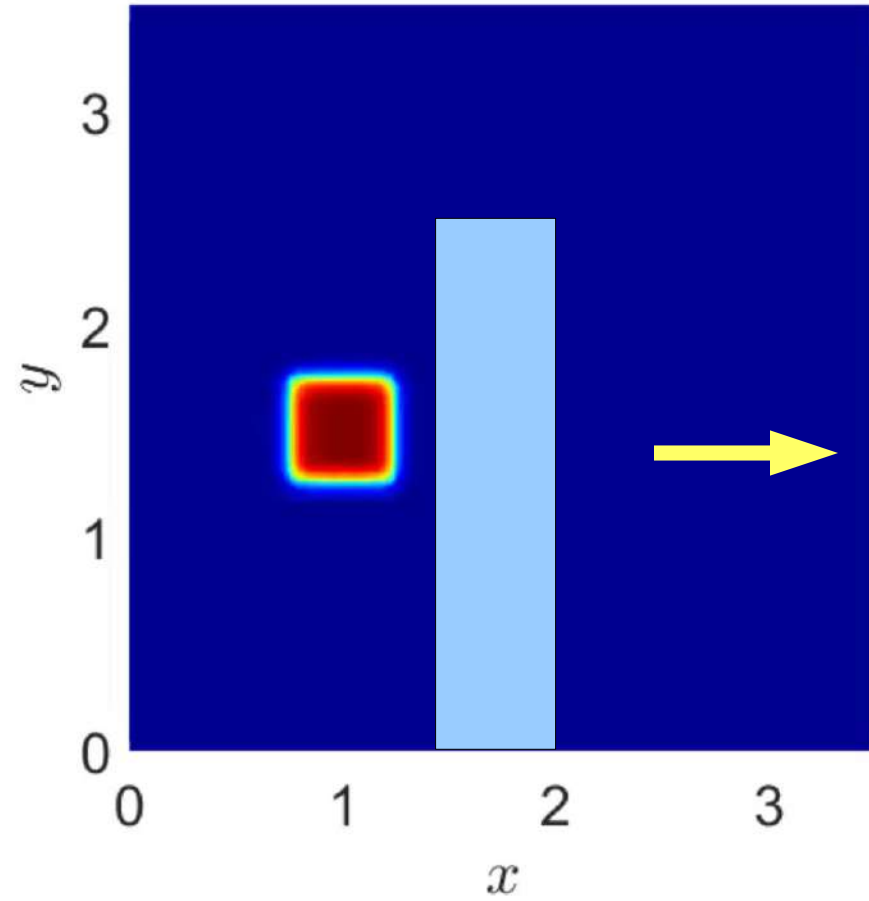


Double bias: Physical limits of migration

Without chemotaxis



With chemotaxis





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Arxiv, 10.48550/ARXIV.2207.01930

Thanks for your attention!